# REVIEW

**Open Access** 

# Targeting deubiquitinating enzymes in cancer stem cells



Hu Lei<sup>\*</sup>, Huizhuang Shan and Yingli Wu<sup>\*</sup>

# Abstract

Cancer stem cells (CSCs) are rare but accounted for tumor initiation, progression, metastasis, relapse and therapeutic resistance. Ubiquitination and deubiquitination of stemness-related proteins are essential for CSC maintenance and differentiation, even leading to execute various stem cell fate choices. Deubiquitinating enzymes (DUBs), specifically disassembling ubiquitin chains, are important to maintain the balance between ubiquitination and deubiquitination. In this review, we have focused on the DUBs regulation of stem cell fate determination. For example, we discuss deubiquitinase inhibition may lead stem cell transcription factors and CSCs-related protein degradation. Also, CSCs microenvironment is regulated by DUBs activity. Our review provides a new insight into DUBs activity by emphasizing their cellular role in regulating stem cell fate and illustrates the opportunities for the application of DUBs inhibitors in the CSC-targeted therapy.

Keywords: Cancer stem cells, Deubiquitinating enzymes, Cancer therapies, CSCs

# Background

The existence of cancer stem cells (CSCs) are considered to play a pivotal role in tumor recurrence, resistance and progression [1, 2]. There are three main aspects to effect CSCs maintenance and differentiation, including transcription factor network, CSC-related proteins and microenvironment [3, 4]. Conventional cancer therapy can't kill cancer stem cells, which will cause cancer relapse and drug resistance under certain conditions (Fig. 1).

Ubiquitination is a post-translational modification process that participates in the covalent conjugation of small, highly conserved 76 amino acid protein ubiquitin with the lysine residues of the substrate protein through the cascade of enzyme reactions, including E1-activating enzymes, E2-conjugating enzymes, and E3 ligases, resulting in protein final degradation, relocalization or activity change. On the contrary, DUB-mediated deubiquitination removes the ubiquitin labels to protect

\*Correspondence: hulei@shsmu.edu.cn; wuyingli@shsmu.edu.cn Hongqiao International Institute of Medicine, Shanghai Tongren Hospital/ Faculty of Basic Medicine, Chemical Biology Division of Shanghai Universities E-Institutes, Key Laboratory of Cell Differentiation and Apoptosis of the Chinese Ministry of Education, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China substrate proteins from above-mentioned changes caused by ubiquitination. It has been reported that the ubiquitination and deubiquitination of the key proteins in stem cells may determine the fate of cells (Fig. 2). Recently, DUBs have been demonstrated as promising targets for cancer therapy [5–7], their functions in cancer cell stemness remains elusive. For example, USP54 is overexpressed in colorectal cancer stem cells and promotes intestinal tumorigenesis [8]. USP28 confers stem-cell-like traits to breast cancer cells [9].

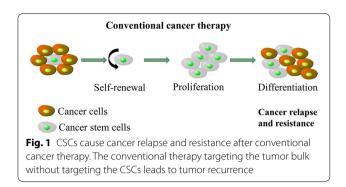
Finding deubiquitinates of transcription factors and key protein can provide better understand of the activation mechanism on CSCs, and further deubiquitination inhibitors can be used to eliminate CSCs in cancer radical treatment.

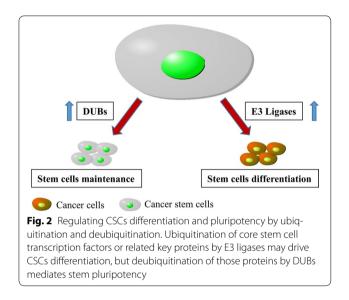
#### **DUBs and CSC-associated transcription factors**

Embryonic stem cells (ESCs) self-renewal and differentiation are known to be regulated by a network of transcription factors including Oct3/4, Sox2, c-Myc, Klf4 and Nanog [10, 11]. Cancer stem cells share significant similarity with normal stem cells in biological characteristics such as quiescence, self-renewal and differentiation [12, 13].



© The Author(s) 2017. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/ publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.





#### Sox2

Sox2 also regulates the differentiation and stemness in cancer stem cells [14]. USP22 is located directly on the Sox2 promoter and negatively regulates Sox2 transcription in ESCs [15]. In brain tumor cells, Usp9x was associated with Sox2 and played key roles in the growth of tumor cells, but the relationship between them was not clear [16]. Sox2 also regulated DUBs activity by binding to the promoter region at the transcriptional level, such as USP7, USP25, USP37, and USP44 [17].

#### c-Myc

c-Myc is a classical CSC-related marker, which can be stabilized by many DUBs. USP37 directly deubiquitinates and stabilizes c-Myc in lung cancer [18]. USP22 positively regulates c-Myc stability and tumorigenic activity in mammalian and breast cancer cells [19]. In a subset of human breast and lung cancers, USP36 interacts with and deubiquitinates c-Myc [20]. USP28 is required for c-Myc stability in human tumor cells, which binds to c-Myc through an interaction with FBW7alpha, an F-box protein that is part of an SCF-type ubiquitin ligase [21].

#### Nanog and ID proteins

Recent studies demonstrated that USP21 maintained the stemness of mouse embryonic stem cells via stabilization of Nanog by removing K48-linked ubiquitin chains [22]. Inhibitor of DNA binding (ID) proteins are transcriptional regulators that control the timing of cell fate determination and differentiation in stem and progenitor cells during normal development and adult life [23]. The small molecule inhibitor of USP1 promotes ID1 degradation and has cytotoxicity to leukemic cells [24]. USP1 deubiquitinated and stabilized ID1, ID2, and ID3 proteins to preserve a mesenchymal stem cell program in osteosarcoma [25].

Some pluripotent factors such as Oct3/4, Klf4 and Lin28 have not been found their DUBs, but all of them are affected by the 26S proteasome, suggesting a potential role of DUB for their stabilization in CSCs.

# **DUBs and CSC-related proteins**

Some CSC-related proteins also control the fate of CSC, such as SIRT1, P53, PTEN, LSD1, PRC and so on. SIRT1, a NAD<sup>+</sup>-dependent histone deacetylase, influences stem cell aging by controlling mitochondrial biogenesis and turnover which may be required for self-renewal [26, 27].

# SIRT1

SIRT1 inhibition represents a potential approach to target leukemia stem cells [28, 29]. USP22 interacts with and stabilizes SIRT1 by removing polyubiquitin chains conjugated onto SIRT1 in mouse embryonic development [30].

#### P53

P53, tumor suppresser, demonstrates a role for p53 deficiency in enhancing the formation of tumors arising from stem cells (embryonal carcinoma cells) [31, 32]. It is reported that USP10 deubiquitinates p53, reversing Mdm2-induced p53 nuclear export and degradation [33]. Ataxin-3, the machado–joseph disease deubiquitinase, interacts with p53 and functions as a novel p53 DUB [34]. USP7 deubiquitinates both p53 and MDM2, one of the ubiquitin ligases that ubiquitylates p53, thereby stabilizing both proteins [35, 36]. OTUD1, OTUD5 and USP11 directly deubiquitinating p53 and functional proteins were required for p53 stabilization [37–39].

## PTEN

PTEN loss leads to the development of cancer stem cells, with the capacity of self-renewal and multi-lineage differentiation [40–43]. ATXN3 acts primarily by repressing PTEN transcription, without altering PTEN protein stability [44]. However, USP18 overexpression could stabilize PTEN protein, and USP18 repression decreases mainly cytoplasmic PTEN [45]. PTEN subcellular compartmentalization can be regulated by USP7 [46, 47].

#### PRC

The dysfunction of polycomb repressive complex (PRC) is closely related to cancer stemness [48, 49]. PRC1 represses transcription is only in part dependent on its ubiquitination activity, and Fbxl10 is reported to recruit PRC1 to CpG islands and regulate H2A ubiquitylation [50, 51]. Polycomb gene silencing may require H2A ubiquitination by PRC1 and H2A deubiquitination by Polycomb repressive deubiquitinase (PR-DUB). In some cancer types, PRC1 can be deubiquitinated by USP7, USP11 and USP26 [52, 53]. PRC2-mediated histone methylation plays an important role in aberrant cancer gene silencing and is a potential target for cancer therapy. The PRC2 proteins EZH2 is frequently overexpressed in mesothelioma with BAP1 mutation [54]. The deubiquitination enzymes of PRC need to be further explored in the future.

#### LSD

Lysine-specific demethylase 1 (LSD1), the first identified histone demethylase, maintains cell stemness during cancer progression [55, 56]. USP7 and USP28 inhibited LSD1 ubiquitination and stabilized LSD1 protein level [9, 57].

Taken together, CSC-related proteins degradation or activity inhibition by targeting DUBs is effective for eliminating cancer stem cells.

# **DUBs and CSC microenvironment**

The microenvironment of CSC has also been reported to play essential roles in maintenance of cancer stemness. Tumor specific microenvironments comprise stromal cells, immune cells, networks of cytokines and growth factors, hypoxic regions, and the extracellular matrix (ECM). We summarize the role of CSC microenvironment from two aspects: hypoxia and inflammation [58–60].

#### Hypoxia

Hypoxia is considered to be a major feature of the tumor microenvironment and is a potential contributor to the CSC phenotype. Hypoxia-inducible factor (HIF) transcription factors (HIF-1 $\alpha$  and HIF-2 $\alpha$ ) are key mediators in cancer hypoxia response and help maintain multiple CSC population [61, 62]. In the presence of oxygen, VHL tumor suppressor protein interacts with HIF proteins and this interaction results in the ubiquitination and degradation of HIF proteins, maintaining low levels of these transcription factors [63]. However, HIF proteins stabilization can be regulated by DUBs, such as USP8, USP19 and USP28 [64–66]. In addition, USP52 is a key component of P-bodies required to prevent HIF1 $\alpha$  mRNA degradation [67].

#### Inflammation

The inflammatory cytokines modify the cancer microenvironment, CSCs secretion factors attract the necessary cells into their areas, enabling them better survive and escape chemotherapy [68]. Transforming growth factor  $\beta$  (TGF $\beta$ ) has the ability to regulate immune cell populations in inhibiting and promoting tumor formation and progression active [69]. Cancer cells exposed to IL-6 are malignant, such as enhanced invasive ability and drug resistance [70, 71]. IL-8 promotes angiogenic activity through the activation of VEGFR2 [78]. USP21 binds to the promoter region of IL-8 and mediates transcriptional initiation in stem-cell like property of human renal cell carcinoma [79]. Also, IL-6 and G-CSF levels have been elevated in lung CSCs [80]. Most inflammatory cytokines

Table 1 The effect of deubiquitinating enzymes in the reg-ulation of target proteins

Proteins	Deubiquitinating enzymes	Effect	References
Sox2	USP22	Transcription	[15]
	USP9X	Unclear	[16]
c-myc	USP37	Protein stabilization	[18]
	USP22		[19]
	USP36		[20]
	USP28		[21]
Nanog	USP21	Protein stabilization	[22]
ID proteins	USP1	Protein stabilization	[24, 25]
SIRT1	USP22	Protein stabilization	[30]
p53	USP10	Protein stabilization	[33]
	Ataxin-3		[34]
	USP7		[35, 36]
	OTUD1		[37]
	OTUD5		[38]
	USP11		[39]
PTEN	ATXN3	Transcription	[44]
	USP18	Protein stabilization	[45]
	USP7	Location	[46, 47]
PRC1	USP7	Protein stabilization	[52]
	USP11		[53]
	USP26		[77]
PRC2	BAP1	Unclear	[54]
LSD1	USP7	Protein stabilization	[57]
	USP28		[9]
HIF-1a	USP8	Protein stabilization	[66]
	USP19		[65]
	USP28		[64]
	USP52	mRNA degradation	[67]
IL-8	USP21	Transcription	[79]
TRAF6	USP4	Activity	[81]
	A20	Protein stabilization mRNA degradation Transcription	[82]

Inhibitors	Targeted DUBs	CSC type	References
Pimozide	USP1	Osteosarcoma, glioblastoma	[25, 83]
ML323	USP1		
P5091	USP7, USP47	Neural, glioblastoma, multiple myeloma	[57, 85–87]
P22077	USP7, USP47		
WP1130	USP9x, USP5, UCHL1, USP14, UCH37	Liver, breast cancer	[72, 73]
IU1	USP14	Gastric, multiple myeloma	[74, 75]
b-AP15	USP14, UCHL5		
VLX1570	USP14		
LDN-57444	UCHL1, UCHL3	Prostate	[76]
TCID	UCHL3, UCHL5	Multiple myeloma	[84]

Table 2 DUB inhibitors for preclinical application in CSC-targeted therapy

are produced by many kinds of signal pathways and the deubiquitination of key proteins in the pathway can block inflammatory cytokines release. For example, TRAF6, a key regulator in toll-like receptor pathway and NF- $\kappa$ B pathway, can be regulated by USP4 and A20 [81, 82].

# Conclusions

CSCs are difficult to eliminate by conventional treatment. mainly due to disorders of signal transduction and epigenetics. The control of ubiquitination and deubiquitination of CSC-related proteins determine the difference in CSCs and the maintenance of pluripotency. DUBs can protect the stemness of the CSC, thereby maintaining its activity and further forming a vicious circle. Therefore, DUBs are very important in the CSC specific treatment. We summarized the effect of deubiquitinating enzymes in the regulation of target proteins in Table 1. The successful inhibition of CSC maintenance and radiation resistance by USP1 specific inhibitor (pimozide) has been provided the basis for further clinical trials [83]. It means that DUB inhibitors may boost more advantages in CSC-specific therapy than other anti-cancer drugs such as proteasome inhibitors. For example, b-AP15, a selective DUB inhibitor, can overcome bortezomib resistance in multiple myeloma [84]. More relevant basic research should be carried out to determine the DUBs related to the CSCs and to identify the mechanisms between them. Currently commercialized DUB inhibitors are summarized in Table 2, showing significant pharmacological effects on cancer cells or cancer stem cells. In general, strategies involving the use of DUB inhibitors to target combination therapy of cancer stem cells and differentiated cancer cells can provide better outcomes for radical cancer treatment.

#### Abbreviations

CSCs: cancer stem cells; DUBs: deubiquitinating enzymes; ESCs: embryonic stem cells; ID: inhibitor of DNA binding; PRC: polycomb repressive complex;

LSD1: lysine-specific demethylase 1; ECM: extracellular matrix; HIF: hypoxia-inducible factor; TGF $\beta$ : transforming growth factor  $\beta$ .

#### Authors' contributions

HL collected materials and wrote the review. HZS collected materials. YLW modified and corrected the review. All authors read and approved the final manuscript.

#### Acknowledgements

Not applicable.

#### **Competing interests**

The authors declare that they have no competing interests.

#### Availability of data and materials

Not applicable.

**Consent for publication** Not applicable.

#### Ethics approval and consent to participate

Not applicable.

#### Funding

National Natural Science Foundation of China (81570118; 81700475).

#### **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 29 August 2017 Accepted: 26 October 2017 Published online: 03 November 2017

#### References

- Krause M, Dubrovska A, Linge A, Baumann M. Cancer stem cells: radioresistance, prediction of radiotherapy outcome and specific targets for combined treatments. Adv Drug Deliv Rev. 2017;109:63–73.
- Clarke MF, Dick JE, Dirks PB, Eaves CJ, Jamieson CH, Jones DL, Visvader J, Weissman IL, Wahl GM. Cancer stem cells–perspectives on current status and future directions: AACR workshop on cancer stem cells. Cancer Res. 2006;66(19):9339–44.
- Qiu GZ, Sun W, Jin MZ, Lin J, Lu PG, Jin WL. The bad seed gardener: deubiquitinases in the cancer stem–cell signaling network and therapeutic resistance. Pharmacol Ther. 2017;172:127–38.
- Suresh B, Lee J, Kim KS, Ramakrishna S. The importance of ubiquitination and deubiquitination in cellular reprogramming. Stem Cells Int. 2016;2016:6705927.

- Fraile JM, Manchado E, Lujambio A, Quesada V, Campos-Iglesias D, Webb TR, Lowe SW, Lopez-Otin C, Freije JM. USP39 deubiquitinase is essential for KRAS oncogene-driven cancer. J Biol Chem. 2017;292(10):4164–75.
- Zhao C, Chen X, Zang D, Lan X, Liao S, Yang C, Zhang P, Wu J, Li X, Liu N, et al. A novel nickel complex works as a proteasomal deubiquitinase inhibitor for cancer therapy. Oncogene. 2016;35(45):5916–27.
- Suresh B, Lee J, Kim H, Ramakrishna S. Regulation of pluripotency and differentiation by deubiquitinating enzymes. Cell Death Differ. 2016;23(8):1257–64.
- Fraile JM, Campos-Iglesias D, Rodriguez F, Espanol Y, Freije JM. The deubiquitinase USP54 is overexpressed in colorectal cancer stem cells and promotes intestinal tumorigenesis. Oncotarget. 2016;7(46):74427–34.
- Wu Y, Wang Y, Yang XH, Kang T, Zhao Y, Wang C, Evers BM, Zhou BP. The deubiquitinase USP28 stabilizes LSD1 and confers stem-cell-like traits to breast cancer cells. Cell Rep. 2013;5(1):224–36.
- Cai N, Li M, Qu J, Liu GH, Izpisua Belmonte JC. Post-translational modulation of pluripotency. J Mol Cell Biol. 2012;4(4):262–5.
- Ramakrishna S, Kim KS, Baek KH. Posttranslational modifications of defined embryonic reprogramming transcription factors. Cell Reprogr. 2014;16(2):108–20.
- 12. Zhao J. Cancer stem cells and chemoresistance: the smartest survives the raid. Pharmacol Ther. 2016;160:145–58.
- Bharti R, Dey G, Mandal M. Cancer development, chemoresistance, epithelial to mesenchymal transition and stem cells: a snapshot of IL-6 mediated involvement. Cancer Lett. 2016;375(1):51–61.
- Liu K, Lin B, Zhao M, Yang X, Chen M, Gao A, Liu F, Que J, Lan X. The multiple roles for Sox2 in stem cell maintenance and tumorigenesis. Cell Sign. 2013;25(5):1264–71.
- Sussman RT, Stanek TJ, Esteso P, Gearhart JD, Knudsen KE, McMahon SB. The epigenetic modifier ubiquitin-specific protease 22 (USP22) regulates embryonic stem cell differentiation via transcriptional repression of sexdetermining region Y-box 2 (SOX2). J Biol Chem. 2013;288(33):24234–46.
- Cox JL, Wilder PJ, Gilmore JM, Wuebben EL, Washburn MP, Rizzino A. The SOX2-interactome in brain cancer cells identifies the requirement of MSI2 and USP9X for the growth of brain tumor cells. PLoS ONE. 2013;8(5):e62857.
- Boyer LA, Lee TI, Cole MF, Johnstone SE, Levine SS, Zucker JP, Guenther MG, Kumar RM, Murray HL, Jenner RG, et al. Core transcriptional regulatory circuitry in human embryonic stem cells. Cell. 2005;122(6):947–56.
- Pan J, Deng Q, Jiang C, Wang X, Niu T, Li H, Chen T, Jin J, Pan W, Cai X, et al. USP37 directly deubiquitinates and stabilizes c-Myc in lung cancer. Oncogene. 2015;34(30):3957–67.
- Kim D, Hong A, Park HI, Shin WH, Yoo L, Jeon SJ, Chung KC. Deubiquitinating enzyme USP22 positively regulates c-Myc stability and tumorigenic activity in mammalian and breast cancer cells. J Cell Physiol. 2017;232(12):3664–76.
- Sun XX, He X, Yin L, Komada M, Sears RC, Dai MS. The nucleolar ubiquitinspecific protease USP36 deubiquitinates and stabilizes c-Myc. Proc Natl Acad Sci USA. 2015;112(12):3734–9.
- Popov N, Wanzel M, Madiredjo M, Zhang D, Beijersbergen R, Bernards R, Moll R, Elledge SJ, Eilers M. The ubiquitin-specific protease USP28 is required for MYC stability. Nat Cell Biol. 2007;9(7):765–74.
- Jin J, Liu J, Chen C, Liu Z, Jiang C, Chu H, Pan W, Wang X, Zhang L, Li B, et al. The deubiquitinase USP21 maintains the stemness of mouse embryonic stem cells via stabilization of Nanog. Nat Commun. 2016;7:13594.
- Lasorella A, Benezra R, lavarone A. The ID proteins: master regulators of cancer stem cells and tumour aggressiveness. Nat Rev Cancer. 2014;14(2):77–91.
- Mistry H, Hsieh G, Buhrlage SJ, Huang M, Park E, Cuny GD, Galinsky I, Stone RM, Gray NS, D'Andrea AD, et al. Small-molecule inhibitors of USP1 target ID1 degradation in leukemic cells. Mol Cancer Ther. 2013;12(12):2651–62.
- Williams SA, Maecker HL, French DM, Liu J, Gregg A, Silverstein LB, Cao TC, Carano RA, Dixit VM. USP1 deubiquitinates ID proteins to preserve a mesenchymal stem cell program in osteosarcoma. Cell. 2011;146(6):918–30.
- 26. Mantel C, Broxmeyer HE. Sirtuin 1, stem cells, aging, and stem cell aging. Curr Opin Hematol. 2008;15(4):326–31.
- Zhou L, Chen X, Liu T, Zhu C, Si M, Jargstorf J, Li M, Pan G, Gong Y, Luo ZP et al. SIRT1-dependent anti-senescence effects of cell-deposited matrix on human umbilical cord mesenchymal stem cells. J Tissue Eng Regen Med. 2017. doi:10.1002/term.2422.

- Li L, Osdal T, Ho Y, Chun S, McDonald T, Agarwal P, Lin A, Chu S, Qi J, Li L, et al. SIRT1 activation by a c-MYC oncogenic network promotes the maintenance and drug resistance of human FLT3-ITD acute myeloid leukemia stem cells. Cell Stem Cell. 2014;15(4):431–46.
- Jin Y, Cao Q, Chen C, Du X, Jin B, Pan J. Tenovin-6-mediated inhibition of SIRT1/2 induces apoptosis in acute lymphoblastic leukemia (ALL) cells and eliminates ALL stem/progenitor cells. BMC Cancer. 2015;15:226.
- Lin Z, Yang H, Kong Q, Li J, Lee SM, Gao B, Dong H, Wei J, Song J, Zhang DD, et al. USP22 antagonizes p53 transcriptional activation by deubiquitinating Sirt1 to suppress cell apoptosis and is required for mouse embryonic development. Mol Cell. 2012;46(4):484–94.
- Puzio-Kuter AM, Levine AJ. Stem cell biology meets p53. Nat Biotechnol. 2009;27(10):914–5.
- 32. Aloni-Grinstein R, Shetzer Y, Kaufman T, Rotter V. p53: the barrier to cancer stem cell formation. FEBS Lett. 2014;588(16):2580–9.
- Yuan J, Luo K, Zhang L, Cheville JC, Lou Z. USP10 regulates p53 localization and stability by deubiquitinating p53. Cell. 2010;140(3):384–96.
- Liu H, Li X, Ning G, Zhu S, Ma X, Liu X, Liu C, Huang M, Schmitt I, Wullner U, et al. The Machado-Joseph disease deubiquitinase Ataxin-3 regulates the stability and apoptotic function of p53. PLoS Biol. 2016;14(11):e2000733.
- Brooks CL, Li M, Hu M, Shi Y, Gu W. The p53–Mdm2–HAUSP complex is involved in p53 stabilization by HAUSP. Oncogene. 2007;26(51):7262–6.
- 36. Brooks CL, Gu W. p53 regulation by ubiquitin. FEBS Lett. 2011;585(18):2803–9.
- Piao S, Pei HZ, Huang B, Baek SH. Ovarian tumor domain-containing protein 1 deubiquitinates and stabilizes p53. Cell Sign. 2017;33:22–9.
- 38. Luo J, Lu Z, Lu X, Chen L, Cao J, Zhang S, Ling Y, Zhou X. OTUD5 regulates p53 stability by deubiquitinating p53. PLoS ONE. 2013;8(10):e77682.
- Ke JY, Dai CJ, Wu WL, Gao JH, Xia AJ, Liu GP, Lv KS, Wu CL. USP11 regulates p53 stability by deubiquitinating p53. J Zhejiang Univ Sci B. 2014;15(12):1032–8.
- Schubbert S, Jiao J, Ruscetti M, Nakashima J, Wu S, Lei H, Xu Q, Yi W, Zhu H, Wu H. Methods for PTEN in stem cells and cancer stem cells. Methods Mol Biol. 2016;1388:233–85.
- Duan S, Yuan G, Liu X, Ren R, Li J, Zhang W, Wu J, Xu X, Fu L, Li Y, et al. PTEN deficiency reprogrammes human neural stem cells towards a glioblastoma stem cell-like phenotype. Nat Commun. 2015;6:10068.
- Liao J, Marumoto T, Yamaguchi S, Okano S, Takeda N, Sakamoto C, Kawano H, Nii T, Miyamato S, Nagai Y, et al. Inhibition of PTEN tumor suppressor promotes the generation of induced pluripotent stem cells. Mol Ther. 2013;21(6):1242–50.
- 43. Hill R, Wu H. PTEN, stem cells, and cancer stem cells. J Biol Chem. 2009;284(18):11755–9.
- Sacco JJ, Yau TY, Darling S, Patel V, Liu H, Urbe S, Clague MJ, Coulson JM. The deubiquitylase Ataxin-3 restricts PTEN transcription in lung cancer cells. Oncogene. 2014;33(33):4265–72.
- Mustachio LM, Kawakami M, Lu Y, Rodriguez-Canales J, Mino B, Behrens C, Wistuba I, Bota-Rabassedas N, Yu J, Lee JJ, et al. The ISG15-specific protease USP18 regulates stability of PTEN. Oncotarget. 2017;8(1):3–14.
- Song MS, Salmena L, Carracedo A, Egia A, Lo-Coco F, Teruya-Feldstein J, Pandolfi PP. The deubiquitinylation and localization of PTEN are regulated by a HAUSP-PML network. Nature. 2008;455(7214):813–7.
- Morotti A, Panuzzo C, Crivellaro S, Pergolizzi B, Familiari U, Berger AH, Saglio G, Pandolfi PP. BCR-ABL disrupts PTEN nuclear-cytoplasmic shuttling through phosphorylation-dependent activation of HAUSP. Leukemia. 2014;28(6):1326–33.
- Gao X, Jin W. The emerging role of tumor-suppressive microRNA-218 in targeting glioblastoma stemness. Cancer Lett. 2014;353(1):25–31.
- Suva ML, Riggi N, Janiszewska M, Radovanovic I, Provero P, Stehle JC, Baumer K, Le Bitoux MA, Marino D, Cironi L, et al. EZH2 is essential for glioblastoma cancer stem cell maintenance. Cancer Res. 2009;69(24):9211–8.
- Laugesen A, Hojfeldt JW, Helin K. Role of the polycomb repressive complex 2 (PRC2) in transcriptional regulation and cancer. Cold Spring Harb Perspect Med. 2016;6(9):a026575.
- Wu X, Johansen JV, Helin K. Fbxl10/Kdm2b recruits polycomb repressive complex 1 to CpG islands and regulates H2A ubiquitylation. Mol Cell. 2013;49(6):1134–46.
- Maertens GN, El Messaoudi-Aubert S, Elderkin S, Hiom K, Peters G. Ubiquitin-specific proteases 7 and 11 modulate Polycomb regulation of the INK4a tumour suppressor. EMBO J. 2010;29(15):2553–65.

- 53. Lecona E, Narendra V, Reinberg D. USP7 cooperates with SCML2 to regulate the activity of PRC1. Mol Cell Biol. 2015;35(7):1157–68.
- Kemp CD, Rao M, Xi S, Inchauste S, Mani H, Fetsch P, Filie A, Zhang M, Hong JA, Walker RL, et al. Polycomb repressor complex-2 is a novel target for mesothelioma therapy. Clin Cancer Res. 2012;18(1):77–90.
- Amente S, Lania L, Majello B. The histone LSD1 demethylase in stemness and cancer transcription programs. Biochim Biophys Acta. 2013;1829(10):981–6.
- Hino S, Kohrogi K, Nakao M. Histone demethylase LSD1 controls the phenotypic plasticity of cancer cells. Cancer Sci. 2016;107(9):1187–92.
- Yi L, Cui Y, Xu Q, Jiang Y. Stabilization of LSD1 by deubiquitinating enzyme USP7 promotes glioblastoma cell tumorigenesis and metastasis through suppression of the p53 signaling pathway. Oncol Rep. 2016;36(5):2935–45.
- Carnero A, Lleonart M. The hypoxic microenvironment: a determinant of cancer stem cell evolution. BioEssays. 2016;38(Suppl 1):S65–74.
- 59. Kise K, Kinugasa-Katayama Y, Takakura N. Tumor microenvironment for cancer stem cells. Adv Drug Deliv Rev. 2016;99(Pt B):197–205.
- Lau EY, Ho NP, Lee TK. Cancer stem cells and their microenvironment: biology and therapeutic implications. Stem Cells Int. 2017;2017:3714190.
- van den Beucken T, Koch E, Chu K, Rupaimoole R, Prickaerts P, Adriaens M, Voncken JW, Harris AL, Buffa FM, Haider S, et al. Hypoxia promotes stem cell phenotypes and poor prognosis through epigenetic regulation of DICER. Nat Commun. 2014;5:5203.
- Zhao M, Zhang Y, Zhang H, Wang S, Zhang M, Chen X, Wang H, Zeng G, Chen X, Liu G, et al. Hypoxia-induced cell stemness leads to drug resistance and poor prognosis in lung adenocarcinoma. Lung Cancer. 2015;87(2):98–106.
- Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, Wykoff CC, Pugh CW, Maher ER, Ratcliffe PJ. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. Nature. 1999;399(6733):271–5.
- Flugel D, Gorlach A, Kietzmann T. GSK-3beta regulates cell growth, migration, and angiogenesis via Fbw7 and USP28-dependent degradation of HIF-1alpha. Blood. 2012;119(5):1292–301.
- Altun M, Zhao B, Velasco K, Liu H, Hassink G, Paschke J, Pereira T, Lindsten K. Ubiquitin-specific protease 19 (USP19) regulates hypoxia-inducible factor 1alpha (HIF-1alpha) during hypoxia. J Biol Chem. 2012;287(3):1962–9.
- Troilo A, Alexander I, Muehl S, Jaramillo D, Knobeloch KP, Krek W. HIF1alpha deubiquitination by USP8 is essential for ciliogenesis in normoxia. EMBO Rep. 2014;15(1):77–85.
- Bett JS, Ibrahim AF, Garg AK, Kelly V, Pedrioli P, Rocha S, Hay RT. The P-body component USP52/PAN2 is a novel regulator of HIF1A mRNA stability. Biochem J. 2013;451(2):185–94.
- Shigdar S, Li Y, Bhattacharya S, O'Connor M, Pu C, Lin J, Wang T, Xiang D, Kong L, Wei MQ, et al. Inflammation and cancer stem cells. Cancer Lett. 2014;345(2):271–8.
- 69. Bierie B, Moses HL. Transforming growth factor beta (TGF-beta) and inflammation in cancer. Cytokine Growth Factor Rev. 2010;21(1):49–59.
- 70. Dethlefsen C, Hojfeldt G, Hojman P. The role of intratumoral and systemic IL-6 in breast cancer. Breast Cancer Res Treat. 2013;138(3):657–64.
- Hollier BG, Evans K, Mani SA. The epithelial-to-mesenchymal transition and cancer stem cells: a coalition against cancer therapies. J Mammary Gland Biol Neoplasia. 2009;14(1):29–43.
- Shen G, Lin Y, Yang X, Zhang J, Xu Z, Jia H. MicroRNA-26b inhibits epithelial-mesenchymal transition in hepatocellular carcinoma by targeting USP9X. BMC Cancer. 2014;14:393.
- Fu P, Du F, Liu Y, Yao M, Zhang S, Zheng X, Zheng S. WP1130 increases cisplatin sensitivity through inhibition of usp9x in estrogen receptornegative breast cancer cells. Am J Transl Res. 2017;9(4):1783–91.

- 74. Zhu Y, Zhang Y, Sui Z, Zhang Y, Liu M, Tang H. USP14 de-ubiquitinates vimentin and miR-320a modulates USP14 and vimentin to contribute to malignancy in gastric cancer cells. Oncotarget. 2016;8(30):48725-36.
- Wang X, Mazurkiewicz M, Hillert EK, Olofsson MH, Pierrou S, Hillertz P, Gullbo J, Selvaraju K, Paulus A, Akhtar S, et al. Corrigendum: the proteasome deubiquitinase inhibitor VLX1570 shows selectivity for ubiquitinspecific protease-14 and induces apoptosis of multiple myeloma cells. Sci Rep. 2016;6:30667.
- Song HM, Lee JE, Kim JH. Ubiquitin C-terminal hydrolase-L3 regulates EMT process and cancer metastasis in prostate cell lines. Biochem Biophys Res Commun. 2014;452(3):722–7.
- Ning B, Zhao W, Qian C, Liu P, Li Q, Li W, Wang RF. USP26 functions as a negative regulator of cellular reprogramming by stabilising PRC1 complex components. Nat Commun. 2017;8(1):349.
- Martin D, Galisteo R, Gutkind JS. CXCL8/IL8 stimulates vascular endothelial growth factor (VEGF) expression and the autocrine activation of VEGFR2 in endothelial cells by activating NFkappaB through the CBM (Carma3/Bcl10/Malt1) complex. J Biol Chem. 2009;284(10):6038–42.
- Peng L, Hu Y, Chen D, Jiao S, Sun S. Ubiquitin specific peptidase 21 regulates interleukin-8 expression, stem-cell like property of human renal cell carcinoma. Oncotarget. 2016;7(27):42007–16.
- Levina V, Marrangoni AM, DeMarco R, Gorelik E, Lokshin AE. Drug-selected human lung cancer stem cells: cytokine network, tumorigenic and metastatic properties. PLoS One. 2008;3(8):e3077.
- Xiao N, Li H, Luo J, Wang R, Chen H, Chen J, Wang P. Ubiquitin-specific protease 4 (USP4) targets TRAF2 and TRAF6 for deubiquitination and inhibits TNFalpha-induced cancer cell migration. Biochem J. 2012;441(3):979–86.
- Sun SC. Deubiquitylation and regulation of the immune response. Nat Rev Immunol. 2008;8(7):501–11.
- Lee JK, Chang N, Yoon Y, Yang H, Cho H, Kim E, Shin Y, Kang W, Oh YT, Mun GI, et al. USP1 targeting impedes GBM growth by inhibiting stem cell maintenance and radioresistance. Neuro-oncology. 2016;18(1):37–47.
- Tian Z, D'Arcy P, Wang X, Ray A, Tai YT, Hu Y, Carrasco RD, Richardson P, Linder S, Chauhan D, et al. A novel small molecule inhibitor of deubiquitylating enzyme USP14 and UCHL5 induces apoptosis in multiple myeloma and overcomes bortezomib resistance. Blood. 2014;123(5):706–16.
- Huang Z, Wu Q, Guryanova OA, Cheng L, Shou W, Rich JN, Bao S. Deubiquitylase HAUSP stabilizes REST and promotes maintenance of neural progenitor cells. Nat Cell Biol. 2011;13(2):142–52.
- Chauhan D, Tian Z, Nicholson B, Kumar KG, Zhou B, Carrasco R, McDermott JL, Leach CA, Fulcinniti M, Kodrasov MP, et al. A small molecule inhibitor of ubiquitin-specific protease-7 induces apoptosis in multiple myeloma cells and overcomes bortezomib resistance. Cancer Cell. 2012;22(3):345–58.
- Fan YH, Cheng J, Vasudevan SA, Dou J, Zhang H, Patel RH, Ma IT, Rojas Y, Zhao Y, Yu Y, et al. USP7 inhibitor P22077 inhibits neuroblastoma growth via inducing p53-mediated apoptosis. Cell Death Dis. 2013;4:e867.

# Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Submit your manuscript at www.biomedcentral.com/submit

• Maximum visibility for your research

