

REVIEW

Open Access



# Circulating exosomal microRNAs as potential prognostic biomarkers in gastrointestinal cancers: a systematic review and meta-analysis

Elmira Gheytauchi<sup>1†</sup>, Fatemeh Tajik<sup>1†</sup>, Mahdieh Razmi<sup>1\*</sup>, Sadegh Babashah<sup>2</sup>, William Chi Shing Cho<sup>3</sup>, Kiarash Tanha<sup>4</sup>, Maryam Sahlolbeigi<sup>5</sup>, Roya Ghods<sup>1,5\*</sup> and Zahra Madjd<sup>1,5\*</sup>

## Abstract

**Background** Recent reports suggested that circulating exosomal microRNAs (exomiRs) may serve as non-invasive prediction biomarkers in gastrointestinal (GI) cancers, yet their clinicopathological and prognostic values need to be more clarified. Hence, the present meta-analysis was aimed to quantitatively assess the evidence regarding the association between circulating exomiRs and prognosis in GI cancer patients.

**Methods** A comprehensive search was carried out in prominent literature databases, including PubMed, ISI Web of Science, Scopus, and Embase. Odds ratios (ORs) or hazard ratios (HRs) with 95% confidence intervals (CIs) were gathered to evaluate the strength of the association. The quality assessment was investigated through the Newcastle-Ottawa Scale (NOS) and publication bias via Eggers' test and funnel plots.

**Results** A total of 47 studies, comprising of 4881 patients, were considered eligible for this meta-analysis. Both up-regulated and down-regulated circulating exomiRs are significantly associated with differentiation (HR = 1.353, P = 0.015; HR = 1.504, P = 0.016), TNM stage (HR = 2.058, P < 0.001; HR = 2.745, P < 0.001), lymph node metastasis (HR = 1.527, P = 0.004; HR = 2.009, P = 0.002), distant metastasis (HR = 2.006, P < 0.001; HR = 2.799, P = 0.002), worse overall survival (OS) (HR = 2.053, P < 0.001; HR = 1.789, P = 0.001) and poorer disease/relapse/progression-free survival (DFS/RFS/PFS) (HR = 2.086, P < 0.001; HR = 1.607, P = 0.001) in GI cancer patients, respectively. In addition, subgroup analyses based on seven subcategories indicated the robustness of the association. The majority of findings were lack of publication bias except for the association between up-regulated exomiRs and OS or DFS/RFS/PFS and for the down-regulated exomiRs and TNM stage.

<sup>†</sup>Elmira Gheytauchi and Fatemeh Tajik contributed equally to this manuscript as co-first authors

<sup>†</sup>Professor Zahra Madjd as the first corresponding author, Dr Roya Ghods as a second and Dr Mahdieh Razmi as a third corresponding authors have contributed to this work

\*Correspondence:

Mahdieh Razmi

Razmi.mahdieh@ut.ac.ir; Razmi\_mahdyeh@yahoo.com

Roya Ghods

ghods.ro@iums.ac.ir; rghods77@yahoo.com

Zahra Madjd

Zahra.madjd@yahoo.com; majdjabari.z@iums.ac.ir

Full list of author information is available at the end of the article



**Conclusion** This study supports that up- and down-regulated circulating exomiRs are associated with poorer survival outcomes and could be served as potential prognostic biomarkers in GI cancers. Given the limitations of the current findings, such as significant heterogeneity, more investigations are needed to fully clarify the exomiRs prognostic role.

**Keywords** Circulating exosomal microRNAs (exomiRs), Gastrointestinal cancers, Prognostic value, Clinicopathological characteristics, Meta-analysis

## Introduction

Gastrointestinal (GI) cancers, which include esophageal, gastric, pancreas, colorectal, rectal, and hepatocellular carcinomas, are the leading cause of cancer-related deaths, with incidence rates varying among industrialized and developing countries [1–4]. As reported in 2022, the number of new GI cancer cases and deaths were estimated at 343,040 and 171,920 in the USA, respectively [3]. Although the mortality rate of GI cancers has decreased over the last decade owing to multidisciplinary treatment approaches, the global burden of the disease remains considerable, with a notable unfavorable prognosis. The primary factors causing inferior survival outcomes for these individuals are late-stage diagnosis, inadequate prognostic biomarkers, metastasis and recurrence progression, and therapeutic resistance [5, 6].

The extent burden of GI cancers resulted in the planning of innovative molecular–omics landscapes [5–7]. Historically, tissue biopsy, as the standard method, still provides insight into cancer diagnosis and prognosis. However, this approach is invasive, costly, and associated with some complications, including a partial snapshot of the whole tumor, inaccessibility of tumor tissue in terms of anatomic location, biopsy sampling errors, and inter-observer variability in some GI tissues [8, 9]. Therefore, an accurate non-invasive detection technique is urgently warranted to completely elucidate the characteristics of the tumor, allow for early detection of cancer, and precisely evaluate the efficacy of treatment approaches. Compared to traditional tissue biopsy, blood-based or liquid biopsy as the minimally invasive tools provide close monitoring to identify cancer-associated changes and predict prognosis and acquired resistance or disease recurrence before the appearance of clinical symptoms [10–15].

Exosomes have received much attention in recent years as circulating biomarkers for cancer. Exosomes are cell-secreted nano-sized membrane (30 to 100 nm) vesicles involved in intercellular communication and various pathological features of cancers, including invasion, angiogenesis, immune response modulation, and inflammation [16, 17]. Extracellular vesicles (EVs), which may be recovered from a variety of physiological fluids, often reflect the genetic makeup of the cancer cells that originally made them up [19]. Importantly,

microRNAs (miRs) in EVs or exomiRs obtained from blood samples demonstrate the specificity of tumors, indicating that exomiRs may be potential indicators for the diagnosis and prognosis of many malignancies as well as in the age of tailored anticancer therapy [17, 18, 20]. MiRs are endogenous non-coding RNAs subtype with 18–22 nucleotides which mainly modulate cellular gene expression, mostly at the post-transcriptional level. They are involved in many physiological cellular processes, including differentiation, proliferation, and apoptosis [18–21]. Recently, many investigations have reported that aberrant expression of miRs is closely associated with the progression of cancers [22, 23], suggesting miRs as putative biomarkers for diagnosis and prognosis in various tumors, such as colorectal cancer, non-small cell lung carcinoma (NSCLC), and glioma [24–26]. Non-exosomal and exosomal miRNAs (exomiRs) in body fluids are considered as stable "tumor-specific" circulating biomarkers in early diagnosis, prognosis, and screening of various cancer types, including colorectal, esophagus, and hepatocellular cancers [10, 11, 27, 28].

This comprehensive systematic review and meta-analysis was designed to verify the prognostic significance of circulating exomiRs in patients with GI cancers and propose their potential as a non-invasive prognostic tool for monitoring mortality, focusing on clinicopathological outcomes. Deregulated circulating exosomal microRNA(s) in gastrointestinal cancers are shown in Fig. 1.

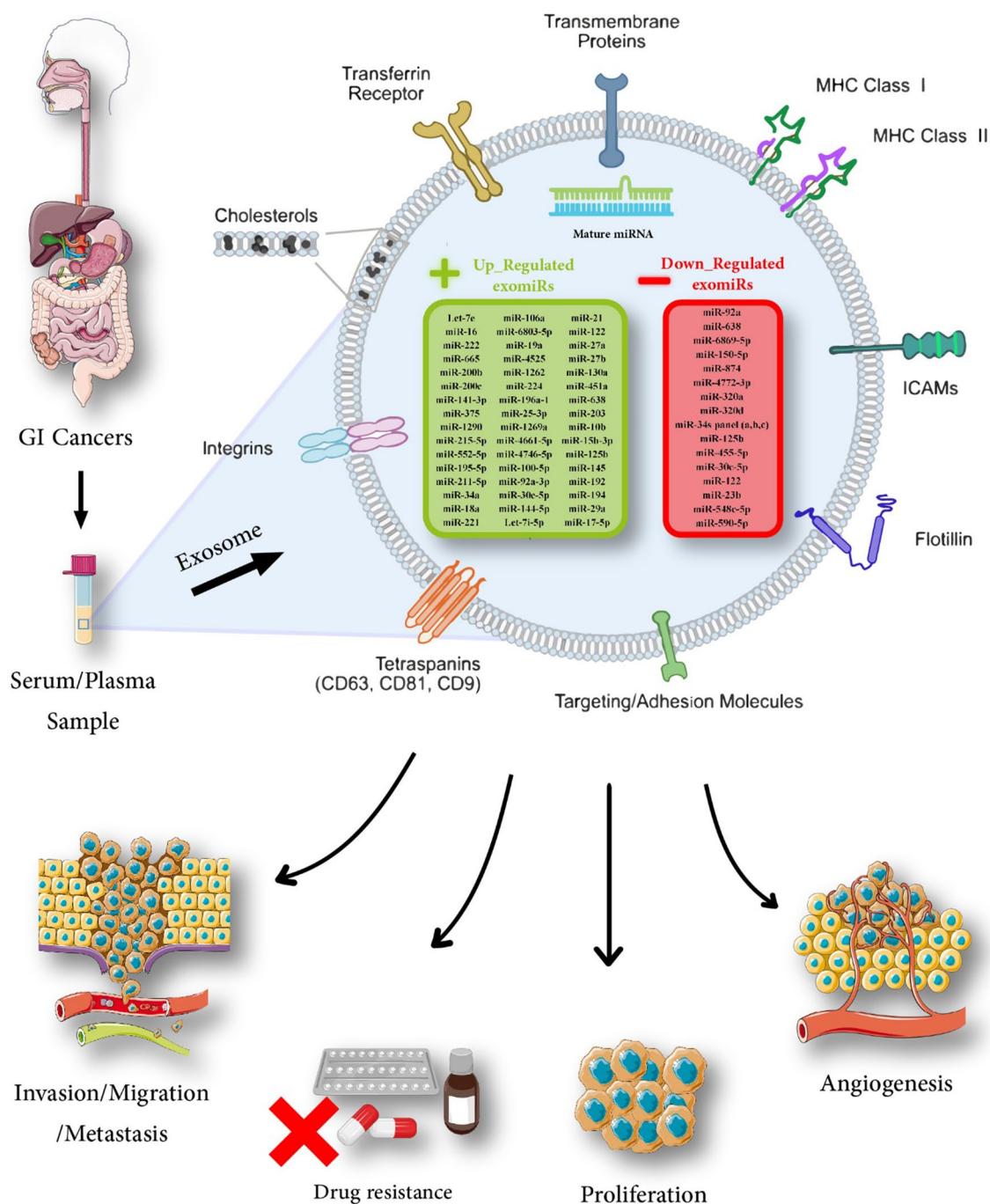
## Material and method

### Protocol and registration

This systematic review and meta-analysis has been registered in the PROSPERO International prospective register of systematic reviews (<http://www.crd.york.ac.uk/PROSPERO>), with the registration number: PROSPERO CRD42017057129; available at [https://www.crd.york.ac.uk/prospero/display\\_record.php?RecordID=57129](https://www.crd.york.ac.uk/prospero/display_record.php?RecordID=57129). In addition, the protocol of the current review has been published in the Systematic Reviews Journal [29].

### Eligibility criteria

The research question has been developed using PFO; "P" as Population, "F" as Prognostic Factors (or models of interest), and "O" as Outcome [30]. The following criteria



<https://smart.servier.com/>  
**Fig. 1** Deregulated circulating exosomal microRNA(s) in gastrointestinal cancers

were incorporated into this systematic review based on the PFO components:

i) Observational studies (case-control, cross-sectional, and cohort studies) assessed the association between circulating exomiRs and GI cancers.

ii) Studies published in English with available full texts.

iii) Studies with human GI cancers, including upper and lower GI and hepatopancreatic biliary.

iv) Studies assessing the association between the circulating exomiRs expression and the prognostic

values consisting of overall survival (OS), disease/relapse/progression-free survival (DFS/RFS/PFS), and/or clinicopathological characteristics of GI cancers.

v) Hazard ratios (HRs) and 95% confidence intervals (CIs) provided in the article, or availability of data to calculate HRs with 95% CIs.

The investigations meeting the following criteria were excluded:

- i) Reviews, meta-analysis, commentaries, case reports, case series studies.
- ii) In vitro and in vivo studies.
- iii) Studies not related to the topic of the interest (e.g., when the studies evaluated the other solid cancers).
- iv) Studies with insufficient and useless data or with unavailable full text.
- v) Studies in which cases received anti-cancer treatment (i.e., chemotherapy and/or radiation therapy) before the biopsy.

#### Information sources

The literature was searched in electronic databases, comprising of Web of Science, PubMed/MEDLINE, Scopus, and Embase until 31st July 2017 and updated on 7th September 2022. Besides, the references of included papers were assessed. Hand searching was performed in key journals that rely on search in Scopus.

#### Search strategy

This study was designed based on the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines (PRISMA 2020) [31]. The following main keywords were used to carry out the search strategy in the mentioned databases: (Neoplasms OR Cancer OR Carcinoma OR Tumor) AND (“Gastrointestinal Tracts” OR “GI Tract” OR “Digestive Tract”) AND (“extracellular vesicles” OR microvesicle OR “Shedding Microvesicles” OR exosomes). The search syntax was adopted in other databases. The search strategy has been fully presented in Additional file 3: Table S1.

#### Selection process

Studies were selected in three phases. Phase 1: duplicated studies were deleted using both EndNote software (version X9.3.3, Thomson Reuters, Philadelphia, USA) and hand searching. Phase 2: two authors (E.Gh. and F.T.) independently screened all records by title and abstract. Phase 3: the same authors independently assessed the full text of each potentially eligible study. Any disagreement

was resolved through consensus and then checked by a third author (M.R.).

#### Data collection process

The data of each eligible study’ was extracted by two authors (E.Gh. and F.T.) independently. The obtained data were entered into a “Data Extraction Form” created by Microsoft Excel for quality assessment and data synthesis. A consensus method was applied between the two reviewers to finalize the validity of all collected data and was then checked by a third author (M.R.).

#### Data items

Data extracted from all eligible papers have consisted of the following items: author’s name, publication year, country, type of exomiRs, type of cancer, expression of exomiRs, detection method, exosome extraction method, sample size, sample type, age, gender, number of patients with up- and down-regulation of circulating exomiRs, clinicopathological parameters (gender, TNM stage of disease, tumor differentiation, lymph node metastasis, distant metastasis), survival data (HR with corresponding 95% CI for OS and DFS/RFS/PFS), cut-off value, and median or mean follow-up times.

#### Study quality assessment

All studies reporting the prognostic values of exomiRs were included in the meta-analysis and assessed according to Newcastle–Ottawa Scale (NOS) tool by two independent investigators (E.Gh. and F.T.). NOS comprises three sections: selection, comparability, and exposure or outcome, with a score ranging from 0 to 9 [32]. This scoring includes four stars for the selection section, two stars for comparability, and three stars for exposure or outcomes. The result of the quality assessment is divided into three categories good, fair, and poor. Moreover, discrepancies between the two authors were resolved by consensus and were then checked by a third author (M.R.).

#### Effect measures and synthesis methods

Comprehensive Meta-Analysis (CMA) software version 2.2.064 was applied to perform all statistical analyses. The HRs and corresponding 95% CIs were recorded for all survival data, including OS and DFS/RFS/PFS. HRs were extracted from both multivariate and univariate statistical tests by preferring information from multivariate statistics if available. For studies without providing HR, we calculated HRs by the Kaplan–Meier curves using the method presented by Parmar et al. [33]. In this regard, survival data were extracted from Kaplan-Meier curves by the software GetData Graph Digitizer (<http://getdata-graph-digitizer.com/>).

Additionally, the combined odds ratios (ORs) and 95% CIs were applied to evaluate the associations between exomiRs expression and clinicopathological features, including gender (male vs. female), TNM stage (III/IV vs. I/II), tumor differentiation (poor vs. moderate/well), lymph node metastasis (positive vs. negative) and distant metastasis (positive vs. negative). A pooled HR/OR larger than one reflected a worse clinical prognostic outcome in GI cancer patients. Heterogeneity among studies was assessed through Cochran's Q statistic and the  $I^2$  index. While an  $I^2$  of over 50% and/or  $P < 0.05$  indicated a large degree of heterogeneity and a random effect model was used, the fixed effect model was utilized in the absence of heterogeneity ( $I^2 \leq 50\%$  or  $P > 0.05$ ). Afterward, subgroup analyses were employed for prognostic outcomes to recognize possible causes of heterogeneity and evaluate the prognostic importance of various subgroups. The level of significance was set at  $P < 0.05$ .

#### Publication bias assessment

Funnel plots were applied to graphically investigate the potential publication bias. In addition, Egger's test was conducted to statistically assess the publication bias [34].

## Results

### Study selection

The preliminary search contained 16,733 references, of which 3120, 4438, 5186, and 3988 papers were retrieved from PubMed/MEDLINE, Web of Science, Scopus, and Embase databases, respectively, published from inception to 7th September 2022. Subsequently, the resulting references were imported to the EndNote reference manager to remove duplicate articles ( $n = 6772$ ). The review articles were excluded ( $n = 649$ ) before screening studies. Of the remained articles ( $n = 9312$ ), 9092 papers were excluded following the subsequent screening of titles and abstracts according to eligibility criteria. As a result, 220 eligible studies remained and entered the next phase. The full text of the remaining studies was evaluated, and of these, 173 were excluded according to the exclusion criteria and unavailable full text. Finally, 47 studies were included in the qualitative synthesis. A flowchart of the search process for eligible studies is shown in Fig. 2.

### Study characteristics

All the enrolled studies were written in English and published between 2015 and 2022, with sample sizes ranging from 4 to 326 patients (Table 1). Geographically, the majority of the papers ( $n = 31$ ) were conducted in China, whereas the remaining papers ( $n = 16$ ) were carried out in other countries (Japan, Egypt, Korea, Spain, Germany, Norway, and Taiwan). In addition, a large number of cases were male in most of the studies.

Of 47 studies, 32 reported circulating exomiRs with high aberrant expression, and 16 reported low expression (Additional file 4: Table S2). There have been various circulating exomiRs, leading to high heterogeneity in our study. Concerning the types of cancer, the two most commonly evaluated GI tract carcinomas were colorectal cancer (CRC) ( $n = 16$ ) and hepatocellular carcinoma (HCC) ( $n = 16$ ), followed by gastric cancer (GC) ( $n = 8$ ), pancreatic ductal adenocarcinoma (PDAC) ( $n = 3$ ), hepatoblastoma (HB) ( $n = 2$ ), pancreatic cancer (PC) ( $n = 1$ ), and locally-advanced rectal cancer (LARC) ( $n = 1$ ). Moreover, the exomiRs were derived from either serum ( $n = 32$ ) or plasma ( $n = 15$ ). Most of the articles ( $n = 38$ ) used quantitative real-time PCR (qRT-PCR) for the detection of exomiR expression; 7 studies employed reverse transcription quantitative PCR (RT-qPCR), and 3 applied RNA sequencing. There have been multiple exosome isolation methods; 28 studies employed extraction kit, and 19 papers applied ultracentrifuge (UC). Besides, 33 studies evaluated the relationship between OS and the expression of circulating exomiRs, while 25 studies investigated the prognostic value of the circulating exomiRs on DFS/RFS.

### Quality assessment in studies

According to the NOS quality assessment tool, the majority of the included studies ( $n = 28$ ) had good quality. The remaining studies ( $n = 18$ ) had poor quality, and only one study had fair quality. A summary of quality assessment for all eligible studies has been shown in Fig. 3.

### Meta-analysis

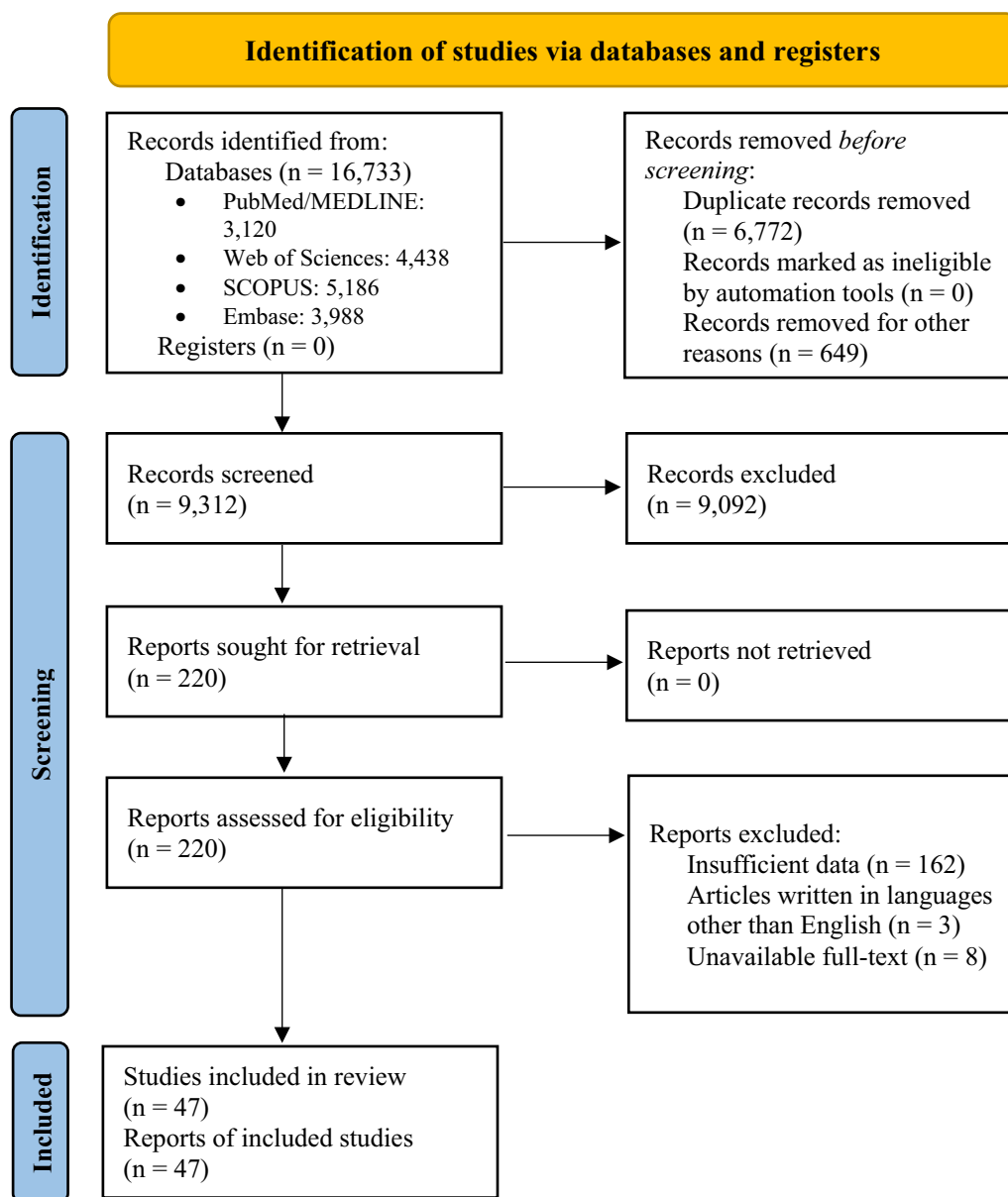
#### Prognostic accuracy and subgroup analyses

A total of 34 studies included in the present meta-analysis assessed the association between deregulated exomiRs ( $n = 52$ ) and OS in individuals suffering from GI cancers. Twenty-four studies provided data regarding exomiRs deregulation ( $n = 29$ ) and DFS/RFS/PFS.

#### Deregulated exomiRs and the overall survival (OS)

Thirty-four studies containing 3833 patients investigated the impact of exomiRs deregulation on OS in patients with GI cancers. As shown in Table 2, since statistical heterogeneity was identified among the investigations ( $I^2 = 76.564\%$ ,  $P < 0.001$ ), a random-effect model was applied to estimate the combined HR. Patients with deregulated exomiRs displayed a statistically significant decrease in OS (pooled HR = 1.998, 95% CI 1.705–2.341,  $P < 0.001$ , Fig. 4).

Additionally, to figure out the overall findings' robustness, subgroup analysis for OS data was conducted according to seven subcategories, including the type of exomiRs deregulation (up or down), type of cancer,



**Fig. 2** Flow-chart for the search strategy according to the Preferred Reporting Items for Systematics Reviews and Meta-Analyses (PRISMA) guideline

sample size, different data extraction methods (direct or indirect), NOS score, and ethnicity (Table 2). Stratified analysis by the different types of exomiRs deregulation indicated a poorer OS for 37 up-regulated exomiRs (HR=2.053, 95% CI 1.720–2.449, P<0.001; I<sup>2</sup>=66.043%, P<0.001), compared to 15 down-regulated exomiRs (HR=1.789, 95% CI 1.251–2.559, P=0.001; I<sup>2</sup>=87.290%, P<0.001). Regarding the cancer types, deregulation of exomiRs was closely associated with inferior OS in cases with CRC (HR=2.928, 95% CI 2.417–3.547, P<0.001, I<sup>2</sup>=19.747%, P=0.204), HCC (HR=1.582,

95% CI 1.264–1.979, P<0.001, I<sup>2</sup>=76.367%, P<0.001), and PDAC (HR=2.514, 95% CI 1.478–4.279, P=0.001, I<sup>2</sup>=48.243%, P=0.085). However, GC (HR=1.353, 95% CI 0.827–2.214, P=0.229, I<sup>2</sup>=88.803%, P<0.001) and LARC (HR=0.958, 95% CI 0.601–1.525, P=0.856, I<sup>2</sup>=0.000%, P=0.506) did not show such association. When the investigations were stratified based on the sample size, a more considerable relationship was identified between worse OS and large sample sizes (≥100, HR=2.306, 95% CI 1.789–2.973, P<0.001; I<sup>2</sup>=81.157%, P<0.001) compared to small sample sizes (<100,

**Table 1** Main characteristics of the studies included in the meta-analysis

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS	
Up-regulation <sup>a</sup>																		
Soeda N 2019 [35]	Japan 2006–2015	miR-21	GC	qRT-PCR	UC	plasma	129	68	90/39	II–III	OS, RFS	OS: R-Multi/ R-Uni RFS: R-Multi/ R-Uni	40.8	76.8	61.6	NA	good	
Sun L 2020 [36]	China NA	miR-122	CRC	qRT-PCR	UC	serum	85	65	46/39	I–IV	OS	R-Multi/ R-Uni	60	NA	NA	NA	poor	
Liu X 2018 [37]	China NA	miR-27a, miR-130a	CRC	qRT-PCR	Exosome isolation kit	plasma	130	64	80/50	I–IV	OS	R-Uni	60	90.9 100	81.8 69.3	0.866 (0.774–0.957) 0.816 (0.730–0.901)	poor	
Wu L 2020 [38]	China 2018–2019	miR-21 miR-210	PC	RT-qPCR	Exosome isolation kit	serum	30	60	20/10 23/7	I–IV	NA	NA	NA	90.0 90.0	80.0 83.0	0.869 0.823	good	
Takahashi K 2018 [39]	Japan 2013–2017	miR-451a	PDAC	qRT-PCR	UC	plasma	50	High expression group: 66 Low expression group: 69	28/22	I–II	OS, DFS	OS: R-Multi/ R-Uni DFS: R-Multi/ R-Uni	30 (range: 5.5–54)	70.8	69.2	NA	good	
Tsukamoto M 2017 [40]	Japan 2002–2012	miR-21	CRC	RT-qPCR	UC	plasma	326	NA	NA	I–IV	OS, DFS	OS: R-Multi DFS: R-Multi	55	NA	NA	NA	poor	
Yokota Y 2021 [41]	Japan 2012–2015	miR-638	HCC	RT-qPCR	Exosome isolation kit	serum	54	High expression group: 73 Low expression group: 70	35/19	NA	DFS	R-Multi/ R-Uni	24	NA	NA	NA	good	
Takano Y 2017 [42]	Japan 2005–2012	miR-203	CRC	RT-qPCR	UC	serum	240	NA	147/93	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: R-Multi/ R-Uni	54	NA	NA	NA	poor	
Tian X 2019 [43]	China 2009–2012	miR-21 miR-10b	HCC	qRT-PCR	ExoQuick kit	serum	124	48	115/9	NA	DFS	R-Multi/ R-Uni	NA	NA	NA	NA	poor	
Wei S.C. 2020 [44]	China NA	miR-15b-3p	GC	qRT-PCR	UC	serum	108	62	80/28	I–IV	OS	E	NA	80.6	74.1	0.820 (0.763–0.876)	good	

**Table 1** (continued)

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS
Xue X. F. 2019 [45]	China 2015–2017	miR-122 miR-125b miR-145 miR-192 miR-194 miR-29a miR-17-5p miR-106a	HCC	qRT-PCR	Exosome isolation kit	serum	80	59	61/19	I–IV	OS	E	24	NA	NA	0.746 (0.650–0.842) 0.650 (0.526–0.774) 0.535 (0.422–0.649) 0.752 (0.658–0.846) 0.738 (0.638–0.838) 0.703 (0.597–0.809) 0.850 (0.764–0.936) 0.704 (0.534–0.873)	good
Yagi T. 2019 [46]	Japan NA	miR-125b	CRC	qRT-PCR	UC	plasma	6	65	4/2	III–IV	PFS	R-Multi/ R-Uni	12	NA	NA	NA	good
Yan S. S. 2018 [47]	China NA	miR-6803-5p	CRC	qRT-PCR	Exosome isolation kit	serum	168	55	100/68	I–IV	OS, DFS	OS; R-Multi DFS;	60	NA	NA	0.739	poor
Matsumura T. 2015 [48]	Japan 1992–2007	miR-19a	CRC	qRT-PCR	UC	serum	209	65	80/129	I–IV	OS, DFS	OS; R-Multi/ R-Uni DFS;	55,68	NA	NA	NA	poor
Kawamura S. 2019 [49]	Japan 2013–2018	miR-4525 miR-451a miR-21	PDAC	qRT-PCR	UC	plasma	55	miR-4525: 66 miR-451a: 69 miR-21: 70	33/22	I–II	OS, DFS	OS; R-Multi/ R-Uni DFS;	60	86.4 77.3 72.7	81.8 72.7 72.7	NA	poor
Abd El Gwad A. 2018 [50]	Egypt 2015–2016	miR-1262	HCC	qRT-PCR	UC	serum	60	58	43/17	N/A	RFS	E	44	80.0	95.0	0.847	good
Cui Y. 2019 [51]	China 2014–2016	miR-224	HCC	qRT-PCR	Exosome isolation kit	serum	89	59	43/46	N/A	OS	E	NA	NA	NA	0.910 (0.840–0.980)	fair



**Table 1** (continued)

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS
Feng C. 2019 [52]	China NA	miR-196a-1	GC	qRT-PCR	Exosome isolation kit	plasma	86	NA	NA	I—IV	OS	E	NA	NA	NA	NA	poor
Liu W. B. 2016 [53]	China 2008–2013	miR-21	HB	qRT-PCR	ExoQuick kit	serum	32	NA	17/15	I—IV	DFS	R-Multi/ R-Uni	42	NA	NA	0.861 (0.752–0.935)	poor
Cho H. J. 2020 [54]	Korea 2014–2018	miR-25-3p miR-1269a miR-4661-5p miR-4746-5p	HCC	Cohort1: NGS-RNA-Seq Cohort 2&3: qRT-PCR	ExoQuick kit	serum	72	55	58/14	I—IV	NA	NA	NA	71.4	38.0	0.704 (0.629–0.772)	good
Han J. Y. 2020 [55]	China 2015–2017	hsa-miR-100-5p hsa-miR-92a-3p hsa-miR-30e-5p hsa-miR-144-5p hsa-let-7f-5p has-miR-16	CRC	qRT-PCR	Exosome isolation kit	plasma	139	60	77/62	III—IV	OS	R-Uni	NA	NA	NA	0.637 (0.545–0.729) 0.659 (0.568–0.751) 0.694 (0.606–0.782) 0.791 (0.718–0.864) 0.650 (0.559–0.742) 0.721 (0.638–0.804) 0.746 (0.664–0.827) 0.778 (0.702–0.854)	good
de Miguel-Pérez D 2020 [56]	Spain NA	miR-92a miR-222	CRC	qRT-PCR	UC	serum	44	55	30/14	NA	OS, DFS/ PFS	miR-92a OS: R-Uni DFS/PFS: R-Multi/ R-Uni miR-222 OS: R-Multi/ R-Uni	27	NA	NA	0.951 (0.900–1.000) 0.896 (0.810–0.980)	good
Qu Z. 2017 [57]	China NA	miR-665	HCC	qRT-PCR	ExoQuick kit	serum	30	60	12/18	I—IV	OS	E	NA	NA	NA	NA	poor

**Table 1** (continued)

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS
Reese M. 2020 [58]	Germany 2015–2018	miR-200b miR-200c	PDAC	qRT-PCR	UC	serum	56	60	36/20	I–IV	OS	miR-200b OS: R-Multi/ R-Uni miR-200c OS: R-Multi/ R-Uni	13	NA	NA	0.790 (0.680–0.890) 0.670 (0.550–0.790)	good
Meltzer S. 2019 [59]	Norway NA	miR-141-3p miR-375	LARC	qRT-PCR	Exosome isolation kit	plasma	64	60	36/28	NA	OS, PFS	OS: R-Uni PFS: R-Uni	65 (range 4–66)	NA	NA	NA	good
Wang Q. 2021 [60]	China NA	miR-1290	HCC	qRT-PCR	Exosome isolation kit/UC	serum	49	55	37/12	I–IV	NA	NA	NA	NA	NA	NA	poor
Zhang Y. 2021 [60]	China NA	miR-215-5p	GC	qRT-PCR	ExoQuick kit	serum	118	60	70/48	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: E	NA	97.1	68.6	0.866	good
Zhu L. 2022 [61]	China 2020–2021	miR-552-5p	GC	qRT-PCR	UC	plasma	30	56	16/14	I–IV	NA	NA	NA	NA	NA	NA	good
Yang J. 2022 [62]	China NA	miR-195-5p miR-211-5p	GC	RT-qPCR	UC	plasma	108	62.4 ± 8.9 62.2 ± 9.4	80/28 79/27	I–IV	NA	NA	NA	NA	NA	0.745 (0.584–0.906) 0.798 (0.656–0.940)	good
Chen S. 2022 [63]	China NA	miR-34a	HCC	qRT-PCR	UC	serum	60	55	60/60	I–IV	OS	R-Multi/ R-Uni	6–60	51.7	78.3	0.664 (0.572–0.747)	good
Huang C. Y. 2021 [64]	China 2017–2018	let-7e miR-18a miR-27b miR-221 miR-20b miR-652	HCC	RT-qPCR	UC	plasma	40	68	13/7	NA	OS, DFS	OS: R-Uni DFS: R-Uni	80	NA	NA	NA	poor
Hao Y. J. 2022 [65]	Taiwan 2019–2020	miR-21	CRC	qRT-PCR	Exosome isolation kit	plasma	113	65	77/36	I–IV	DFS, PFS	E	NA	NA	NA	NA	poor
Soeda N. 2019 [35]	Japan 2006–2015	miR-92a	GC	qRT-PCR	UC	plasma	129	68	90/39	II–III	OS, RFS	OS: R-Multi/ R-Uni RFS: E	40.8	60.7	63.0	NA	good

Down-regulation<sup>a</sup>

**Table 1** (continued)

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS
Yan S. 2017 [66]	China 2008–2014	miR-638	CRC	qRT-PCR	Exosome isolation kit	serum	192	58	108/84	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: E	47	NA	NA	NA	poor
Yan S. 2018 [67]	China 2012–2014	miR-6869-5p	CRC	qRT-PCR	Exosome Isolation Kit	serum	142	59	85/57	I–IV	OS	R-Multi/ R-Uni	36	NA	NA	NA	good
Zou S. L. 2019 [68]	China NA	miR-150-5p	CRC	qRT-PCR	ExoQuick kit	serum	133	60	49/84	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: E	60	76.1	81.0	0.870	poor
Zhang N. 2020 [69]	China NA	miR-874	CRC	RT-qPCR	Exosome Isolation kit	serum	125	60	76/49	I–IV	OS	R-Multi/ R-Uni	NA	78.6	80.8	0.818	poor
Liu C. 2016 [70]	China 2006–2011	miR-4772-3p	CRC	qRT-PCR RNA-seq	ExoQuick kit	serum	84	57	53/31	II–III	OS	R-Multi	51 (range: 45–64)	77.1	78.6	0.720 (0.590–0.850)	poor
Hao X. 2020 [71]	China 2012–2013	miR-320a	HCC	qRT-PCR	ExoQuick kit	serum	104	60	77/27	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: E	NA	80.0	77.9	0.860	good
Jiao C. W. 2017 [72]	China 2007–2015	miR-34 s panel miR-34a miR-34b miR-34c	HB	qRT-PCR	ExoQuick kit	serum	89	NA	52/37	I–IV	DFS	R-Multi/ R-Uni	54	NA	NA	0.831(0.071–0.371) 0.813(0.023–0.296) 0.837(0.004–0.342)	good
Li W. 2020 [73]	China NA	miR-320d	HCC	qRT-PCR	Exosome isolation kit	serum	110	60	98/12	I–IV	OS, DFS	OS: R-Multi DFS: E	NA	NA	NA	0.869	good
Liu W. F. 2017 [74]	China 2012	miR-125b	HCC	qRT-PCR	ExoQuick kit	serum	128	50	110/18	I–III	OS	R-Multi/ R-Uni	2.9–52.4	53.4	82.5	0.702 (0.602–0.802)	good
Sheng L. Q. 2020 [75]	China 2017–2018	miR-455-5p miR-30c-5p	HCC	exomiRs sequencing	Exosome isolation kit	plasma	cohort1:24 cohort2:27	NA	cohort1:36/8 cohort2:8/4	N/A	OS	E	NA	NA	NA	NA	poor
Shi M. 2018 [76]	China 2008–2011	miR-638	HCC	qRT-PCR	Exosome isolation kit	serum	126	65	70/56	I–IV	OS	R-Multi/ R-Uni	81.5	NA	NA	NA	good
Suehiro T. 2018 [76]	Japan 2006–2013	miR-122	HCC	qRT-PCR	ExoQuick kit	serum	75	73	49/26	NA	DFS	R-Multi	47	NA	NA	NA	poor

**Table 1** (continued)

Authors, publication year	Country, Duration (year)	Exosomal miR(s)	Cancer type	Detection method	Exosome extraction method	sample	Sample size (cancer)	mean/median age (year)	Gender cancer (M/F)	TNM Stage	Outcome	Hazard ratio (HR)	Follow up (month)	Spec %	Sen %	AUC (95% CI)	NOS	
Kumata Y. 2018 [77]	Japan 2006–2013	miR-23b	GC	qRT-PCR	UC	plasma	232	NA	165/67	I–IV	OS, DFS	OS: R-Multi/ R-Uni DFS: R-Multi/ R-Uni	45.6 (range: 4.8–127.2)	NA	NA	NA	NA	poor
Peng Z. Y. 2018 [78]	China 2008–2014	miR-548c-5p	CRC	qRT-PCR	Exosome isolation kit	serum	108	58	61/47	I–IV	OS	R-Multi/ R-Uni	44	NA	NA	NA	NA	poor
Zheng G-D. 2021 [79]	China 2008–2011	miR-590-5p	GC	qRT-PCR	UC	serum	168	60	117/51	I–IV	OS	R-Multi/ R-Uni	64.2 (range: 43.3–92)	86.0	63.7	0.810 (0.751–0.860)	good	

GC Gastric Cancer, CRC Colorectal Cancer, LARC Locally Advanced Rectal Cancer, HCC Hepatocellular Carcinoma, HB Hepatoblastoma, PC Pancreatic Cancer, PDAC Pancreatic Ductal Adenocarcinoma, UC Ultracentrifuge, microRNA miR, NA Not Applicable, OS Overall Survival, DFS Disease Free Survival, PFS Progression Free Survival, RFS Relapse Free Survival, E Estimated, R-Multi Reported Multivariate, R-Uni Reported Univariate, qRT-PCR quantitative real-time PCR, RT-qPCR Reverse transcription quantitative PCR, Sen Sensitivity, Spec Specificity, AUC Area Under Curve.

<sup>a</sup>The up- and down-regulation of exomiR(s) has been reported in comparison with the control group

**Table 2** The results of meta-analyses for the association between deregulated exomiRs and overall survival (OS), and disease/relapse/progression-free survival (DFS/RFS/PFS) in patients with GI cancers

Study groups	Included exomiRs	Test of association		Test of heterogeneity	
		HR (95% CI)	P-value	I <sup>2</sup> %	P-value
OS					
All studies	52	1.998 (1.705–2.341)	<0.001	76.564	<0.001
The type of exomiRs deregulation					
Up-regulation	37	2.053 (1.720–2.449)	<0.001	66.043	<0.001
Down-regulation	15	1.789 (1.251–2.559)	0.001	87.290	<0.001
Type of cancer					
Colorectal	21	2.928 (2.417–3.547)	<0.001	19.747	0.204
Gastric	7	1.353 (0.827–2.214)	0.229	88.803	<0.001
Hepatocellular	16	1.582 (1.264–1.979)	<0.001	76.367	<0.001
Pancreatic	6	2.514 (1.478–4.279)	0.001	48.243	0.085
locally advanced rectal	2	0.958 (0.601–1.525)	0.856	0.000	0.506
Sample size					
≥ 100	29	2.306 (1.789–2.973)	<0.001	81.157	<0.001
< 100	23	1.659 (1.359–2.025)	<0.001	65.182	<0.001
Sample type					
Plasma	27	1.890 (1.497–2.386)	<0.001	82.109	<0.001
Serum	25	2.125 (1.752–2.578)	<0.001	51.047	0.002
Survival analysis					
Direct	44	2.114 (1.719–2.601)	<0.001	79.360	<0.001
Indirect	8	1.559 (1.368–1.776)	<0.001	1.170	0.420
NOS score					
≥ 7	25	2.165 (1.577–2.972)	<0.001	78.093	<0.001
< 7	27	1.850 (1.534–2.231)	<0.001	74.863	<0.001
Ethnicity					
Asian	46	2.085 (1.746–2.489)	<0.001	78.377	<0.001
Caucasian	6	1.327 (0.934–1.886)	0.115	25.255	0.245
DFS/RFS/PFS					
All studies	29	1.920 (1.641–2.245)	<0.001	55.871	<0.001
Deregulation					
Up-regulation	21	2.086 (1.725–2.522)	<0.001	41.988	0.023
Down-regulation	8	1.607 (1.218–2.122)	0.001	72.320	0.001
Type of cancer					
Colorectal	9	2.105 (1.736–2.554)	<0.001	0.000	0.734
Gastric	4	1.408 (0.781–2.539)	0.256	86.773	<0.001
Hepatocellular	10	1.923 (1.651–2.239)	<0.001	0.000	0.740
Pancreatic	4	3.195 (2.019–5.056)	0.000	0.000	0.839
Locally advanced rectal	2	1.034 (0.758–1.410)	0.834	0.000	0.567
Sample size					
≥ 100	15	1.911 (1.520–2.402)	<0.001	64.800	<0.001
< 100	14	1.890 (1.506–2.373)	<0.001	45.085	0.034
Sample type					
Plasma	13	1.819 (1.317–2.510)	<0.001	74.592	<0.001
Serum	16	1.906 (1.680–2.163)	<0.001	0.000	0.633
Survival analysis					
Direct	20	2.019 (1.594–2.558)	<0.001	67.219	<0.001
Indirect	9	1.809 (1.552–2.109)	<0.001	0.000	0.725

**Table 2** (continued)

Study groups	Included exomiRs	Test of association		Test of heterogeneity	
		HR (95% CI)	P-value	I <sup>2</sup> %	P-value
NOS score					
≥ 7	14	1.803 (1.498–2.170)	<0.001	41.843	0.050
< 7	15	2.062 (1.571–2.706)	<0.001	65.905	<0.001
Ethnicity					
Asian	26	1.998 (1.705–2.341)	<0.001	50.425	0.002
Caucasian	3	1.109 (0.800–1.538)	0.535	10.141	0.329

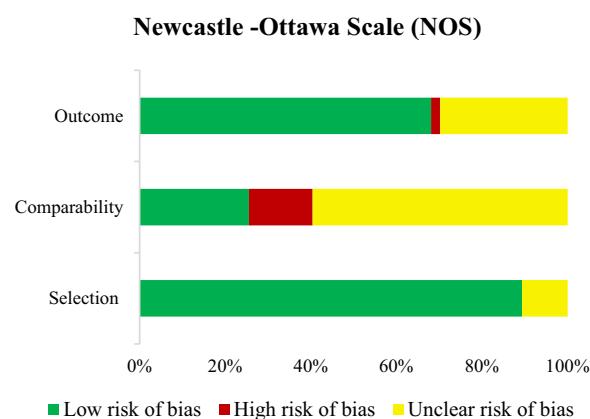
HR = 1.659, 95% CI 1.359–2.025,  $P < 0.001$ ;  $I^2 = 65.182\%$ ,  $P < 0.001$ ). Subgroup analysis of OS for exomiRs de-regulation in plasma demonstrated considerable association (HR = 1.890, 95% CI 1.497–2.386,  $P < 0.001$ ;  $I^2 = 82.109\%$ ,  $P < 0.001$ ), like serum (HR = 2.125, 95% CI 1.752–2.578,  $P < 0.001$ ;  $I^2 = 51.047\%$ ,  $P = 0.002$ ). In subgroup analysis stratified by different data extraction methods, exomiRs deregulation revealed a more significant connection with the worse OS in the HR presented in the articles (HR = 2.114, 95% CI 1.719–2.601,  $P < 0.001$ ;  $I^2 = 79.360\%$ ,  $P < 0.001$ ) than that calculated from the survival curves (HR = 1.559, 95% CI 1.368–1.776,  $P < 0.001$ ;  $I^2 = 1.170\%$ ,  $P = 0.420$ ). When categorized by quality assessment, deregulated exomiRs was related to poorer OS in both high- (HR = 2.165, 95% CI 1.577–2.972,  $P < 0.001$ ;  $I^2 = 78.093\%$ ,  $P < 0.001$ ) and low-quality publications (HR = 1.850, 95% CI 1.534–2.231,  $P < 0.001$ ;  $I^2 = 74.863\%$ ,  $P < 0.001$ ) (Additional file 1: Figure S1). Finally, in subgroup analysis stratified by ethnicity, deregulation of exomiRs revealed a significant connection with the worse OS in the articles published in Asian countries (HR = 2.085, 95% CI 1.746–2.489,  $P < 0.001$ ;  $I^2 = 78.377\%$ ,  $P < 0.001$ ) unlike articles from Caucasian countries

with an HR of 1.327 (95% CI 0.934–1.886,  $P = 0.115$ ;  $I^2 = 25.255\%$ ,  $P = 0.245$ ).

#### Deregulated exomiRs and the disease/relapse/progression-free survival (DFS/RFS/PFS)

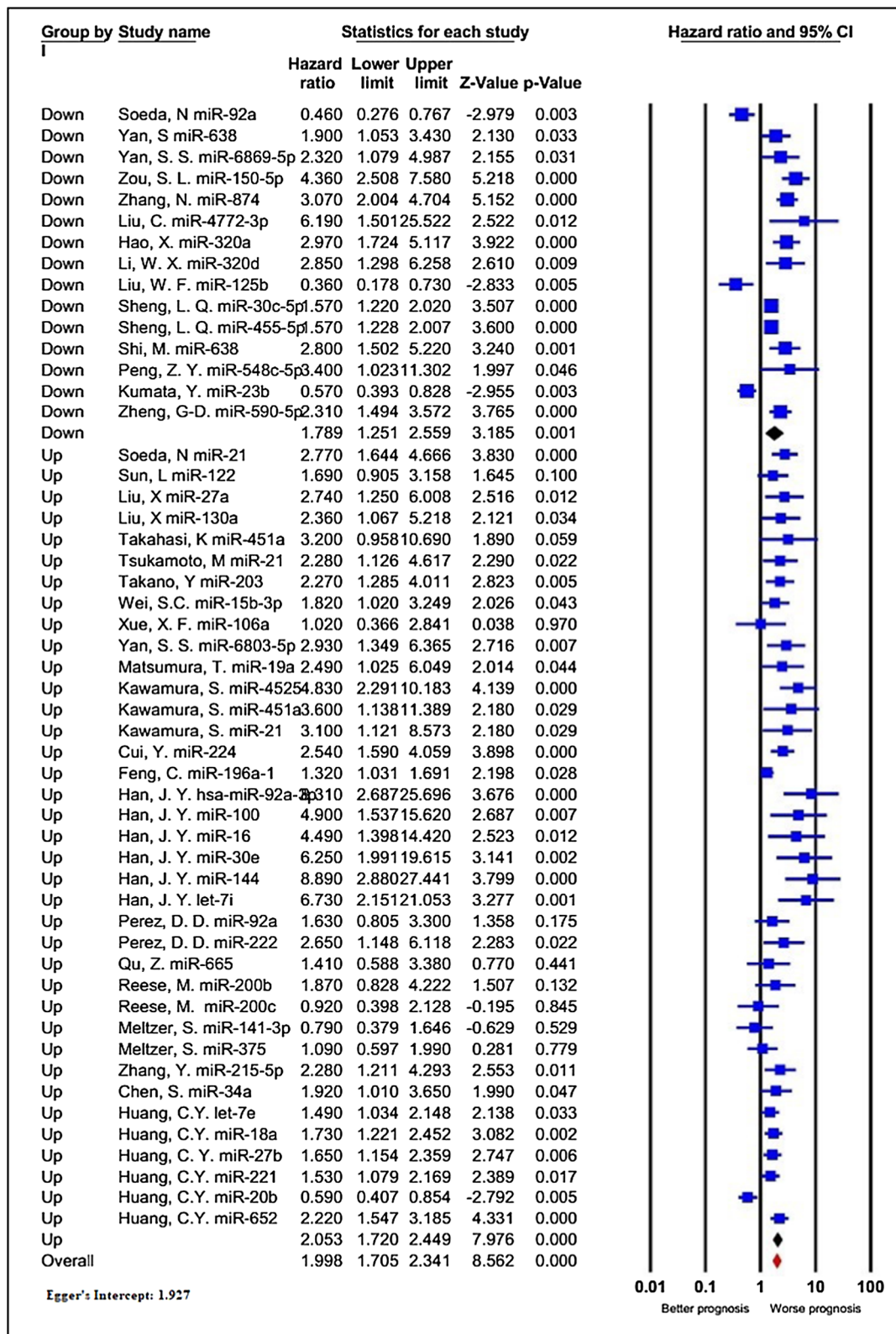
As indicated in Table 2, twenty-nine studies containing 2767 patients reported the data regarding DFS/RFS/PFS, and relatively significant heterogeneity was identified among these investigations ( $I^2 = 55.871\%$ ,  $P < 0.001$ ). The pooled results through a random-effects model revealed a significant link between deregulated exomiRs and poorer DFS/RFS/PFS in patients (HR = 1.920, 95% CI 1.641–2.245,  $P < 0.001$ , Fig. 5).

Following that, subgroup analyses were performed to further investigate the potential predictive significance of the SMYD family members according to the same seven categories utilized for OS (Table 2). The results demonstrated a remarkable connection between deregulated exomiRs and worse DFS/RFS/PFS in all the stratified analyses performed, except patients with GC (HR = 1.408, 95% CI 0.781–2.539,  $P = 0.256$ ;  $I^2 = 86.773\%$ ,  $P < 0.001$ ), LARC (HR = 1.034, 95% CI 0.758–1.410,  $P = 0.834$ ;  $I^2 = 0.000\%$ ,  $P = 0.567$ ), and articles published in Caucasian (HR = 1.109, 95% CI 0.800–1.538,  $P = 0.535$ ;  $I^2 = 10.141\%$ ,  $P = 0.329$ ) (Table 2).

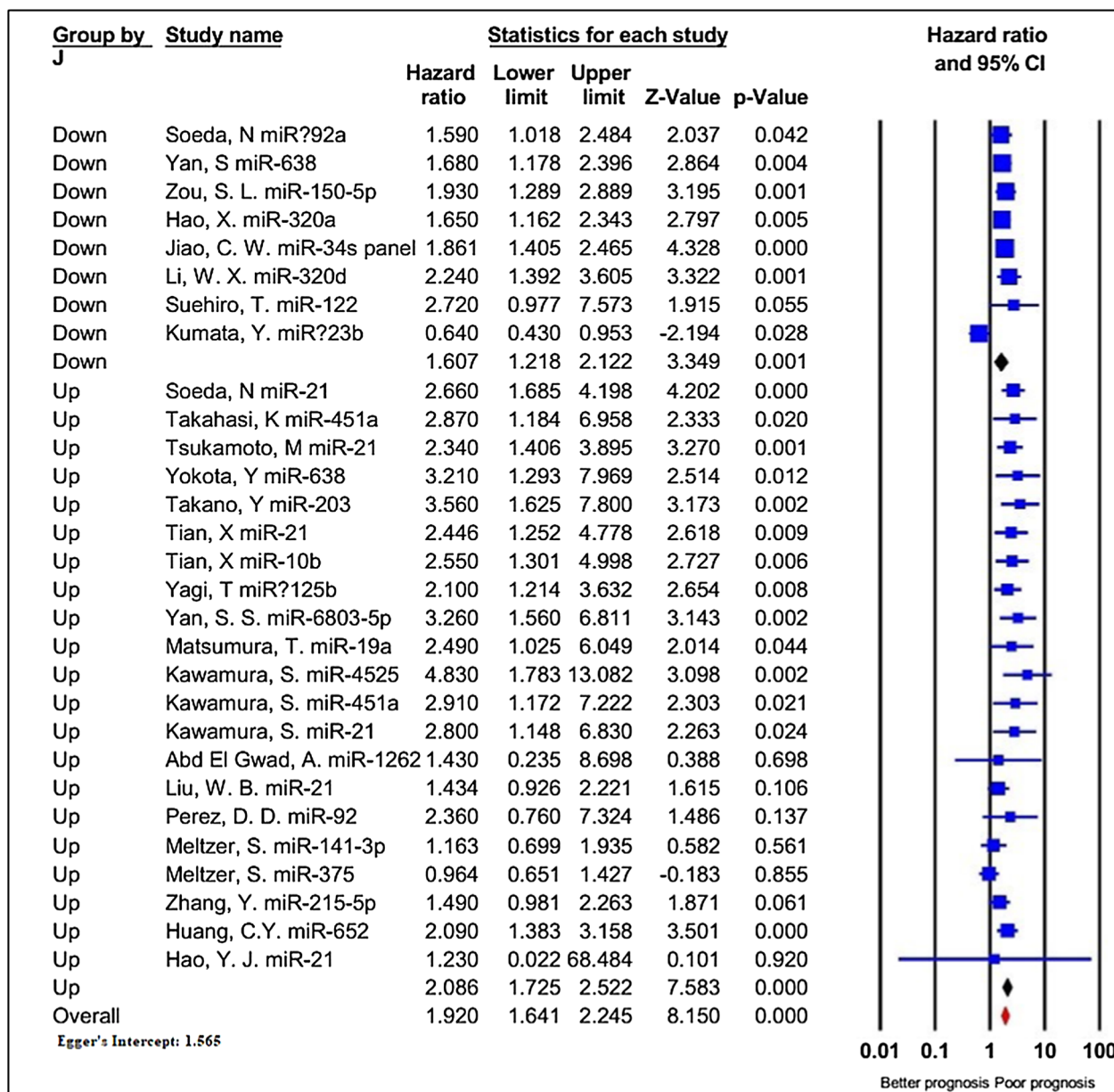
**Fig. 3** Summary of quality assessment

#### Association between deregulated exomiRs and clinicopathological characteristics

**Gender** The relationship between up-regulated exomiRs and gender was evaluated in 27 studies with 2479 patients, while down-regulated exomiRs were reported in 13 studies with 1736 patients. As shown in Additional file 5: Table S3, pooled results from fixed-effects framework ( $I^2 = 19.601\%$ ,  $P = 0.182$ ) indicated that up-regulated exomiRs did not associate with gender (OR = 0.992, 95% CI 0.826–1.190,  $P = 0.927$ ), as in down-regulated exomiRs (OR = 0.986, 95% CI 0.796–1.222,  $P = 0.900$ ;  $I^2 = 0.000\%$ ,  $P = 0.826$ ).



**Fig. 4** Forest plot of the association between exomiRs deregulation and overall survival in patients with GI cancers, stratified by the type of exomiRs deregulation (down-regulated exomiRs and up-regulated exomiRs)



**Fig. 5** Forest plot of the association between exomiRs deregulation and disease/relapse/progression-free survival (DFS/RFS/PFS) in patients with GI cancers, stratified by the type of exomiRs deregulation (down-regulated exomiRs and up-regulated exomiRs)

**TNM stage** Data from 21 and 13 investigations comprising of 2288 and 1735 patients were collected and pooled to reveal a connection between up- and down-regulated exomiRs and TNM stage, respectively. Based on the random-effect framework ( $I^2=71.244\%$ ,  $P<0.001$ ), it was found that the GI cancer patients with up-regulated exomiRs tended towards the advanced TNM stage (OR=2.058, 95% CI 1.410–3.003,  $P<0.001$ , Fig. 6A, Additional file 5: Table S3). In addition, the pooled OR indicated that down-regulated exomiRs were directly

correlated with a higher TNM stage (HR=2.745, 95% CI 1.621–4.648,  $P<0.001$ , Fig. 6B, Additional file 5: Table S3).

**Differentiation** A total of 20 studies 2,425 patients, evaluated a possible connection between up-regulated exomiRs and differentiation (Fig. 6C and Additional file 5: Table S3). The pooled OR through a random effects model ( $I^2=37.862\%$ ,  $P=0.045$ ) found statistically significant results (OR=1.353, 95% CI: 1.060–1.726,  $P=0.015$ ). Analysis based on 13 studies through a random-effects





model ( $I^2=55.128\%$ ,  $P=0.008$ ) showed that the down-regulated exomiRs correspondent with poorly-differentiated cancer cells (OR=1.504, 95% CI: 1.079–2.099,  $P=0.016$ ; Fig. 6D and Additional file 5: Table S3).

**Lymph node metastasis** A total of 19 studies consisting of 2121 cases focused on the dependability between the up-regulated exomiRs and LNM. The overall pooled HR, under a random-effect model ( $I^2=47.603\%$ ,  $P=0.011$ ), indicated that up-regulated exomiRs had a statistically significant relationship with LNM (OR=1.527, 95% CI 1.141–2.042,  $P=0.004$ ; Fig. 6E, Additional file 5: Table S3). Afterward, a clear association was identified between down-regulated exomiRs and LNM (OR=2.009, 95% CI 1.293–3.122,  $P=0.002$ ; Fig. 5F, Additional file 5: Table S3) with significant heterogeneity ( $I^2=60.459\%$ ,  $P=0.013$ ).

**Distant metastasis** The relationship between up-regulated exomiRs and distant metastasis was demonstrated in eight studies with 876 cases, while down-regulated exomiRs were reported in three studies with 467 patients. Statistically, non-significant heterogeneity was observed for high expression exomiRs ( $I^2=2.450\%$ ,  $P=0.411$ ) and low expression exomiRs ( $I^2=0.000\%$ ,  $P=0.930$ ); consequently, a fixed-effect model was employed to combine the results. The findings showed that cases with down-regulated exomiRs were more likely to develop distant metastasis (OR=2.799, 95% CI 1.448–5.408,  $P=0.002$ ; Fig. 6H, Additional file 5: Table S3), as in up-regulated exomiRs (OR=2.006, 95% CI 1.358–2.962,  $P<0.001$ ; Fig. 6G, Additional file 5: Table S3).

#### Publication Bias

Funnel plot (Fig. 7) and Egger's test (Additional file 5: Table S3) were also conducted to identify potential publication bias of all analyses. The P-values for Egger's test, measuring the asymmetry of performed analyses, indicated statistically non-significant publication bias for all analyses except for OS (overall deregulated exomiRs ( $P=0.002$ ) and up-regulated exomiRs ( $P<0.001$ ), DFS/RFS/PFS (overall deregulated exomiRs ( $P=0.021$ ) and up-regulated exomiRs ( $P=0.023$ ), TNM stage (down-regulated exomiRs ( $P=0.027$ ) and distant metastasis (up-regulated exomiRs ( $P=0.026$ )). The funnel plots, displaying potential publication bias, are shown in Fig. 7.

#### Association between circulating exomiR-21 and prognostic/clinicopathological characteristics

To analyze the same types of circulating exomiRs in GI cancers, we considered the exomiR-21 as the most overexpressed miRNA in GI tumors. The results of our analysis indicated a statistically significant association between the exomiR-21 and overall survival (HR=2.655, 95% CI 1.802–3.912,  $P<0.001$ ), disease/relapse/progression-free survival (HR=2.127, 95% CI 1.674–2.703,  $P<0.001$ ), and Lymph node metastasis (HR=1.568, 95% CI 1.118–2.198,  $P=0.009$ ) (Additional file 2: Figure S2). Furthermore, no association was found between circulating exomiR-21 and other clinicopathological characteristics.

#### Discussion

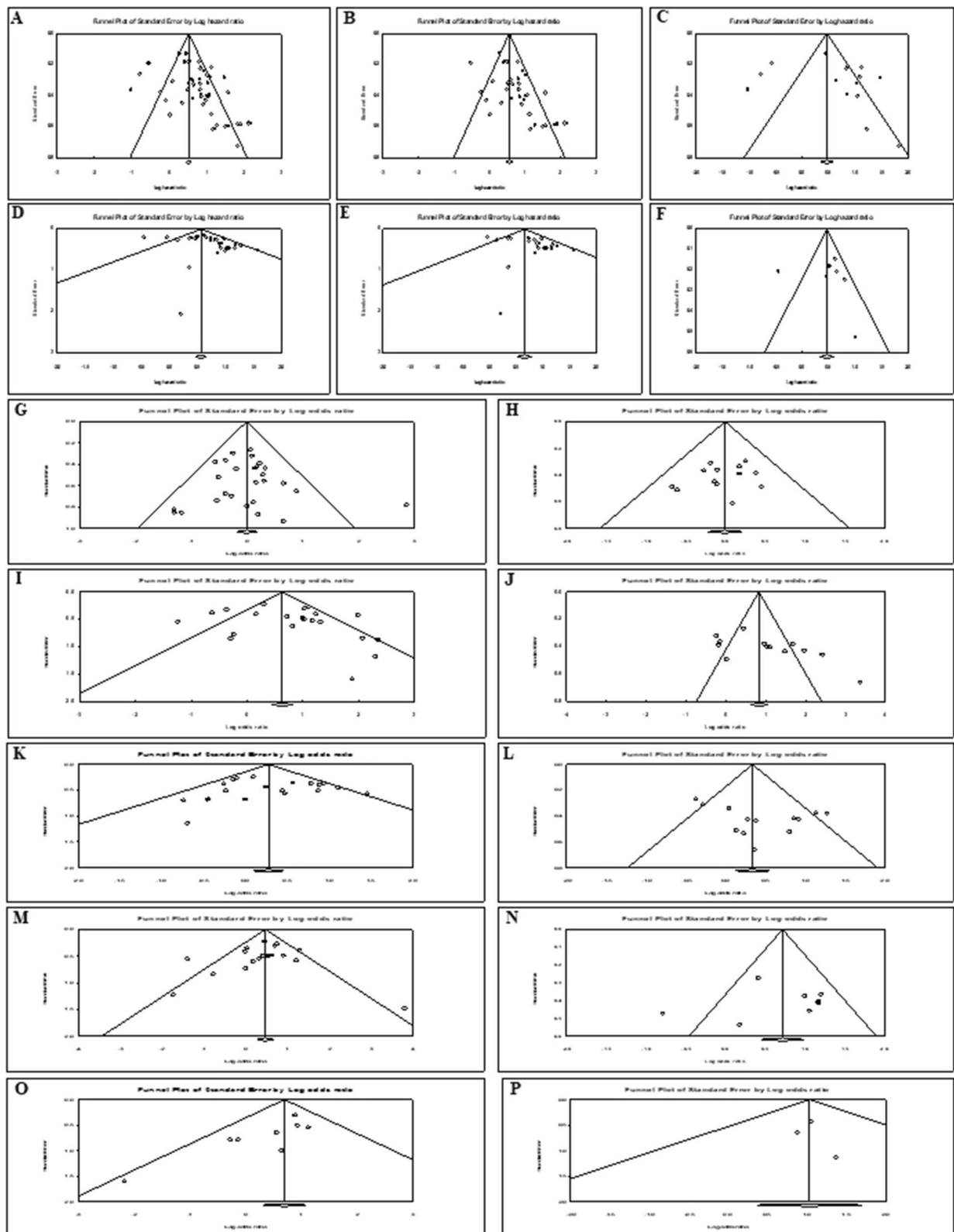
Exploring novel prognostic biomarkers is crucial for improving management, therapy, and prognosis in patients with GI cancers. Compelling evidence has cleared that deregulated exomiRs, as the hallmarks of most human cancers, regulate a plethora of biological pathways, including maintaining proliferative potential, escaping suppressors, repelling cell death, and triggering invasion metastasis, drug resistance, and angiogenesis [80].

Recently, serum exomiRs were applied as feasible and non-invasive serological biomarkers for several cancers, including GI cancers [41, 48, 81–88]. Therefore, in this systematic review and meta-analysis, we aimed to evaluate the prognostic capacity of circulating exomiRs GI cancers. The four principal databases (Web of Science, PubMed/MEDLINE, Scopus, and Embase) were searched to ensure the identification of all relevant studies. To the best of our knowledge, this is the first comprehensive meta-analysis comprising a wide range of circulating exomiRs with 4881 participants to provide strong evidence regarding the potential clinical applicability of circulating exomiRs in GI cancers. Notably, we attempted to clarify study heterogeneity and publication bias.

In general, we combined the data regarding the deregulation of all exomiRs in our meta-analysis to perceive their potential roles to predict the prognosis. The results emerging from this meta-analysis advocated that deregulated exomiRs were significantly associated with reduced OS (HR=1.998) and dismal DFS/RFS/

(See figure on next page.)

**Fig. 7** Funnel plot of the publication bias. **A** the association between overall exomiRs deregulation and OS. **B, C** the association between up-regulated **B** and down-regulated **C** exomiRs and OS. **D** the association between overall exomiRs deregulation and disease/relapse/progression-free survival (DFS/RFS/PFS). **E, F** the association between up-regulated **E** and down-regulated **F** exomiRs and DFS/RFS/PFS. **G–P** the association between up-regulated and down-regulated exomiRs and clinicopathological characteristics, including gender **G, H**, TNM stage **I, J**, Differentiation **K, L**, Lymph node metastasis **M, N**, Distant metastasis **O, P**, respectively



**Fig. 7** (See legend on previous page.)

PFS (HR=1.920), supporting the statement that the aberrant expression of exomiRs as minimally invasive biomarkers can predict prognostic outcomes in GI cancers patients. The majority of subcategories, including the type of exomiRs deregulation (up or down), the majority of cancer types (colorectal, hepatocellular, and pancreatic cancers), sample size ( $\geq 100$  or  $< 100$ ), various data extraction methods (direct or indirect), sample type (plasma or serum), and NOS score (high or low-quality publications), were also strongly associated with poor survival outcomes (OS or DFS/RFS/PFS), strengthening the prognosis. Only some types of cancers (gastric cancer and locally advanced rectal cancer) and Caucasian ethnicity did not show a link with poor survival outcomes. Based on obtained findings from OS, CRC (HR=2.928), HCC (HR=1.582), and PDAC (HR=2.514) are likely to have a significant association with poor prognosis while other cancers of the GI tract, including GC (HR=1.353,  $P=0.229$ ) and LARC (HR=0.958,  $P=0.857$ ) did not show such association. Even the association between DFS/RFS/PFS and the same types of cancers including CRC (HR=2.105), HCC (HR=1.923), and PDAC (HR=3.195) remained significant. This finding highlights the similarity of results related to different types of prognostic reports (OS vs. DFS/RFS/PFS) and GI cancers. Moreover, we did not find any significant association between survival findings and LARC or GC. It is assumed that the assorted outcomes and the insufficient number of included studies might have affected these findings [89, 90]. It was reported that variability in outcomes prediction and categorization of LARC is due to the diversity of tumors and the complexity of diagnostic and prognostic tools in clinical practice [91–93].

We observed partially high heterogeneity in most of our prognostic findings, which is mainly due to the considerable variability within circulating exomiRs, type of cancer, and follow-up duration of the included studies. Moreover, publication bias was only detected between OS and overall deregulated exomiRs ( $P=0.002$ ) or up-regulated exomiRs ( $P<0.001$ ), as well as DFS and overall deregulated exomiRs ( $P=0.021$ ) or up-regulated exomiRs ( $P=0.023$ ).

The clinical significance of circulating exomiRs with abnormally elevated or reduced expression in various malignancies has been highlighted by an increasing body of research [79, 98, 99]. We examined 47 studies on 62 distinct exomiRs in GI malignancies in the current research, including 50 exomiRs with high expression, 16 with low expression, and 4 with both low and high expression (miR-122, 638, 125b, 92a). All circulating exomiRs were categorized into two main subgroups as either up- or down-regulated exomiRs. According

to our findings, aberrant expression of exomiRs, in terms of both up and down-regulation, are strongly associated with inferior prognostic outcomes, including OS and DFS/RFS/PFS, as well as clinicopathological characteristics, including differentiation, TNM stage, lymph node metastasis, and distant metastasis. However, we did not identify any association between up or down-regulated circulating exomiRs and gender. Similar to our findings, a prior meta-analysis has demonstrated that aberrant exomiRs expression, in terms of both up and down-regulation, is correlated with a worse prognosis in colorectal cancer [23]. The above findings propose that developing the panel of exomiRs could be utilized as valuable biomarkers in GI cancer prognosis.

Among the various up-regulated miRs, miR-21 has been identified to be the most overexpressed miRNA in tumors, which may affect the development of cancer via different signaling cascades [94]. Overexpression of miR-21 was recognized as a prognostic and diagnostic biomarker in various cancers [95–99]. Furthermore, it was studied that high expression of miR-21 was correlated with low OS in glioma patients [100]. Our findings, which indicated a statistically significant correlation between the exomiR-21 and OS (HR=2.655,  $P<0.001$ ), DFS/RFS/PFS (HR=2.127,  $P<0.001$ ), and Lymph node metastasis (HR=1.568,  $P=0.009$ ), are consistent with these findings. Similarly, overexpression of miR-222 can enhance tumorigenesis, migration, and invasion properties in breast cancer (BC) and thyroid cancer [101, 102], along with its association with worse OS and DFS/RFS/PFS in glioma and NSCLC [103, 104]. Besides, high expression of the miR-200 family plays a significant role in tumorigenesis and metastasis in ovarian cancer and endometrial adenocarcinoma [105, 106] and is associated with shorter OS in breast cancer [107]. Regarding the results of our meta-analysis and other investigations, overexpression of some specific exomiRs has the chance to become predictors of long-term survival and metastasis in the patients with GI cancers.

Regarding down-regulated miRs, a low level of miR-320 is correlated with advanced stage, LNM, and poor OS in BC patients [108]. It was reported that miR-34 is expressed at low levels in BC, NSCLC, and bladder cancer and its down-regulation is correlated with recurrence, metastasis, and poor survival outcomes [109–111]. Furthermore, the down-regulation of miR-30c was correlated with poor prognostic outcomes in patients with BC [112, 113]. Thus, our findings revealed that up- or down-regulated exomiRs can be considered new biomarkers in predicting clinical outcomes in GI cancers.

Moreover, the expression of some circulating exomiRs could be either increased or decreased based on the cancer type. Among them, miR-122, a tumor suppressor miR, could regulate metastasis of HCC [114], and circulating miR-122 was up-regulated in BC, NSCLC [115, 116], which is associated with distant metastasis and lowered OS and PFS, considered as a prognostic factor [115]. Other groups of researchers, however, have shown that miR-122 expression is down-regulated in various cancer cells, including bladder and colon tumors [105]. These investigations support our findings, which show that several circulating exomiRs (miR-122, 638, 125b, and 92a) are members of both expression-high and expression-low subgroups in GI cancers. Although our large meta-analysis sheds light on the pathological characteristics and prognostic potentials of GI circulating exomiRs in clinical practice, several limitations should be considered when interpreting the results of the current study. Firstly, while all relevant studies were included in this study, relatively high heterogeneity was observed when analyzing the overall findings for OS or DFS/RFS/PFS. Therefore, subgroup analysis was performed by considering some subcategories to find the factors that caused the heterogeneity. Regarding this consideration, we did not underestimate the variability in patients' clinicopathological characteristics. Therefore, variations in population and research methodology might affect the heterogeneity. Secondly, the publication bias was identified for the relationship between OS or DFS/RFS/PFS and circulating exomiRs with high expression, affecting the validity of prognostic findings. Third, additional bias may possibly have arisen from (i) the number of investigations for some GI cancers was insufficient; (ii) all articles published in English; and (iii) most of the included publications were performed in Asian ethnicity, possibly leading to selection bias. Finally, some statistical errors may affect the credibility of findings in our meta-analysis, including (i) the indirect estimation of HR and 95% CI via the Kaplan–Meier curves in some studies; and (ii) using univariate analysis information instead of multivariate analysis for some studies without providing the statistical methodology. Despite these limitations, this meta-analysis suggests that circulating exomiRs would be useful as new prognostic biomarkers in GI cancers. However, multi-parameter and large-scale clinical studies with a strong methodology are required to implement exomiRs as biomarkers in the prognosis of GI cancers robustly.

## Conclusions

In conclusion, our study demonstrated the widest meta-analysis conducted on the prognostic importance of circulating exomiRs in GI cancers. Our results indicated that up- and down-regulated circulating exomiRs might serve as effective indicators of inferior survival outcomes in patients with gastrointestinal malignancies. In addition, exomiR dysregulation is related to advanced clinical stage, poor differentiation, and tumor spread in GI carcinomas. The current review advocates using a combined panel of circulating exomiRs for better risk stratification and clinical outcomes prediction in GI cancer patients, which could compensate for the unreliability of individual exomiRs in estimating prognosis. We envision our results would bring the attention of researchers and clinicians to the significance of deregulated circulating exomiRs as prognostic biomarkers that may aid better prediction.

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12935-023-02851-8>.

**Additional file 1: Figure S1.** Forest plot of the association between NOS and overall survival (A), disease/relapse/progression-free survival (B).

**Additional file 2: Figure S2.** Forest plot of the association between exomiR-21 and overall survival (A), disease/relapse/progression-free survival (B), and clinicopathological characteristics in patients with GI cancers. Lymph node metastasis (C), Differentiation (D), Distant metastasis (E), TNM stage (F), Gender (G).

**Additional file 3: Table S1.** Search strategy of electronic databases.

**Additional file 4: Table S2.** Significantly dysregulated circulating exomiR(s) in GI cancer patients.

**Additional file 5: Table S3.** Summary of meta-analyses for the association between exomiRs deregulation and clinicopathologic features in patients with GI cancers.

## Acknowledgements

Not applicable.

## Author contributions

ZM, RGh, SB, MR, EGh, and FT contributed to the concept and study design. The search syntax was provided by KT. The search strategy was developed by EGh, and FT. Data screening and selecting phases were performed by EGh, FT, and MS. Quality assessment was performed by EGh, FT, and MR. Data extraction and preparing the draft of the manuscript were performed by EGh, FT, and MR. All extracted data were analyzed by MR. Moreover, WCSC critically revised and edited the manuscript. Besides, ZM, RG, and MR were responsible for reviewing the manuscript and editing the final manuscript. All authors read and approved the final manuscript.

## Funding

The funding source is a college institute Iran University of Medical Sciences (IUMS) and has sponsored the project financially and approved (Grant Number #30805).

## Availability of data and materials

All recorded data from the data extraction process were available on request to the extent that they were not included in the systematic review article.

## Declarations

### Ethics approval and consent to participate

As this systematic review was only based on published data already in the public domain, ethical approval is not required. The findings of this systematic review have been disseminated through an international peer-reviewed journal publication and several scientific conference presentations. To the best of our knowledge, there are no systematic reviews that have specifically looked at the frequency of exosome encapsulated miRNA(s) in the circulating blood of patients with GI cancers.

### Consent for publication

Not applicable.

### Competing interests

There are no competing interests that the authors declare.

### Author details

<sup>1</sup>Oncopathology Research Center, Iran University of Medical Sciences, Tehran, Iran. <sup>2</sup>Department of Molecular Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran. <sup>3</sup>Department of Clinical Oncology, Queen Elizabeth Hospital, Kowloon, Hong Kong Special Administrative Region, China. <sup>4</sup>Department of Biostatistics, School of Public Health, Iran University of Medical Sciences, Tehran, Iran. <sup>5</sup>Department of Molecular Medicine, Faculty of Advanced Technologies in Medicine, Iran University of Medical Sciences, Tehran, Iran.

Received: 5 July 2022 Accepted: 12 January 2023

Published online: 20 January 2023

## References

- Arnold M, Abnet CC, Neale RE, Vignat J, Giovannucci EL, McGlynn KA, et al. Global burden of 5 major types of gastrointestinal cancer. *Gastroenterology*. 2020;159(1):335–49.e15.
- Nagtegaal ID, Odze RD, Klimstra D, Paradis V, Rugge M, Schirmacher P, et al. The 2019 WHO classification of tumours of the digestive system. *Histopathology*. 2020;76(2):182–8.
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics 2022. *CA A Cancer J Clin*. 2022;72(1):7–33.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA A Cancer J Clin*. 2021;71(3):209–49.
- Arnold M, Abnet CC, Neale RE, Vignat J, Giovannucci EL, McGlynn KA, et al. Global burden of 5 major types of gastrointestinal cancer. *Gastroenterology*. 2020;159(1):335–49.
- Bijlsma MF, Sadanandam A, Tan P, Vermeulen L. Molecular subtypes in cancers of the gastrointestinal tract. *Nat Rev Gastroenterol Hepatol*. 2017;14(6):333–42.
- Abyadeh M, Meyfour A, Gupta V, Zabet Moghaddam M, Fitzhenry MJ, Shahbazian S, et al. Recent advances of functional proteomics in gastrointestinal cancers- a path towards the identification of candidate diagnostic, prognostic, and therapeutic molecular biomarkers. *Int J Mol Sci*. 2020;21:22.
- Melton SD, Genta RM, Souza RF. Biomarkers and molecular diagnosis of gastrointestinal and pancreatic neoplasms. *Nat Rev Gastroenterol Hepatol*. 2010;7(11):620–8.
- Yamamoto Y, Kikuchi D, Nagami Y, Nonaka K, Tsuji Y, Fujimoto A, et al. Management of adverse events related to endoscopic resection of upper gastrointestinal neoplasms: review of the literature and recommendations from experts. *Dig Endosc*. 2019;31:4–20.
- Alves dos Santos K, Clemente dos Santos IC, Santos Silva C, Gomes Ribeiro H, de Farias Domingos I, Nogueira Silbiger V. Circulating exosomal miRNAs as biomarkers for the diagnosis and prognosis of colorectal cancer. *Int J Mol Sci*. 2021;22(1):346.
- Danac JMC, Uy AGG, Garcia RL. Exosomal microRNAs in colorectal cancer: overcoming barriers of the metastatic cascade. *Int J Mol Med*. 2021;47(6):1–16.
- Francavilla A, Turoczi S, Tarallo S, Vodicka P, Pardini B, Naccarati A. Exosomal microRNAs and other non-coding RNAs as colorectal cancer biomarkers: a review. *Mutagenesis*. 2020;35(3):243–60.
- Li X, Jiang W, Gan Y, Zhou W. The application of exosomal microRNAs in the treatment of pancreatic cancer and its research progress. *Pancreas*. 2021;50(1):12–6.
- Wang J, Yan F, Zhao Q, Zhan F, Wang R, Wang L, et al. Circulating exosomal miR-125a-3p as a novel biomarker for early-stage colon cancer. *Sci Rep*. 2017;7(1):1–9.
- Hayashi T, Lombaert IM, Hauser BR, Patel VN, Hoffman MP. Exosomal MicroRNA transport from salivary mesenchyme regulates epithelial progenitor expansion during organogenesis. *Dev Cell*. 2017;40(1):95–103.
- Ghosh S, Bhowmik S, Majumdar S, Goswami A, Chakraborty J, Gupta S, et al. The exosome encapsulated microRNAs as circulating diagnostic marker for hepatocellular carcinoma with low alpha-fetoprotein. *Int J Cancer*. 2020;147(10):2934–47.
- Wu L, Zhou WB, Zhou J, Wei Y, Wang HM, Liu XD, et al. Circulating exosomal microRNAs as novel potential detection biomarkers in pancreatic cancer. *Oncol Lett*. 2020;20(2):1432–40.
- Xu H, Ling M, Xue J, Dai X, Sun Q, Chen C, et al. Exosomal microRNA-21 derived from bronchial epithelial cells is involved in aberrant epithelium-fibroblast cross-talk in COPD induced by cigarette smoking. *Theranostics*. 2018;8(19):5419–33.
- Inamura K, Ishikawa Y. MicroRNA in lung cancer: novel biomarkers and potential tools for treatment. *J Clin Med*. 2016;5:3.
- Su Z, Ni L, Yu W, Yu Z, Chen D, Zhang E, et al. MicroRNA-451a is associated with cell proliferation, migration and apoptosis in renal cell carcinoma. *Mol Med Rep*. 2015;11(3):2248–54.
- Mahabady MK, Mirzaei S, Saebfar H, Gholami MH, Zabolian A, Hushmandi K, et al. Noncoding RNAs and their therapeutics in paclitaxel chemotherapy: mechanisms of initiation, progression, and drug sensitivity. *J Cell Physiol*. 2022;237(5):2309–44.
- Lamichhane SR, Thachil T, Gee H, Millic N. Circulating microRNAs as prognostic molecular biomarkers in human head and neck cancer: a systematic review and meta-analysis. *Dis Markers*. 2019. <https://doi.org/10.1155/2019/8632018>.
- Zhou J, Guo H, Yang Y, Zhang Y, Liu H. A meta-analysis on the prognosis of exosomal miRNAs in all solid tumor patients. *Medicine*. 2019;98:16.
- Liu Q, Yu Z, Yuan S, Xie W, Li C, Hu Z, et al. Circulating exosomal microRNAs as prognostic biomarkers for non-small-cell lung cancer. *Oncotarget*. 2017;8(8):13048.
- Yi R, Li Y, Wang FL, Miao G, Qi RM, Zhao YY. MicroRNAs as diagnostic and prognostic biomarkers in colorectal cancer. *World J Gastrointest Oncol*. 2016;8(4):330–40.
- Lan F, Qing Q, Pan Q, Hu M, Yu H, Yue X. Serum exosomal miR-301a as a potential diagnostic and prognostic biomarker for human glioma. *Cell Oncol*. 2018;41(1):25–33.
- Tanaka Y, Kamohara H, Kinoshita K, Kurashige J, Ishimoto T, Iwatsuki M, et al. Clinical impact of serum exosomal microRNA-21 as a clinical biomarker in human esophageal squamous cell carcinoma. *Cancer*. 2013;119(6):1159–67.
- Sugimachi K, Matsumura T, Hirata H, Uchi R, Ueda M, Ueo H, et al. Identification of a bona fide microRNA biomarker in serum exosomes that predicts hepatocellular carcinoma recurrence after liver transplantation. *Br J Cancer*. 2015;112(3):532–8.
- Gheytauchi E, Madjd Z, Janani L, Rasti A, Ghods R, Atyabi F, et al. Exosomal microRNAs as potential circulating biomarkers in gastrointestinal tract cancers: a systematic review protocol. *Syst Rev*. 2017;6(1):1–6.
- Munn Z, Stern C, Aromataris E, Lockwood C, Jordan Z. What kind of systematic review should I conduct? a proposed typology and guidance for systematic reviewers in the medical and health sciences. *BMC Med Res Methodol*. 2018;18(1):5.
- Page MJ, Moher D, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. PRISMA 2020 explanation and elaboration: updated guidance and exemplars for reporting systematic reviews. *BMJ*. 2021;372:n160.
- Stang A. Critical evaluation of the newcastle-ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol*. 2010;25(9):603–5.
- Parmar MK, Torri V, Stewart L. Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. *Stat Med*. 1998;17(24):2815–34.

34. Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315(7109):629–34.
35. Soeda N, Iinuma H, Suzuki Y, Tsukahara D, Midorikawa H, Igarashi Y, et al. Plasma exosome-encapsulated microRNA-21 and microRNA-92a are promising biomarkers for the prediction of peritoneal recurrence in patients with gastric cancer. *Oncol Lett*. 2019;18(5):4467–80.
36. Sun L, Liu X, Pan B, Hu X, Zhu Y, Su Y, et al. Serum exosomal miR-122 as a potential diagnostic and prognostic biomarker of colorectal cancer with liver metastasis. *J Cancer*. 2020;11(3):630–7.
37. Liu X, Pan B, Sun L, Chen X, Zeng K, Hu X, et al. Circulating exosomal miR-27a and miR-130a act as novel diagnostic and prognostic biomarkers of colorectal cancer. *Cancer Epidemiol Biomarkers Prev*. 2018;27(7):746–54.
38. Wu L, Zhou WB, Zhou J, Wei Y, Wang HM, Liu XD, et al. Circulating exosomal microRNAs as novel potential detection biomarkers in pancreatic cancer. *Oncol Lett*. 2020;20(2):1432–40.
39. Takahasi K, Iinuma H, Wada K, Minezaki S, Kawamura S, Kainuma M, et al. Usefulness of exosome-encapsulated microRNA-451a as a minimally invasive biomarker for prediction of recurrence and prognosis in pancreatic ductal adenocarcinoma. *J Hepatobiliary Pancreat Sci*. 2018;25(2):155–61.
40. Tsukamoto M, Iinuma H, Yagi T, Matsuda K, Hashiguchi Y. Circulating exosomal MicroRNA-21 as a biomarker in each tumor stage of colorectal cancer. *Oncology*. 2017;92(6):360–70.
41. Yokota Y, Noda T, Okumura Y, Kobayashi S, Iwagami Y, Yamada D, et al. Serum exosomal miR-638 is a prognostic marker of HCC via down-regulation of VE-cadherin and ZO-1 of endothelial cells. *Cancer Sci*. 2021;112(3):1275–88.
42. Takano Y, Masuda T, Iinuma H, Yamaguchi R, Sato K, Tobo T, et al. Circulating exosomal microRNA-203 is associated with metastasis possibly via inducing tumor-associated macrophages in colorectal cancer. *Oncotarget*. 2017;8(45):78598–613.
43. Tian XP, Wang CY, Jin XH, Li M, Wang FW, Huang WJ, et al. Acidic micro-environment up-regulates exosomal miR-21 and miR-10b in early-stage hepatocellular carcinoma to promote cancer cell proliferation and metastasis. *Theranostics*. 2019;9(7):1965–79.
44. Wei S, Peng L, Yang J, Sang H, Jin D, Li X, et al. Exosomal transfer of miR-15b-3p enhances tumorigenesis and malignant transformation through the DYNLT1/caspase-3/caspase-9 signaling pathway in gastric cancer. *J Exp Clin Cancer Res*. 2020;39(1):32.
45. Xue XF, Zhao YB, Wang XN, Qin L, Hu RK. Development and validation of serum exosomal microRNAs as diagnostic and prognostic biomarkers for hepatocellular carcinoma. *J Cell Biochem*. 2019;120(1):135–42.
46. Yagi T, Iinuma H, Hayama T, Matsuda K, Nozawa K, Tsukamoto M, et al. Plasma exosomal microRNA-125b as a monitoring biomarker of resistance to mFOLFFOX6-based chemotherapy in advanced and recurrent colorectal cancer patients. *Mol Clin Oncol*. 2019;11(4):416–24.
47. Yan SS, Jiang Y, Liang CH, Cheng M, Jin CW, Duan QH, et al. Exosomal miR-6803-5p as potential diagnostic and prognostic marker in colorectal cancer. *J Cell Biochem*. 2018;119(5):4113–9.
48. Matsumura T, Sugimachi K, Iinuma H, Takahashi Y, Kurashige J, Sawada G, et al. Exosomal microRNA in serum is a novel biomarker of recurrence in human colorectal cancer. *Br J Cancer*. 2015;113(2):275–81.
49. Kawamura S, Iinuma H, Wada K, Takahashi K, Minezaki S, Kainuma M, et al. Exosome-encapsulated microRNA-4525, microRNA-451a and microRNA-21 in portal vein blood is a high-sensitive liquid biomarker for the selection of high-risk pancreatic ductal adenocarcinoma patients. *J Hepatobiliary Pancreat Sci*. 2019;26(2):63–72.
50. Abd El Gwad A, Matboli M, El-Tawdi A, Habib EK, Shehata H, Ibrahim D, et al. Role of exosomal competing endogenous RNA in patients with hepatocellular carcinoma. *J Cell Biochem*. 2018;119(10):8600–10.
51. Cui Y, Xu HF, Liu MY, Xu YJ, He JC, Zhou Y, et al. Mechanism of exosomal microRNA-224 in development of hepatocellular carcinoma and its diagnostic and prognostic value. *World J Gastroenterol*. 2019;25(15):1890–8.
52. Feng C, She JJ, Chen XB, Zhang QC, Zhang X, Wang YS, et al. Exosomal miR-196a-1 promotes gastric cancer cell invasion and metastasis by targeting SFRP1. *Nanomedicine*. 2019;14(19):2579–93.
53. Liu WB, Chen S, Liu B. Diagnostic and prognostic values of serum exosomal microRNA-21 in children with hepatoblastoma: a Chinese population-based study. *Pediatr Surg Int*. 2016;32(11):1059–65.
54. Cho HJ, Baek GO, Seo CW, Ahn HR, Sung S, Son JA, et al. Exosomal microRNA-4661-5p-based serum panel as a potential diagnostic biomarker for early-stage hepatocellular carcinoma. *Cancer Med*. 2020;9(15):5459–72.
55. Han JY, Sun W, Liu R, Zhou Z, Zhang HY, Chen X, et al. Plasma exosomal miRNA expression profile as oxaliplatin-based chemoresistant biomarkers in colorectal adenocarcinoma. *Front Oncol*. 2020. <https://doi.org/10.3389/fonc.2020.01495>.
56. de Miguel PD, Rodríguez Martínez A, Ortigosa Palomo A, Delgado Ureña M, García Puche JL, Robles Remacho A, et al. Extracellular vesicle-miRNAs as liquid biopsy biomarkers for disease identification and prognosis in metastatic colorectal cancer patients. *Sci Rep*. 2020;10(1):3974.
57. Qu Z, Wu J, Wu J, Ji A, Qiang G, Jiang Y, et al. Exosomal miR-665 as a novel minimally invasive biomarker for hepatocellular carcinoma diagnosis and prognosis. *Oncotarget*. 2017;8(46):80666–78.
58. Reese M, Flammang I, Yang Z, Dhayat SA. Potential of exosomal microRNA-200b as liquid biopsy marker in pancreatic ductal adenocarcinoma. *Cancers*. 2020;12:1.
59. Meltzer S, Bjørntrø T, Lyckander LG, Flatmark K, Dueland S, Samiappan R, et al. Circulating exosomal miR-141-3p and miR-375 in metastatic progression of rectal cancer. *Transl Oncol*. 2019;12(8):1038–44.
60. Zhang YF, Huang FC, Xu N, Wang J, Li D, Yin L. Overexpression of serum extracellular vesicle micro-RNA-215-5p is associated with early tumor recurrence and poor prognosis of gastric cancer. *Clinics*. 2021. <https://doi.org/10.6061/clinics/2021/e2081>.
61. Zhu LY, Zhang SS, Chen SD, Wu HJ, Jiang MJ, Liu AQ. Exosomal miR-552-5p promotes tumorigenesis and disease progression via the PTEN/TOB1 axis in gastric cancer. *J Cancer*. 2022;13(3):890–905.
62. Yang JJ, Li X, Wei SC, Peng L, Sang HM, Jin DC, et al. Evaluation of the diagnostic potential of a plasma exosomal miRNAs panel for gastric cancer. *Front Oncol*. 2021. <https://doi.org/10.3389/fonc.2021.683465>.
63. Chen S, Mao Y, Chen W, Liu C, Wu H, Zhang J, et al. Serum exosomal miR-34a as a potential biomarker for the diagnosis and prognostic of hepatocellular carcinoma. *J Cancer*. 2022;13(5):1410–7.
64. Huang CY, Tang SY, Shen D, Li XZ, Liang L, Ding YQ, et al. Circulating plasma exosomal miRNA profiles serve as potential metastasis-related biomarkers for hepatocellular carcinoma. *Oncol Lett*. 2021;21:2.
65. Hao YJ, Yang CY, Chen MH, Chang LW, Lin CP, Lo LC, et al. Potential values of circulating microRNA-21 to predict early recurrence in patients with colorectal cancer after treatments. *J Clin Med*. 2022;11:9.
66. Yan S, Dang G, Zhang X, Jin C, Qin L, Wang Y, et al. Downregulation of circulating exosomal miR-638 predicts poor prognosis in colon cancer patients. *Oncotarget*. 2017;8(42):72220–6.
67. Yan SS, Liu GY, Jin CW, Wang ZF, Duan QH, Xu J, et al. MicroRNA-6869-5p acts as a tumor suppressor via targeting TLR4/NF- $\kappa$ B signaling pathway in colorectal cancer. *J Cell Physiol*. 2018;233(9):6660–8.
68. Zou SL, Chen YL, Ge ZZ, Qu YY, Cao Y, Kang ZX. Downregulation of serum exosomal miR-150-5p is associated with poor prognosis in patients with colorectal cancer. *Cancer Biomarker*. 2019;26(1):69–77.
69. Zhang N, Zhang PP, Huang JJ, Wang ZY, Zhang ZH, Yuan JZ, et al. Reduced serum exosomal miR-874 expression predicts poor prognosis in colorectal cancer. *Eur Rev Med Pharmacol Sci*. 2020;24(2):664–72.
70. Liu C, Eng C, Shen J, Lu Y, Takata Y, Mehdi zadeh A, et al. Serum exosomal miR-4772-3p is a predictor of tumor recurrence in stage II and III colon cancer. *Oncotarget*. 2016;7(46):76250–60.
71. Hao X, Xin R, Dong W. Decreased serum exosomal miR-320a expression is an unfavorable prognostic factor in patients with hepatocellular carcinoma. *J Int Med Res*. 2020;48(4):300060519896144.
72. Jiao CW, Jiao XH, Zhu AZ, Ge JT, Xu XQ. Exosomal miR-34s panel as potential novel diagnostic and prognostic biomarker in patients with hepatoblastoma. *J Pediatr Surg*. 2017;52(4):618–24.
73. Li W, Ding X, Wang S, Xu L, Yin T, Han S, et al. Downregulation of serum exosomal miR-320d predicts poor prognosis in hepatocellular carcinoma. *J Clin Lab Anal*. 2020;34(6):e23239.
74. Liu WF, Hu J, Zhou KQ, Chen FY, Wang Z, Liao BY, et al. Serum exosomal miR-125b is a novel prognostic marker for hepatocellular carcinoma. *Onco Targets Ther*. 2017;10:3843–51.

75. Sheng LQ, Li JR, Qin H, Liu L, Zhang D, Zhang Q, et al. Blood exosomal micro ribonucleic acid profiling reveals the complexity of hepatocellular carcinoma and identifies potential biomarkers for differential diagnosis. *World J Gastrointest Oncol.* 2020;12(10):1195–208.
76. Suehiro T, Miyaaki H, Kanda Y, Shibata H, Honda T, Ozawa E, et al. Serum exosomal microRNA-122 and microRNA-21 as predictive biomarkers in transarterial chemoembolization-treated hepatocellular carcinoma patients. *Oncol Lett.* 2018;16(3):3267–73.
77. Kumata Y, Iinuma H, Suzuki Y, Tsukahara D, Midorikawa H, Igarashi Y, et al. Exosome-encapsulated microRNA-23b as a minimally invasive liquid biomarker for the prediction of recurrence and prognosis of gastric cancer patients in each tumor stage. *Oncol Rep.* 2018;40(1):319–30.
78. Peng ZY, Gu RH, Yan B. Downregulation of exosome-encapsulated miR-548c-5p is associated with poor prognosis in colorectal cancer. *J Cell Biochem.* 2018. <https://doi.org/10.1002/jcb.27291>.
79. Zheng GD, Xu ZY, Hu C, Lv H, Xie HX, Huang T, et al. Exosomal miR-590-5p in serum as a biomarker for the diagnosis and prognosis of gastric cancer. *Front Mol Biosci.* 2021;8:636566.
80. Peng Y, Croce CM. The role of MicroRNAs in human cancer. *Signal Transduct Target Ther.* 2016;1:15004.
81. Nik Mohamed Kamal N, Shahidan WNS. Non-exosomal and exosomal circulatory micromas: which are more valid as biomarkers? *Front Pharmacol.* 2019;10:1500.
82. Ogata-Kawata H, Izumiya M, Kurioka D, Honma Y, Yamada Y, Furuta K, et al. Circulating exosomal microRNAs as biomarkers of colon cancer. *PLoS ONE.* 2014;9(4):e92921.
83. Lin J, Li J, Huang B, Liu J, Chen X, Chen XM, et al. Exosomes: novel biomarkers for clinical diagnosis. *ScientificWorldJournal.* 2015;2015:657086.
84. Zöller M. Pancreatic cancer diagnosis by free and exosomal miRNA. *World J Gastrointest Pathophysiol.* 2013;4(4):74–90.
85. Que R, Ding G, Chen J, Cao L. Analysis of serum exosomal microRNAs and clinicopathologic features of patients with pancreatic adenocarcinoma. *World J Surg Oncol.* 2013;11:219.
86. Tanaka Y, Kamohara H, Kinoshita K, Kurashige J, Ishimoto T, Iwatsuki M, et al. Clinical impact of serum exosomal microRNA-21 as a clinical biomarker in human esophageal squamous cell carcinoma. *Cancer.* 2013;119(6):1159–67.
87. Sohn W, Kim J, Kang SH, Yang SR, Cho JY, Cho HC, et al. Serum exosomal microRNAs as novel biomarkers for hepatocellular carcinoma. *Exp Mol Med.* 2015;47(9):e184.
88. Xiao D, Ohlendorf J, Chen Y, Taylor DD, Rai SN, Waigel S, et al. Identifying mRNA, microRNA and protein profiles of melanoma exosomes. *PLoS ONE.* 2012;7(10):e46874.
89. Tokunaga M, Kurokawa Y, Machida R, Sato Y, Takiguchi S, Doki Y, et al. Impact of postoperative complications on survival outcomes in patients with gastric cancer: exploratory analysis of a randomized controlled JCOG1001 trial. *Gastric Cancer.* 2021;24(1):214–23.
90. Osseis M, Esposito F, Lim C, Doussot A, Lahat E, Fuentes L, et al. Impact of postoperative complications on long-term survival following surgery for T4 colorectal cancer. *BMC Surg.* 2018;18(1):87.
91. Kokelaar RF, Evans MD, Davies M, Harris DA, Beynon J. Locally advanced rectal cancer: management challenges. *Onco Targets Ther.* 2016;9:6265–72.
92. Apffelstaedt JP, Driscoll DL, Spellman JE, Velez AF, Gibbs JF, Karakousis CP. Complications and outcome of external hemipelvectomy in the management of pelvic tumors. *Ann Surg Oncol.* 1996;3(3):304–9.
93. Nacion AJD, Park YY, Kim NK. Contemporary management of locally advanced rectal cancer: resolving issues, controversies and shifting paradigms. *Chin J Cancer Res.* 2018;30(1):131–46.
94. Chan JA, Krichevsky AM, Kosik KS. MicroRNA-21 is an antiapoptotic factor in human glioblastoma cells. *Cancer Res.* 2005;65(14):6029–33.
95. Sun M, Song J, Zhou Z, Zhu R, Jin H, Ji Y, et al. Comparison of serum microRNA21 and Tumor markers in diagnosis of early non-small cell lung cancer. *Dis Markers.* 2016;2016:3823121.
96. Feng YH, Tsao CJ. Emerging role of microRNA-21 in cancer. *Biomed Rep.* 2016;5(4):395–402.
97. Chen J, Wang X. MicroRNA-21 in breast cancer: diagnostic and prognostic potential. *Clin Transl Oncol.* 2014;16(3):225–33.
98. Qu J, Yang J, Chen M, Cui L, Wang T, Gao W, et al. MicroRNA-21 as a diagnostic marker for hepatocellular carcinoma: A systematic review and meta-analysis. *Pak J Med Sci.* 2019;35(5):1466–71.
99. Wang Y, Xing QF, Liu XQ, Guo ZJ, Li CY, Sun G. MiR-122 targets VEGFC in bladder cancer to inhibit tumor growth and angiogenesis. *Am J Transl Res.* 2016;8(7):3056–66.
100. Li C, Sun J, Xiang Q, Liang Y, Zhao N, Zhang Z, et al. Prognostic role of microRNA-21 expression in gliomas: a meta-analysis. *J Neurooncol.* 2016;130(1):11–7.
101. Chatterjee A, Jana S, Chatterjee S, Wastall LM, Mandal G, Nargis N, et al. MicroRNA-222 reprogrammed cancer-associated fibroblasts enhance growth and metastasis of breast cancer. *Br J Cancer.* 2019;121(8):679–89.
102. Huang Y, Yu S, Cao S, Yin Y, Hong S, Guan H, et al. MicroRNA-222 promotes invasion and metastasis of papillary thyroid cancer through targeting protein phosphatase 2 regulatory subunit B alpha expression. *Thyroid.* 2018;28(9):1162–73.
103. Zhang R, Pang B, Xin T, Guo H, Xing Y, Xu S, et al. Plasma miR-221/222 family as novel descriptive and prognostic biomarkers for glioma. *Mol Neurobiol.* 2016;53(3):1452–60.
104. Ulivi P, Petracci E, Marisi G, Baglivo S, Chiari R, Billi M, et al. Prognostic role of circulating miRNAs in early-stage non-small cell lung cancer. *J Clin Med.* 2019;8:2.
105. Choi PW, Ng SW. The functions of microRNA-200 family in ovarian cancer: beyond epithelial-mesenchymal transition. *Int J Mol Sci.* 2017;18:6.
106. Snowdon J, Zhang X, Childs T, Tron VA, Feilotter H. The microRNA-200 family is up-regulated in endometrial carcinoma. *PLoS ONE.* 2011;6(8):e22828.
107. Thi Chung Duong T, Nguyen THN, Thi Ngoc Nguyen T, Huynh LH, Ngo HP, Thi Nguyen H. Diagnostic and prognostic value of miR-200 family in breast cancer: a meta-analysis and systematic review. *Cancer Epidemiol.* 2022;77:102097.
108. Luo L, Yang R, Zhao S, Chen Y, Hong S, Wang K, et al. Decreased miR-320 expression is associated with breast cancer progression, cell migration, and invasiveness via targeting aquaporin 1. *Acta Biochim Biophys Sin.* 2018;50(5):473–80.
109. Zhang L, Wang L, Dong D, Wang Z, Ji W, Yu M, et al. MiR-34b/c-5p and the neurokinin-1 receptor regulate breast cancer cell proliferation and apoptosis. *Cell Prolif.* 2019;52(1):e12527.
110. Tafsiiri E, Darbouy M, Shadmehrb MB, Zagryazhskaya A, Alizadeh J, Karimipour M. Expression of miRNAs in non-small-cell lung carcinomas and their association with clinicopathological features. *Tumour Biol.* 2015;36(3):1603–12.
111. Juracek J, Stanik M, Vesela P, Radova L, Dolezel J, Svoboda M, et al. Tumor expression of miR-34a-3p is an independent predictor of recurrence in non-muscle-invasive bladder cancer and promising additional factor to improve predictive value of EORTC nomogram. *Urol Oncol.* 2019;37(3):184.e1-7.
112. Rodríguez-González FG, Sieuwerts AM, Smid M, Look MP, Meijer-van Gelder ME, de Weerd V, et al. MicroRNA-30c expression level is an independent predictor of clinical benefit of endocrine therapy in advanced estrogen receptor positive breast cancer. *Breast Cancer Res Treat.* 2011;127(1):43–51.
113. Han W, Cui H, Liang J, Su X. Role of microRNA-30c in cancer progression. *J Cancer.* 2020;11(9):2593–601.
114. Tsai WC, Hsu PW, Lai TC, Chau GY, Lin CW, Chen CM, et al. MicroRNA-122, a tumor suppressor microRNA that regulates intrahepatic metastasis of hepatocellular carcinoma. *Hepatology.* 2009;49(5):1571–82.
115. Saleh AA, Soliman SE, Habib MSE, Gohar SF, Abo-Zeid GS. Potential value of circulatory microRNA122 gene expression as a prognostic and metastatic prediction marker for breast cancer. *Mol Biol Rep.* 2019;46(3):2809–18.
116. Fan X, Wu X. MicroRNA-122-5p promotes the development of non-small cell lung cancer via downregulating p53 and activating PI3K-AKT pathway. *J Buon.* 2019;24(1):273–9.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.