REVIEW



The renin-angiotensin-aldosterone system (RAAS) signaling pathways and cancer: foes versus allies



Bahareh Hassani^{1†}, Zeinab Attar^{2†} and Negar Firouzabadi^{3*}

Abstract

The renin-angiotensin-aldosterone system (RAAS), is an old system with new fundamental roles in cancer biology which influences cell growth, migration, death, and metastasis. RAAS signaling enhances cell proliferation in malignancy directly and indirectly by affecting tumor cells and modulating angiogenesis. Cancer development may be influenced by the balance between the ACE/Ang II/AT1R and the ACE2/Ang 1–7/Mas receptor pathways. The interactions between Ang II/AT1R and Ang I/AT2R as well as Ang1-7/Mas and alamandine/MrgD receptors in the RAAS pathway can significantly impact the development of cancer. Ang I/AT2R, Ang1-7/Mas, and alamandine/MrgD interactions can have anticancer effects while Ang II/AT1R interactions can be involved in the development of cancer. Evidence suggests that inhibitors of the RAAS, which are conventionally used to treat cardiovascular diseases, may be beneficial in cancer therapies.

Herein, we aim to provide a thorough description of the elements of RAAS and their molecular play in cancer. Alongside this, the role of RAAS components in sex-dependent cancers as well as GI cancers will be discussed with the hope of enlightening new venues for adjuvant cancer treatment.

Keywords Renin-angiotensin-aldosterone system, Cancer biology, ACE inhibitors, ARBs, sex hormone-dependent cancer, GI cancer

Introduction

Cancer is the greatest clinical, social, and economic burden measured in cause-specific Disability-Adjusted Life Years (DALYs). The accelerating prevalence of cancer and the rising expense of cancer therapy highlight the urgent need for more efficient and cost-effective therapies [1]. Nearly every organ in the human body expresses functional RAAS at various levels [2]. The relationship

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changes in the function and activity of RAAS elements
play various roles in carcinogenesis [4]. Numerous tumor
types, including sex hormone-dependent cancers such
as breast, prostate, cervical and ovarian cancers as well
as GI cancers like pancreatic, stomach, liver and colon
cancer along the cancers of the brain, lung, skin, and
hematopoietic cells have been shown to exhibit increased
RAAS activity [5, 6]. The involvement of RAAS in cell
proliferation, inflammation, metastasis, and angiogenesis has recently received great attention. Hence, applying therapeutic options which target the RAAS pathways

between RAAS signaling and cancer has recently been

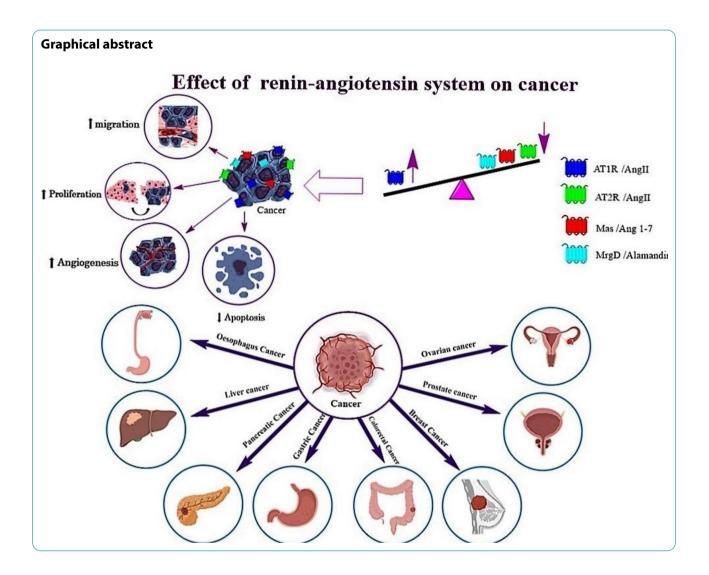
highlighted. RAAS signaling is a key player in maintain-

ing many physiological conditions [3]. Dysregulation of

RAAS signaling frequently happens in cancer. Indeed,



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might inhibit tumor growth and metastasis, and increase overall survival.

Receptors and effectors in the RAAS pathway can have opposite effects in preventing or developing cancer. Therefore, a better understanding of the RAAS signaling pathway and the effective factors and drugs affecting each of the receptors can lead to new therapeutic options. RAAS proteins are small peptides with opposing effects on cancerous cells. Angiotensin II (Ang II), the major peptide of the classical axis is a product of the angiotensin-converting enzyme (ACE). It has various effects on affecting AT1R and AT2R. Ang II enhances cell proliferation and angiogenesis through the signaling of AT1 receptor, whereas Ang II induces anti-proliferative effects through AT2 receptor [7]. Angiogenesis, fibrosis, increased tumor invasion, and metastasis can all be attributed to the binding of Ang II to AT1R. Contrarily, when Ang II binds to AT2R, it causes opposite effects that may prevent cancer [8]. The ACE2/Ang1-7/Mas axis is described as the counter-regulatory axis of the classical ACE/Ang II/AT1R axis. Ang 1–7, the peptide produced by ACE2, generally inhibits cell proliferation via Mas receptors [9]. Ang 1–7/Mas axis along with the Ang II/ AT2R axis are antagonists of the classic axis of ACE/ Ang II/AT1R, particularly under pathological conditions [10]. Ang 1–7 has anti-inflammatory, anti-fibrotic, antiproliferative, and anti-migratory effects when it binds to the Mas receptor [8]. Therefore, RAAS pathway can have dualistic effects.

Here we emphasized on the dualistic role of RAAS in cancer. We discussed the elements of the RAAS pathways and their role in sex-hormone-dependent cancers (breast, ovary, and prostate) as well as common GI cancers. Moreover, drugs affecting RAAS along with their potential role in the above-mentioned malignancies are discussed.

The RAAS pathway

Along with the prominent function of the renin-angiotensin-aldosterone system as the most important blood pressure regulator, RAAS could be related to different pathological conditions such as cancer and oncogenesis [11]. RAAS is mainly regulated by the heart and kidney, and its main function is to maintain body homeostasis and tissue perfusion. This cooperation is done in such a way that allows the kidney to maintain blood volume and the body's water content whereas the cardiovascular system supplies blood circulation [12]. Angiotensin II (Ang II), a potent vasoconstrictor, is produced by the RAAS pathway with the aid of a sequence of enzymatic processes. Researchers have discovered the existence of RAAS in various organs and tissues such as the gastrointestinal and nervous system, liver, muscle, bone, pancreas, and adipose tissue [13].

The knowledge of the traditional RAAS has progressed through identification of new actors, Angiotensin-converting enzyme 2 (ACE2), prorenin receptor (PRR), and the Mas receptor [14]. New perspectives on the physiology and pathology of the RAAS have been achieved by proposing the RAAS pathway as a two-axis system consisting of the Angiotensin-converting enzyme (ACE)/Ang II/Ang II type 1 receptor (AT1R) and ACE2/Angiotensin 1–7 (Ang 1–7)/Mas receptor [15, 16], along discovering PRR and its physiologic function [17, 18], as well as discovering angiotensin 1–9 (Ang 1–9) as an active protein in the RAAS [19, 20]. These new perspectives on the RAAS enable researchers to target RAAS to treat various diseases [14].

Angiotensinogen and angiotensin derivatives

Angiotensinogen can be considered the precursor of other Angs. In fact, angiotensinogen is a glycoprotein which is the starting point of the RAAS pathway. There are 485 amino acids in human angiotensinogen which include a signal peptide of 33 amino acids. Angiotensinogen belongs to the non-inhibitory serpin superfamily which are serine protease inhibitors [21]. Production and secretion of Ang occurs in the liver, but it should be noted that although the liver is the main producing organ, angiotensinogen is additionally produced in the brain, lungs, kidneys, vasculature, and heart [12]. Renin can cleave the N-terminal amino acids of angiotensinogen to generate the inactive decapeptide Ang I. Ang I can be considered a source of active Ang peptides. The ratelimiting step for the release of Ang I is the cleavage of angiotensinogen by renin [21]. Renin cleavage of angiotensinogen is different in various species [22, 23].

An overview of the angiotensin derivatives is displayed in Fig. 1. Ang I cleaves into an active octapeptide called Ang II. This cleavage is carried out by ACE or chymase. Ang 1–7 can be derived from Ang I or Ang II through endo-peptidases or carboxy-peptidases respectively. Ang I can be converted into Ang 1-7 by some peptidases for instance TOP (thimet oligopeptidase), NEP (neutral endopeptidase), and PEP (prolyl-endopeptidase). The Pro⁷-Phe⁸ linkage in Ang I can break and convert Ang I to Ang 1–7. Ang II is also able to be hydrolyzed by ACE2 to produce Ang 1–7 [24, 14]. In addition, Ang I can be cleaved by ACE2 to create Ang 1–9, which can be cleaved by ACE or NEP to generate Ang 1–7 [25, 14]. In other words, ACE2 can cut phenylalanine amino acid from Ang II and convert it to Ang 1–7. Furthermore, Ang I can be converted to Ang 1-9 by ACE2 by removing the amino acid leucine. Therefore, ACE2 appears to be crucial for the formation of Ang 1–7 and Ang 1–9 [26, 27]. Ang III (Ang 2-8) is another Ang derivative that is derived from Ang II by aminopeptidase A (APA). It has the ability to bind to AT1R and induce the same pressor response as Ang II, but with a more significant effect on the brain [28]. As Ang III is further cleaved, it produces Ang IV (Ang 3-8), which can interact with AT4R and promote vasodilation in the cerebral and renal vascular beds and facilitate sodium excretion and renal blood flow [29]. Another derivative of Ang is Ang A (Ang A) which is considered a physiologically active metabolite of Ang II. Ang A is an octapeptide whose amino acid sequence only differs by one amino acid from Ang II. Thus, in Ang A, there is an Ala¹ amino acid instead of Asp¹. Ang A probably generates from Ang II via decarboxylation of Asp¹. Ang A can interact with both types of AT1R and AT2R receptors [30]. Alamandine could be produced from Ang A by ACE2. Additionally, decarboxylation of the aspartate residue in Ang 1-7 can also generate alamandine directly. In fact, Alamandine simply varies from Ang 1–7 in that its N-terminal residue is an alanine rather than an aspartate [31]. Alamandine can exert its effects by interacting with Mas-related G protein-coupled receptor D (MrgD). Therefore, the Ang A/alamandine/MrgD signaling pathway emerges [32]. Research indicates that the function of various organs and systems could be related to the balance between the activation of the ACE/Ang II/AT1R and the ACE2/Ang 1-7/Mas receptor pathway. Factors that cause imbalance towards the ACE/Ang II/ AT1R pathway can result in cardiovascular diseases. It is worth mentioning that the function of this axis is beyond the cardio-renal and vascular due to its pleiotropic effects and can affect a wide range of diseases [33, 34, 35]. Inflammation, oxidative stress, fibrosis, and hypertrophy are associated with the activation of the ACE/Ang II/AT1R pathway [36].

Angiotensin 1-7

An active heptapeptide known as Ang 1-7 is another Ang derivative [36]. Ang 1-7 can interact with the endogenous Mas receptor which can antagonize the

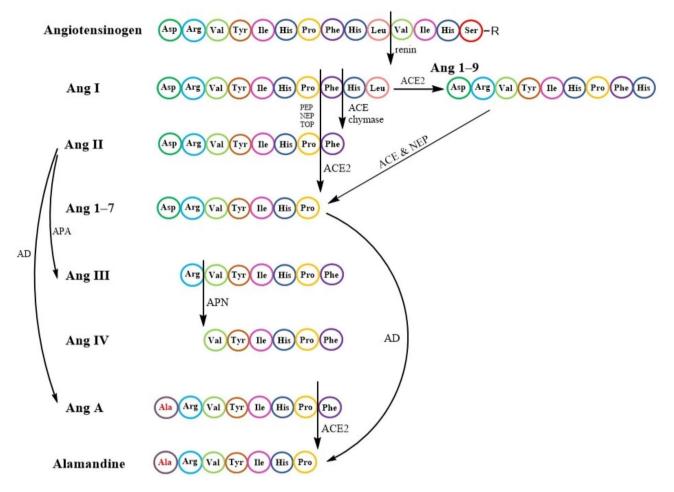


Fig. 1 RAAS peptides, receptors and enzymes

Angiotensinogen is the precursor of all angiotensin derivatives. Angiotensinogen can be cleaved by renin to produce Ang (I) Ang I cleaves into Ang II by ACE or chymase. Some peptidases, like TOP, NEP, and PEP, can convert Ang I into Ang 1–7. ACE2 can hydrolyze Ang II to create Ang 1–7 as well. In addition, Ang I can be cleaved by ACE2 to create Ang 1–9, which can be cleaved by ACE or NEP to generate Ang 1–7. Ang III (Ang 2–8) can be driven from Ang II by APA. As Ang III is further cleaved, it produces Ang IV (Ang 3–8). Through the decarboxylation of Asp1, Ang A is produced from Ang (II) Ang A could be converted to alamandine by ACE2. Alamandine can also be produced by decarboxylation of the aspartate residue in Ang 1–7

actions of Ang II and can counter-regulate most of Ang II's negative effects. Therefore, it has positive cardiovascular effects and is able to decrease blood pressure. In addition to its initially identified cardiovascular and renal functions, Ang 1–7 has different functions in various tissues and organs [37]. Studies have shown that the Mas antagonist, A779, can block most of Ang 1-7's actions [38, 39, 40, 41]. Furthermore, the effects of Ang 1–7 are not found in Mas-deficient animals [42, 40, 43, 44]. MrgD can also mediate some of the actions of Ang 1-7 [42, 40, 43, 44]. Therefore, it was concluded that the effect of Ang 1-7 can be through the Mas receptor. The Ang 1-7/Masreceptor interaction can regulate several signaling pathways, including the ERK and phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT) pathways, as well as affect downstream effectors like NO, cyclo-oxygenase-2 (COX-2) and forkhead box O1 (FOXO1) [14]. These mechanisms allow Ang 1-7 to improve pathological disorders such as inflammation and fibrosis in various organs like the liver, lungs, and kidneys. Due to its potential to prevent angiogenesis and cell proliferation, Ang 1–7 can be considered a potential anticancer therapy [14].

Renin and prorenin

Prorenin is a precursor of renin. This transformation occurs in juxtaglomerular granular cells of the renal macula densa. Juxtaglomerular granular cells are specialized smooth muscle cells found primarily in the walls of the renal afferent arterioles that are involved in the generation, storage, and release of renin [45]. Various conditions trigger renin secretion from the kidneys into the bloodstream among which a reduction in blood pressure is considered the most crucial factor. Other stimulators of renin secretion include sympathetic stimulators and humoral factors such as Ang II, vasopressin, NO, histamine, prostaglandins, endothelins, and dopamine [45]. 46]. In addition to the kidney, the uterus, placenta, testicles, adrenal gland, retina, and submandibular glands also produce renin [47].

ACE & ACE2

ACE is a dipeptide carboxypeptidase and is responsible for converting Ang I to Ang II. On the other hand, ACE2 is a carboxypeptidase that is responsible for converting Ang II to Ang 1–7 [59, 25]. Although ACE2 and ACE have considerable similarities, ACE inhibitors do not block ACE2 activity [34]. ACE2 is a transmembrane glycoprotein that has a mono-carboxy-peptidase function which has a catalytically active ectodomain. Although ACE2 is expressed in most tissues of the body, the highest expression of ACE2 is in the lungs, kidneys, heart, and endothelium [60, 61]. It can be considered that the major function of ACE2 is the production of Ang 1-7 from Ang II, which can antagonize the action of Ang II [34]. Although the main substrate of ACE2 is Ang II, other peptides with lower affinity can be degraded by ACE2 [61, 34]. ACE2 can lessen the deleterious effects of RAAS in a variety of ways. Firstly, it is responsible for the cleavage of Ang I and Ang II, therefore it can reduce the available substrate of the ACE/Ang II/AT1R pathway, and in addition, it is responsible for the production of Ang 1-7 so it can increase the available substrate of the protective ACE2/Ang 1–7/Mas receptor pathway [62].

AT1R & AT2R

AT1R and AT2R are G protein-coupled receptors and have pivotal roles in the RAAS pathway. Both receptors have a similar affinity for Ang II [63, 64]. While AT1R's activities have long been known, AT2R was only discovered recently, and many of its activities are still not completely understood [65, 66]. AT2R is effective in the protective arm of RAAS and has the potential to be targeted for drug development. It has beneficial effects in improving cardiovascular, renal, lung, neural, and cutaneous diseases in addition to cancer [67]. The maximum relative affinities for AT1R and AT2R are possessed by Ang II and Ang III, respectively [68]. Most of the harmful and negative effects of Ang II are through its effect on AT1R [69]. A variety of protective actions are mediated by AT2R in the pathophysiological conditions, which are very similar to the effects of Ang 1–7 via Mas receptor, and include immune modulation, anti-inflammatory, antiapoptotic, antifibrotic, neuro-regeneration actions and inhibition of sympathetic outflow [70, 36]. Activation of AT2R may inhibit cell proliferation, promote vasodilation, and reduce oxidative stress and inflammation [71]. The proper balance between AT1R and AT2R function has a significant effect on the physiological activities of major organs [72, 69].

Prorenin receptor (PRR)

PRR is a transmembrane protein with three domains: an extracellular domain with the N-terminal region, a transmembrane domain, and a cytoplasmic domain with the C-terminal region. [48]. It is commonly agreed that PRR is an essential receptor for regulating RAAS. The primary function of PRR is Ang II formation [49]. This receptor was known primarily for its ability to enhance RAAS by interacting with prorenin, and renin and also activating the intracellular mitogen-activated protein kinase (MAPK)/ extracellular signal-regulated kinase (ERK) pathway independently of the RAAS, thus having a crucial effect on renal and cardiovascular function [50]. PRRs are implicated in different physiologic and pathologic conditions and pathways, including Vacuolar ATPase (V-ATPase) activity [51], Wnt/β-catenin signaling [52], Par3 system, and tyrosine-phosphorylation-dependent signaling pathways [49]. Latest research has demonstrated that benign tumors and human cancers express PRR much more than normal tissues. It was found to be overexpressed in breast cancer [53], glioma [54], pancreatic ductal adenocarcinoma [55, 56], aldosterone-producing adenoma [57], and colorectal cancer [58].

Mas receptors

Mas receptor is a transmembrane protein and a member of G protein-coupled receptors [73]. There is a subtype of Mas receptor called Mas-Related G-Protein Coupled Receptor D (MrgD), with an affinity for alamandine [74]. Alamandine does not attach to the Mas receptor, however, it binds to the MrgD receptor with the same features [75]. Activation of the MAS receptor in the RAAS reduces chronic hypertension, fibrosis, blood pressure, and sympathetic tone and increases vasodilation, nitric oxide production, parasympathetic tone, baroreflex, and natriuresis [40]. Positive effects related to the Mas receptor are produced by its interaction with Ang 1–7 [76].

The RAAS pathway and cancer

Nearly every organ in the human body expresses a functional RAAS at various levels [2]. Numerous tumor types, including cancer of the breast, kidney, pancreatic, prostate, stomach, bladder, cervix, brain, lung, liver, colon, skin, and hematopoietic cells, have been shown to have increased RAAS activity [5, 6, 5]. The involvement of RAAS in cell proliferation, inflammation, metastasis, and angiogenesis has received more attention recently.

The relation between RAAS and cancer has recently been highlighted, and various researches have emphasized the role of RAAS dysregulation in cancer. It is noteworthy that different components of RAAS can have different roles in carcinogenesis [4]. (Fig. 2 displays an

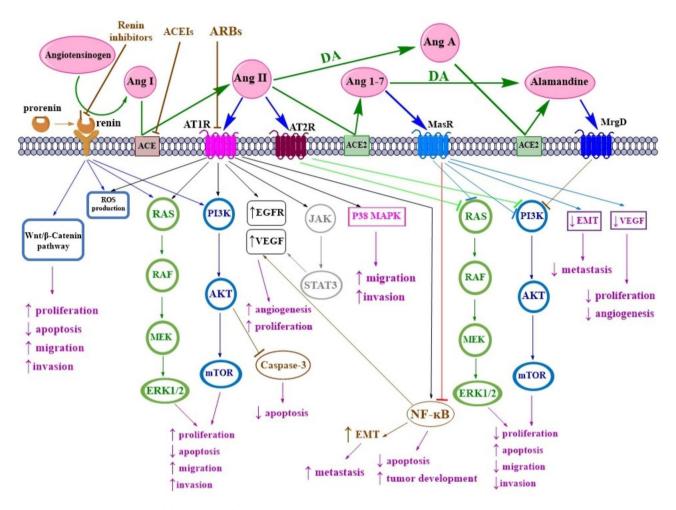


Fig. 2 Angiotensin derivatives and their associated receptors

The downstream pathways affecting proliferation, apoptosis, angiogenesis, and cancer as well as an overview of the receptors and effectors affecting the RAAS pathway is displayed and the dual effect of this pathway on cancer is depicted. The interactions between Ang II/AT1R and Ang I/AT2R and Ang1-7/ Mas and alamandine/MrgD receptors in the RAAS pathway can have a significant impact on the development of cancer. Ang I/AT2R, Ang1-7/Mas, and alamandine/MrgD interactions can have anticancer effects while Ang II/AT1R interactions can be involved in the development of cancer. Activation or inhibition of the PI3K/AKT/mTOR pathway and the RAS/RAF/ERK1/2 pathway, which are downstream of the RASS pathway, are the most effective pathways in cancer development or cancer prevention

overview of different receptors and effectors of the RAAS pathway and its downstream effectors)

Different types of receptors and effectors can affect RAAS regulation. PRR, AT1R, AT2R, and Mas receptor are among the receptors involved in the pathophysiology of RAAS, whose inhibition or activation can impact cancer progression. Among the effectors involved in this pathway, Ang II, Ang 1–7, and Ang 1–9 are of major concern [78].

Endothelial cells and fibroblasts, as well as other major components of the tumor microenvironment, together with tumor cells themselves, could be involved in producing and expressing RAAS components that promote angiogenesis [79]. The creation and secretion of cytokines (TNF, IL-1, and IL-6), growth factors (VEGF), and the production of ROS by neutrophils and macrophages in inflammatory and hypoxic environments are also facilitated by RAAS signaling pathways [80].

Ang II/AT1R Pathway

Along with the effects of AT1R on the regulation of blood pressure, it can also lead to enhanced proliferation, migration, motility, invasion, angiogenesis, and inhibition of apoptosis [81, 82]. It appears that the AT1R pathway seems to be the basic RAAS pathway that is implicated in tumor development, tumor proliferation, and angiogenesis by increasing the expression of epidermal growth factor receptor (EGFR) or vascular endothelial growth factor (VEGF) [2, 83, 84]. Furthermore, research evidence has revealed that AT1R activation in DU145, LNCaP, and PrSC cells can lead to elevated Janus kinase signal transducers and activators of transcription (JAK-STAT) signaling, mitogen-activated protein kinase (MAPK) activation, and cell proliferation [85, 86]. In fact, AT1R-activated cells recruit nonreceptor tyrosine kinases, for instant Src, that recruit a molecular complex containing JAKs [81]. ATR1 activation can exert antiapoptotic effects in vascular endothelial cells by promoting cell survival and also by inhibiting caspase-3 activity via activating the PI3K/ Akt signaling pathway [87, 88]. AT1R upregulation has been found in many different types of cancer, for example, breast, pancreatic, and lung cancers [83].

The involvement of AT1R in tumorgenesis may be explained as part of a complicated process involving a number of cell signaling pathways within endothelial cells, including EGFR and MAPK/ERK1/2 [89, 90, 86, 91]. Ang II can induce VEGF-A expression in endothelial cells through AT1R and ERK1/2 signaling, which are necessary for neovascularization and tumor growth. This promotes neo-angiogenesis in different tumors such as hepatocellular, ovarian, pancreatic, and bladder cancers [92, 93, 94, 95]. Aside from promoting angiogenesis, Ang II/AT1R also has the potential to relate to malignancy via EGFR transactivation. This can hijack downstream signaling pathways associated with malignant transformation and activate molecular cascades that promote cancer cell proliferation [96]. In summary, the PI3K/ AKT/mTOR, Jun activating kinase II (JAK)/signal transducer and activator of transcription 3 (STAT3), and RAS/ RAF/ERK1/2 pathways are three molecular cascades that can be involved in downstream of the AT1R [96, 97, 98]. The PI3K/AKT/capsase3 and p38MAPK pathways are all activated via VEGF receptors in endothelial cells, which further prevents apoptosis and promotes proliferation [96]. AT1R activation can increase cell proliferation in prostate cancer cell lines and cells generated from prostate stromal cell lines by activating the MAPK and STAT3 signaling pathways [85, 99, 86]. In patients with high blood pressure, overstimulation of AT1R is observed, which can affect vascular remodeling and facilitate cell migration during metastasis [100].

It has been discovered that Ang II can work synergistically with NF-κB in cancer development [101]. The transcription factor NF-KB is expressed in all cell types and is important for metastasis and cancer progression and is considered a survival transcription factor that inhibits apoptosis and increases cancer progression. NF-KB is also capable of controlling metastasis by inducing adhesion molecules which can help the spread of cancerous cells by adhering to vascular endothelial cells and entering the target site [102]. NF-κB can also promote angiogenesis by upregulating VEGF, which promotes neo-angiogenesis throughout migration and invasion [102, 103]. A study demonstrated that Ang II can promote cellular migration as well as induce phosphorylation of PI3K/Akt. Therefore, in general, Ang II has the ability to activate the AT1R/PI3K/Akt pathway, which can ultimately lead to elevated NF-κB and IKK α /β activity. This can increase matrix metalloproteinase, MMP2 and MMP9 expression and cell migration [97]. In breast and gastric cancers, Ang II/AT1R activation can promote survival and invasion through the NF-κB pathway, which is suppressed by the Ang 1–7/MAS receptor [97, 87]. In endothelial cells, NADPH oxidase and NF-κB induce angiogenesis [96]. Therefore, NF-κB activation through Ang II/AT1R signaling enhances tumor development and inhibits apoptosis [104]. It is noteworthy to mention that ACEIs and ARBs might be able to decrease NF-κB activity [105].

Ang II enhances cell growth and proliferation in various ways, such as through affecting tyrosine kinase [106], transforming growth factor-beta (TGF- β) [107], and activating mTOR pathways [106]. Ang II stimulation can increase the proliferation of breast cancer cells, which is regulated through MEK and PI3K signaling [108, 90]. Some studies have suggested that EGFR inhibition is required to fully prevent Ang II-mediated proliferation. In other words, Ang II-mediated proliferation in cancer involves complex mechanisms other than direct signaling from AT1R, such as EGFR transactivation [109, 90, 110]. Ang II can increase ROS production. A direct RAAS activation by Ang II in the tumor microenvironment might result in the production of ROS generation and proangiogenic and pro-inflammatory signaling [111]. ROS can function as signaling molecules that help regulate the pathways critical for cytoskeletal contraction control and cell proliferation stimulation. Additionally, the Ang II/ AT1R complex activates signaling pathways that regulate the production of extracellular matrix and enhances cell adhesion [112]. It is noteworthy that Ang II proliferation is usually related to certain types of cancer, but there is significant variation in responses even within a cancer type, suggesting that the genetic background of the cancerous cells can influence the cell proliferation potential [111].

Angiotensin-converting enzyme inhibitors (ACEIs) such as Captopril, Enalapril, Lisinopril, etc. can prevent the conversion of Ang I to Ang II by inhibiting the ACE enzyme. Therefore, less Ang II is produced and as a consequence the function of Ang II/AT1R pathway is reduced. Therefore, the use of ACEIs may have a preventive effect in cancer [100]. It should be noted that due to the existing conflicting results, more studies are required to draw definitive conclusions. Angiotensin receptor blockers (ARBs) such as Losartan, Valsartan, Telmisartan, Candesartan, etc. can block AT1 receptors. Therefore, it is possible that ARBs lead to the inhibition of the downstream pathway of this receptor by blocking AT1R and ultimately lead to a decrease in proliferation and cancer prevention. [113, 8]. However it is noteworthy due to conflicting results more studies are needed.

PRR pathway

It is well recognized that PRR is an important receptor involved in the regulation of RAAS. PRR can be involved in PI3K/AKT/mTOR, MAPK/ERK, and Wnt/β-catenin pathways, thus it can play a role in different pathological and physiological conditions, such as cancer [49]. Upregulation of PRR has been found in some types of cancers for instance pancreas cancer [56], prostate cancer [114], and leukemia [115]. It has been found that PRR can activate MAPK/ERK as well as PI3K/AKT/mTOR signaling pathways [116]. In pancreatic cancer cells, PRR pathway silencing reduces the expression of AKT, ERK1/2, and NF-KB [56]. In different cell types such as mesangial cells, monocytes, neurons, and collecting duct cells, PRR can activate MAPK/ERK signaling and up-regulate ERK1/2 [117]. ERK1/2 activation thus promoting cellular proliferation and inducing TGF- β generation, which plays a role in the pathogenesis and metastasis [118, 119]. PRR can also induce ROS production in an Ang II-independent manner and thus can contribute to PI3K/AKT/ mTOR and MAPK/ERK signal transduction [120]. It is generally recognized that the Wnt/ β -catenin pathway is involved in the progress of a variety of cancers, including breast [121], gastric [122], prostate [123], colorectal [124], and adrenocortical cancers [125]. PRR can be considered a crucial part of the Wnt receptor complex that can act as an adapter protein and therefore facilitate the attachment of Wnt ligands to the Wnt receptor complex [52]. Further research demonstrated that PRR probably can promote colorectal [58], pancreatic [55], and brain [54] cancers through the Wnt/-catenin signaling pathway. These data collectively suggest that PRR probably exerts its cancer-promoting effects through the PI3K/AKT/ mTOR, the MAPK/ERK, and the Wnt/β-catenin pathways [116]. Renin-inhibitors can exert their anticancer effects through decreasing the plasma renin activity by fulfilling its active sites and preventing the conversion of Angiotensinogen to Ang I [126].

Mas receptor and the AT2R pathways

Components of RAAS can have different roles in carcinogenesis. The signaling pathways triggered by the Ang II/AT2R signaling are poorly understood and recognized, compared with those.

triggered by Ang II/AT1R signaling, however generally, it has been demonstrated that activation of AT2R has protective effects, in contrast to AT1R [127, 81]. Therefore, while AT1R has proliferative properties, AT2R and Mas receptor have shown anti-proliferative effects [128, 129]. Ang 1–7 suppresses angiogenesis along with cell proliferation contrary to Ang II [130, 131]. The Ang 1–7/ Mas receptor and the Ang II/AT2R pathways are believed to be antagonistic to several cellular functions of the Ang II/AT1R pathway [79].

AT2R can inhibit cell proliferation while it can stimulate apoptosis in different types of cells, such as lung cells, vascular smooth muscle cells, endothelial cells, cardiomyocytes, and prostate cancer [132, 133, 134, 135]. Additionally, AT2R overexpression can cause apoptosis without the involvement of Ang II via the activation of caspase 3, caspase 8, and p38 MAPK, which is regulated by an extrinsic cell death-signaling pathway that is partly dependent on p53 [136, 137]. The major signaling pathways that AT2R activates include the PLA2/arachidonic acid signaling, the NO-cGMP system, and the activation of protein phosphatases. There are three different classes of protein phosphatases that are significantly activated by the Ang II/AT2R complex which consist of SH2 domaincontaining phosphatase (SHP-1), protein phosphatase 2 A (PP2A), and MAP kinase phosphatase 1 (MKP-1). Therefore, these proteins can regulate the inhibition of the PI3K/AKT, ERK/MAPK, and JAK phosphorylation cascades [81]. It has been discovered that AT2R signaling can inhibit cell survival, cell proliferation, invasion, and migration by inhibiting the MEK1/2 and Ras proteins and MAPK pathways, which is facilitated by phosphatasemediated dephosphorylation events [138, 139].

Mas receptor interference by Ang 1–7 can exert antiproliferative, anti-inflammatory, anti-migratory, and antifibrotic effects. It can affect vascular and cellular growth mechanisms and exert inhibitory effects [140] as well as preventing the growth of tumor cells in various cancers like esophageal squamous cell carcinoma [141], nasopharyngeal carcinoma [142], lung cancer [143], and prostate cancer [144]. Ang1-7/Mas axis can exert its beneficial effects on cancer through various mechanisms, such as exerting inhibitory effects on cellular proliferation and preventing metastasis and invasion [145]. In cultured tumor cells, the anti-proliferative Mas receptor/Ang 1-7 axis is probably mediated by the inhibition of ERK signaling [146]. In human nasopharyngeal carcinoma xenografts, it was reported that Ang 1-7 can down-regulate PI3K/Akt/mTOR pathway and inhibit tumor growth via autophagy [147]. A similar effect was reported in breast cancer as well, where Ang 1-7 reduced activation of the PI3K/AKT pathway, and VEGF expression [148].

AT2R receptor-interacting proteins (ATIP) are proteins that can interfere with the balance of AT1R and AT2R. The first ATIP member to be identified as constitutively interacting with AT2R at the cellular membrane is ATIP-1. The inhibitory effect of AT2R on ERK phosphorylation, receptor tyrosine kinase activation, and cell proliferation is predominantly mediated by ATIP-1 [149]. Additionally, ATIP-1 is involved in the transport and signaling of AT2R receptors. Another protein with tumor-suppressing properties that interacts with AT2R is ATIP-3. ATIP-3 levels is found to be lower in invasive breast cancer, and also the proliferation of tumor cells is decreased when ATIP3 expression is restored in breast cancer cells [150].

The endogenous substance alamandine exists in the blood circulation and cardiovascular system. The biological activities of alamandine are mediated via MrgD and are similar to those of Ang 1-7, including antihypertensive, antifibrotic, vasodilation, and central cardiovascular effects [151]. Alamandine can also reduce pulmonary fibrosis through the MrgD receptor. Studies have shown that alamandine/MrgD receptor axis appears to be critical for defending the body from SARS-CoV-2 disease [152]. Studies have revealed that alamandine/MrgD signaling, which is currently poorly understood, may be involved in the activation of different signaling cascades, such as the c-Src/p38 MAPK pathway [153]. Alamandine/ MrgD signaling can also activate protein kinase A and protein kinase C signaling pathways [154, 155]. Additionally, it has been reported that alamandine can activate the AMPK pathway [156], inhibit the MAPK pathway [157], and activate Jun-N-terminal kinase (JNK) phosphorylation to inhibit the NF- κ B pathway [158]. Signaling cascades triggered by alamandine/MrgD activation may mediate almandine protective functions and counteract Ang II's detrimental effects [81]. Further studies on the physiological effects of the alamandine/MrgD seem necessary and can be helpful in better understanding of the downstream of this pathway and the effects it probably has beyond the cardiovascular system effects, such as possible anti-proliferative effect, anti-cancer effect, etc. It has been reported that alamandine can induce a shift in anaerobic metabolism to aerobic metabolism in tumor cells, negatively regulate PI3K/AKT/mTOR signaling pathway, and activate FoxO1 transcriptional factor. These events may account for the observed anti-proliferative effects of alamandine, at least partially [159].

Drugs inhibiting RAAS elements

ACEIs and ARBs are the two primary RAAS inhibitors. These two are usually used in renal and cardiovascular disease management [160, 161, 162]. While ARBs specifically prevent Ang II from binding to downstream angiotensin receptors, ACEIs are able to inhibit the synthesis of Ang II [163]. Studies have indicated that ACEI/ARBs can alleviate cancer [83]. Since down-regulating RAAS has inhibitory effects on metastasis, cancer growth, and angiogenesis, ACEI and ARBs have been proposed as possible treatment options for cancer [164]. Following the effect of ARBs, the ability of Ang II for binding to AT1R is diminished, limiting proliferation capacity [78, 128]. The favorable effects of ARBs on the development of cancer may be described by the dominance of Ang 1-7 and AT2R over Ang II and AT1R. A study conducted on breast cancer showed that Ang 1–7 level has decreased, and the ACE2/Ang 1-7/Mas pathway has been

down-regulated [165, 166]. Researchers have discovered that the common ARB medicine, losartan, plays a role in lowering tumor growth rates in human pancreatic, breast, and prostate cancer cell xenografts [77, 167, 168]. Similarly to this, captopril, which belongs to the ACEI drugs group decreased the extent of liver metastases in addition to inhibiting tumor angiogenesis [169]. According to a study conducted on prostate cancer, ARBs such as telmisartan and candesartan have been able to suppress the expression of AT1R, induce apoptosis, and suppress cell proliferation [170, 171, 172]. A study showed that individuals with rectal cancer who consumed ARBs or ACEIs responded better to neoadjuvant therapy [173]. The use of RAAS inhibitors greatly enhanced overall survival rates and patient outcomes in different types of cancers, according to several other trials [4, 174]. (Some of the studies conducted by ACEI drugs and ARBs are listed in Table 1.)

Sex Hormone-dependent Cancers

RAAS components in ovarian cancer

The high mortality rate of ovarian cancer is due to its nonspecific symptoms. [175, 175] Despite greater knowledge of ovarian cancer, trends in treatment and survival have not changed considerably because it is still difficult to make an early diagnosis [175].

Angiogenesis is related to tumor invasion. AT1R has recently been determined to be highly and effectively expressed in ovarian cancer cells [176] and has been demonstrated that Ang II increases tumor cell invasion and VEGF expression through AT1R. In a mouse model, an AT1R blocker also inhibited cancer angiogenesis and prevented peritoneal dissemination of ovarian cancer [95]. Patients with positive AT1R staining exhibited considerably overall lower survival and reduced progression-free survival when compared to patients with negative AT1R staining. These findings demonstrate that in ovarian cancer, AT1R expression correlates with tumor angiogenesis and a poor prognosis, supporting the idea that AT1R could be considered a crucial molecule in tumor angiogenesis. This demonstrates that AT1R blockade therapy can probably be potentially a new molecular target for ovarian cancer therapy [176, 95]. According to a study that aims to evaluate the probable relationship between circulating ACE enzyme levels and ovarian cancer, it has been determined that ovarian cancer patients have increased serum ACE levels, and circulating ACE may be related to a pathobiological process in the carcinogenesis of ovarian. Therefore, ovarian cancer treatment in the future may involve targeting the RAAS pathway [177]. Dual targeting of AT1R and AT2R has also been investigated in epithelial ovarian carcinoma. It has been demonstrated that a novel therapeutic approach for epithelial ovarian carcinoma may involve dual regulation of

Table 1	Effect of RAAS inhibitors and their associated targets in various cancers	

Type of cancer	Drug	Target receptors	Clinical study	Outcome	Reference
Ovarian cancer		Ang II/AT1R	Human epithelial ovar- ian cancer tissues	In ovarian cancer patients, AT1R could be associated with tumor angiogenesis and a poor prognosis.	[339]
	Candesartan and PD123319	ATIR	- Human epithelial ovar- ian tumor tissues - SKOV-3 and HRA cell line - BALB/c mouse model of peritoneal carcinomatosis	 Ovarian cancer expresses AT1R, which is involved in angiogenesis and tumor growth. A promising approach to treating ovarian cancer may be AT1R blocking therapy. 	[95]
	losartan and captopril	PAX8	- OVCAR-4, OVCAR-8 and HEK293T cell lines	- Losartan and captopril can inhibit PAX8 expression and function. Therefore, they may be a suitable phar- macological target for various forms of HGSC.	[179]
	losartan	Ang II and AT1R and MCS formation	- A2780 and Ovca429 cell lines - The isogenic non- metastatic (NM) and highly metastatic (HM) cells - NOD-SCID mouse	 Ang II increases MCS formation and rise peritoneal metastasis of epithelial ovarian cancer cells. By overexpression of SCD1, AT1R activation enhances lipid desaturation, which finally can lessen endoplas- mic reticulum stress in MCS. 	[183]
		sPRR	– 197 primary epithelial ovarian cancer patients	- sPRR probably doesn't have prognostic, predictive, or diagnostic value in epithelial ovarian cancer	[176]
		Ang 1–7, Ang 1–9 and Ang 3–7	- OVPA8 cell line	 Peptides from RAS have the ability to regulate how nutrient-deprived tumors adapt to starvation. The availability of growth factors and nutrition has a significant impact on how angiotensin peptides affect cancer cells. 	[340]
		ACE	41epithelial ovarian cancer patients and 19 healthy controls	 serum ACE levels has been increased circulating ACE is probably related to pathobiologic process 	[177]
	losartan and CGP42112A	AT1R and AT2R	 – 58 human epithelial ovarian cancer tissues – 2774, A2780, PA-1, SKOV3ip1, HeyA8 cell lines - Human umbilical vein endothelial cells (HUVEC) 	Dual regulation of AT1R and AT2R, which inhibits angiogenesis and cancer cell survival, might be a potential for treating epithelial ovarian carcinoma.	[178]
	losartan	AT1R	- SKOV3ip1 and Hey-A8 ovarian cancer cells - orthotopic human ovarian carcinoma xenograft models - mouse fibroblast (10T1/2) - mice (<i>Mus muscu-</i> <i>lus</i>), and rats (<i>Rattus</i> <i>norvegcus</i>)	losartan therapy can improve paclitaxel's effectiveness losartan can increase medication delivery and im- prove vessel perfusion. losartan it can lower both the incidence and the volume of ascites produced.	[184]

Table 1 (continued)

Ang 1–9 and Ang 3–7

Ang 1–9 and Ang 3–7

AT1R

AT2R

AT4R

AT1R

AT2R

AT4R

Mas receptor

Mas receptor

PNT1A cell line

LNCaP cell line

PC3 cell line

Type of cancer	Drug	Target receptors	Clinical study	Outcome	Reference
Prostate cancer		Ang II and relaxin 2	- PNT1A cells	The development of some malignancies, particularly prostate cancer, may be influenced by the dysregula- tion of locally released peptide hormones including Ang II and relaxin 2.	[189]
	recombinant adenovi- ruses encoding AT2R (Ad5-CMV-AT2R-EGFP)	AT1R and AT2R	- DU145 cell lines - BALB/c nude mice	The findings demonstrated that AT2R overexpression can prevent prostate cancer tumor growth in vivo by preventing tumor cell proliferation and triggering apoptosis. The gene AT2R has the potential to be help- ful in prostate gene therapy.	[185]
	an AT2R agonist (C21)	AT2R	- LNCaP cells - transgenic rat for adenocarcinoma of prostate (TRAP) model	A potential new chemo preventive drug against human prostate cancer is AT2R agonist.	[186]
	fimasartan, losartan, eprosartan and valsartan	AT1R	PC-3, DU-145 and LNCa- pLN3 cell lines	Prostate cancer cells may exhibit anti-metastatic action and autophagy-associated cell death when exposed to ARBs.	[189]
	candesartan	AT1R and Caveolin-1	- Metastatic prostate adenocarcinoma cells (PC-3)	 The mRNA and protein levels of caveolin expression may be regulated by candesartan. growth inhibition of metastatic PC3 cell line 	[341]
	olmesartan	AT1R, ACE and Ang I/II	- DU145 and LNCaP cells - human prostate cancer cell lines - prostate stromal cells (PrSCs)	Prostatic RAS is overexpressed in hormone-resistant prostate cancer tissue, and different types of hormonal stimulation affect the expression of its components.	[342]
	Ang 1–7	Ang 1–7	- Human PC3 prostate cancer cells - SCID mice	For advanced prostate cancer, Ang 1–7 may function as an anti-metastatic agent and anti-angiogenic.	[343]
	Ang 1–7	Ang 1–7	- LNCaP human prostate cancer cells - Male athymic mice	A potential anti-angiogenic therapy for prostate cancer may be Ang 1–7.	[344]

Ang1-9 and Ang3-7 have opposite effects

Ang1-9: cause cell divisions, increase cell motility, and

stimulate the expression of genes including VEGF, HIF-

Ang 1–9 and Ang 3–7 might alter the overall number

the switch from hormone-dependent prostate cancer

to hormone-refractory illness may be facilitated by

1, VIM, and REL Ang3-7: exhibit no mitogenic action

of steroid receptors in aggressive prostate cells.

Ang 1–9 and Ang 3–7

[193]

[194]

Table 1 (continued)

Type of cancer	Drug	Target receptors	Clinical study	Outcome	Reference
Breast	Candesartan	AT1R	Xenograft mice model	Inhibition of tumor growth and angiogenesis	[345]
cancer	Ibesartan	AT1R	BC cell line	Suppression of cell proliferation	[346]
	Olmesartan	AT1R	BC cell line	Apoptosis induction	[347]
	Losartan	AT1R	Mouse model	Suppression of tumor cell proliferation, Reduction of inflammatory cytokines.	[348]
			BC Cell line	Reverse the Ang II-induced cancer cell proliferation and VGEF-A upregulation, Reduction in cell adhesion and invasion	[198, 345]
			4T1 and EMT6 murine TNBC cell lines	Improves the Antitumor Efficacy of Dox-L Chemotherapy, Combination Therapy Involved α-PD1 Immunotherapy Alleviates Primary Tumor Burden and Lung Metastases	[240]
			mouse (E0771 and 4T1 models)	Reduces stromal fibrosis, signalling Reduced solid stress, increase blood perfusion in tumours, enhance drug and oxygen delivery.	[77]
	Captopril	ACE1	TAM resistant BC cell line	Prevention of TAM resistant cells	[349]
			BC cell line with adipo- cyte-CM exposure	Reduction of cancer growth and inflammation in TME	[350]
			BC cell line	Inhibition of TF and VEGF expression	[351]
	Perindopril	ACE1	Mouse tumor xenograft and corneal micropock- et model	Inhibition of tumor growth and angiogenesis	[352]
	ACEI and / or ARBs	ACE 1and / or AT1R	Human studies (Meta-Analysis)	Decreased breast cancer risk with long-term ACEI / ARB use	[353]
			Human studies: Retro- spective study	No association with ACEI /ARB use and with disease free or overall survival	[242]
				higher incidence of invasive lobular carcinoma subtype	[354]
CRC	Telmisartan	AT1	In vitro	Inhibit cell proliferation, G0/G1 cell cycle arrest,	[254]
	Captopril	ACE	Mouse Model	Reduce EMT and invasive phenotype, Reduced tumor growth, reduction in volume of colorectal cancer liver metastases	[169, 355]
	Ibesartan	AT1R	Mouse Model	Reduced tumor growth, reduction in volume of colorectal cancer liver metastases	[169]
	Candesartan	AT1R	In vitro / CRC mouse models	Inhibition of CRC cell growth and migration, enhanced necrosis in CRC cells and tumors	[356]
	Valsartan	AT1R	In vitro/CRC mouse models	Inhibition of tumor cell growth, Induction of apoptosis, Inhibition of cell migration, ameliorate inflammation	[357]
	Losartan	AT1R	Xenograft Model	Inhibition of tumor growth, migration and angiogen- esis, increase pro-inflammatory cytokines	[358]
	LP2	AT2R	Xenograft model	Apoptosis in patient-derived xenografts of colorectal carcinoma, synergism demonstrated in combination of LP2 with 5-FU and the EGFR inhibitor erlotinib	[251]
	ACEis/ARBs	ACE1/AT1R	Human studies: Retro- spective cohort study	association with a lower colorectal cancer risk in a duration-response manner	[359]
			Human studies: Cohort analysis of SEER-Medi- care database	ACE inhibitors associated with decreased mortality	[257]
			Human studies : Retro- spective study	ACEI/ARB + BB exposure was associated with de- creased mortality compared to unexposed individuals, decrease in cancer progression, decrease cancer- related hospitalizations	[259]
				ACEI/ARB treatment may reduce tumor recurrence in left-sided CRC and early-stage CRC.	[260]

Table 1 (continued)

Type of cancer	Drug	Target receptors	Clinical study	Outcome	Reference
Gastric cancer	Candesartan Telmisartan Candesartan	AT1R	Human GC cell line Xenograft Model	Reduce cellular proliferation Inhibition of Tumor growth, Suppression of EMT and fibrosis	[266]
	TCV-116	AT1R	Xenograft Model	Inhibition of tumor growth, reduce VEGF	[360]
	Olmesartan	AT1R	N87 and MKN45 cell lines (Human GC cell line)	significant decrease in invasive ability of cancer cells	[263]
	RAAS inhibitors	ACE1/AT1R	Human studies: Cohort studies	Lower incidence of GC rate	[361]
Pan- creatic cancer	Losartan	AT1R	Orthotopic PDAC in mice	Increase drug uptake and functional microvasculature, Inhibition of Cell proliferation and VEGF expression, improve survival rate	[362, 363]
			A Phase 2 Clinical Trial (Total Neoadjuvant Therapy With FOLFIRI- NOX in Combination With Losartan Followed by Chemoradiotherapy for Locally Advanced Pancreatic Cancer)	Prolonged progression-free and overall survival.	[243]
	Captopril	ACE1	In vitro and In vivo studies	Targeted delivery of captopril to the CAFs, significantly down-regulated the deposition of ECM by blocking the TGF- β 1-Smad2 and consequently enhance the liposome-encapsulated chemotherapeutic agent gemcitabine.	[364]
	Irbesartan	AT1R	In vitro and In vivo studies	effectively reduced chemoresistance in PDAC	[292]
	Telmisartan	AT1	In vitro	Inhibit the human PDAC cell pro;iferation by cell cycle arrest, The miRNA expresseion profile was markedly altered	[365]
			Orthotopic PDAC mice model	increased vascular perfusion and reduced hypoxia,	[366]
			Wistar Bonn/Kobori (WBN/Kob) rat	Prevented the development of chronic pancreatitis and fibrosis.	[276]
			Human studies: Retro- spective analysis	longer overall survival in PDAC patients with non- metastatic disease and resected PDAC patients, improve clinical outcomes in patients with advanced pancreatic cancer in combination with Gemcitabine	[367–369]
			A population study	Improve survival	[293, 370]
Hepatic cancer	Candesartan	AT1R	Hepatic carcinoma cell line	Inhibition of proliferation	[306]
	Telmisartan	AT1R	non-alcoholic steato- hepatitis (NASH) rats	Prevent hepatocarcinogenesis through inhibition of angiogenesis	[371]
	Irbesartan	AT1R	Xonegraft model	Inhibition of HCC growth and Metastasis, Weaken adhesion to endothelial cells	[311]
	Losartan	AT1R	diethylnitrosamine-	Decrease cell migration, Metastasis, Inhibition of	[372]
	Fosinopril	ACE	induced hepatocellular	Neovascularization	
	Perindopril	ACE	carcinoma in mice		
	Azilsartan	AT1R	hepatocellular adeno- carcinoma cell line	Induce cell apoptosis	[373]
	Losartan	AT1R	Mouse model of HCC	reduced liver and peritumoral fibrosis , lenhance anti-PD-1-triggered HCC regression in	[374]
Esopha- geal	Telmisartan	AT1R	EAC Cell lines Xenograft Model	Inhibition of proliferation and cell cycle arrest Inhibition of tumor growth	[332]
cancer	Benazepril	ACE	Xenograft Model	Inhibition of tumor growth and angiogenesis	[335]
	ACEi and/ or ARBs	ACE/AT1R	Human studies: a nested case control study		[375]

AT1R and AT2R, which prevents angiogenesis and cancer cell survival [178].

Paired box transcription factor 8 (PAX8) is one of the most commonly used biomarkers in High-Grade Serous Ovarian Cancer (HGSC). It is expressed in about 90% of patients with HGSC. Although PAX8 is not expressed by the ovarian surface epithelium it is expressed by animal models of HGSC that are derived from it. It is indicated that it could be an effective factor in the development of cancer in addition to being a hallmark of Müllerian origin. PAX8 deficiency in HGSC cells can cause cellular tumor death and decreases invasion and cell migration. PAX8 deficiency can also decrease the amount of TGFbeta secretion, which is a cytokine that is crucial for the modification of the tumor microenvironment. Therefore, PAX8 inhibitors may be a suitable pharmacological target for various forms of HGSC. Losartan and Captopril are both RAAS pathway inhibitors that can prevent PAX8 expression and activity. Therefore, Losartan and Captopril can have beneficial effects on the treatment of ovarian cancer [179]. Losartan and Captopril were identified as drugs that can constantly suppress the expression of the PAX8 protein and inhibit the function of the PAX8 promoter. PAX8 can promote invasion and tumor cell migration through modulating FOXM1 and PKCa expression [180]. In OVCAR8 cells, Captopril and Losartan both inhibited the expression of fibronectin, PAX8, and PAX8 downstream proteins such as PKCa and FOXM1 [179].

The prognostic and predictive efficacy of soluble PRR as a biomarker for clinicopathological outcomes in primary epithelial ovarian cancer patients has been investigated. However, no correlation between levels of sPRR with clinicopathological factors and prognostic data was observed. sPRR seems to have no diagnostic, prognostic, or predictive value in epithelial ovarian cancer [176].

In ovarian cancer, multicellular spheroids (MCS) can be formed, which can lead to disseminating cells to accumulate together and ultimately promote cancer metastasis and prevent cell death [181]. In contrast to the majority of solid tumors that spread through the blood, ovarian cancer typically disseminates through the peritoneal cavity [182]. It has been found that Ang II/AT1R signaling is probably effective in the progress and promotion of ovarian cancer, so it could be considered a critical therapeutic target [183]. Ovarian tissue expresses RAAS components locally, and it is likely that enhanced expression of its components, such as Ang II and AT1R, can be related to the progression of ovarian cancer [177]. Ovarian cancer cells can overexpress Ang II as well as stimulate AGT gene expression to generate more Ang II, which can ultimately result in cancer development and MCS formation [183]. Through lipid homeostasis and stimulation of the lipid desaturation pathway, cancer cells can decrease endoplasmic reticulum stress brought on by saturated fatty acid overload, which ultimately prevents cancer cells from going into apoptosis. Sterol Regulatory Element-Binding Protein (SREBP) is a signaling pathway that Ang II/AT1R signaling can activate to induce lipogenesis. Stearoyl-CoA Desaturase-1 (SCD1) is an endoplasmic reticulum enzyme that is in charge of lipid desaturation. It is one of the lipogenesis enzymes that are upregulated by the SREBP pathway. Therefore, SCD1 can help cancer cells to prevent cell death [183]. AT1R activation can increase lipid desaturation through up-regulation of SCD1, which finally can decrease the endoplasmic reticulum stress of MCS. The above mechanism can clarify the link between elevated levels of AT1R and worse clinical outcomes in ovarian cancer patients [183].

Tumor fibrosis and Ang-driven fibrogenic signaling have been demonstrated to negatively correlate with survival in ovarian cancer patients. A study aimed to improve drug delivery and treatment efficacy by remodeling the dense extracellular matrix in two orthotopic human ovarian cancer xenograft models. It has been discovered that by using losartan to target the Ang signaling axis, extracellular matrix content, and related "solid stress" can be decreased, improving the anticancer therapeutic impact. It was also found that losartan therapy can improve paclitaxel's effectiveness by improving the tumor microenvironment. Consequently, it may lead to increased medication delivery and improved vessel perfusion. Although losartan medication alone does not diminish tumor burden, it can lower both the incidence and the volume of ascites produced. These results offer the justification and supporting information for a clinical investigation on losartan and chemotherapy combination therapy for ovarian cancer patients [184].

RAAS components in prostate cancer

Metastasis occurs in a significant percentage of prostate cancer cases. Most patients eventually develop chemo resistance and lose their responsiveness to additional hormonal therapy. Therefore, the development of novel therapeutic strategies for the treatment of prostate cancer is necessary [185].

AT1R and AT2R receptors can be considered possible therapeutic targets for the treatment of prostate cancer. AT1R inhibition and AT2R activation can have beneficial effects on prostate cancer treatment. Investigation on the effects of AT2R stimulation revealed that in vivo and in vitro treatment of an AT2R agonist increases apoptosis and reduces androgen receptors expression, hence lowering cell proliferation. Thus, AT2R activators were recommended as potential chemo-preventive therapy candidates for human prostate cancer [186]. Early stages of prostate cancer were shown to have an overexpression of Ang II receptors [187]. Also, there is an association between AT1R expression and prostate cancer cells' potential for metastasis [188]. AT2R overexpression in prostate cancer can reduce tumor growth, decrease Ki-67 expression, and induce apoptosis, according to an in vivo investigation. Furthermore, there was an inverse relationship between AT2R expression and prostate cancer aggressiveness [185].

RAAS was previously found to express itself in both healthy prostate tissue and cancer cells, demonstrating its importance in both normal cell function and tumor cell transformation. In healthy prostate cells, RAAS can be involved in sperm motility, spermiogenesis, and sperm survival. Long-term exposure of healthy prostate cells to Ang II can alter cellular morphology, and increase cell survival and cell proliferation via upregulating BCL2/ BAX ratios. It can also increase the degradation of the extracellular matrix. In summary, Ang II expression dysregulation in the prostate can increase the risk of prostate cancer [189].

It is reported that prolonged administration of ARBs or ACEIs in hypertensive patients can reduce the incidence of prostate cancer [190]. Moreover, consumption of high concentrations (100–400 μ M) of ARBs has been reported to enhance the death of prostate cancer cells via increasing cancer cell autophagy and increasing the expression of autophagy-related genes. In addition to the advantages of ARBs, Fimasartan was capable of reducing prostate cancer migration; as a result, it might be a potential treatment agent for prostate cancer in hypertensive patients [191].

Protein Tyrosine Kinase (PTK) activity was reported to be decreased by Ang 1–7 and Ang II/AT2R in androgenindependent prostate cancer. PTKs were shown to be elevated in the advanced stages of prostate cancer. This suggests that Ang 1–7 and Ang II may be involved in providing protection against prostate cancer in advanced stages, or in androgen-independent prostate cancer [192].

In a study, the effects of Ang 1–9 and Ang 3–7 in prostate epithelial cells have been investigated. The findings of this study demonstrate that Ang 1-9 and Ang 3-7 have opposite impacts on the in vitro biological characteristics of prostate cells. By causing cell divisions, increasing cell motility, and stimulating the expression of genes including vascular endothelial growth factor (VEGF), hypoxia-inducible factors (HIF-1), vimentin (VIM), and REL proto-oncogene, NF-kB subunit (REL), Ang1-9 appears to have pro-cancer effects. Ang3-7, on the other hand, exhibited no mitogenic action [193]. Further investigation on the effects of Ang 1-9 and Ang 3-7 on prostate cancer was also conducted. The results imply that Ang 1–9 and Ang 3–7 can affect the biological characteristics of prostate cancer cells via modulating genes involved in the inflammatory and steroidogenesis pathways. Additionally, it was demonstrated that Ang 1–9 and Ang 3–7 might alter the overall number of steroid receptors in aggressive prostate cells. This indicates that the switch from hormone-dependent prostate cancer to hormone-refractory illness may be facilitated by these peptides [194, 186]

RAAS components in breast cancer

RAAS components are presented in both normal and cancerous breast tissues. The physiological activities of local RAAS, particularly in the epithelial tissue are tissue growth and modeling [195–197]. Much evidence suggests that dysregulation of this system influences cancer progression, metastasis, and resistance to cancer therapeutics [198, 199]. Various components of RAAS play different roles in the biology of breast cancer which are discussed in the following sections.

AT1R pathway

The Ang II/AT1 receptor axis as the major and most well-known RAAS pathway plays a pivotal role in many cellular processes including proliferation, migration, angiogenesis, and inflammation, and is strongly associated with tumorigenesis [200]. AT1R activation through binding of Ang II, induces cell proliferation and attenuates cell death via PI3K/AKT signaling and ERK/MAPK signaling pathways [201]. Furthermore, PI3K/AKT activation by AT1R influences NF-KB which is involved in cell migration [97]. It also causes angiogenesis by increasing the VEGF generation to maintain tumor growth [79, 202]. In the triple-negative breast cancer cell line, RAAS plays a role in angiogenesis by increasing HIF-2 α , and TIMP-1 [203]. RAAS activation through the AT1 receptor is involved in migration and metastasis through up-regulation of MMP-2 and MMP-9 expression and enzymatic activity in breast cancer cells [204]. It has also been proven that over-expression of Intracellular Adhesion Molecule-1(ICAM-1) which is associated with the aggressive phenotype of breast cancer cells results in response to Ang II treatment [205].

A study by Ma et al. proposed that the role of AT1R for lymph node metastasis is AT1R-mediated and is through CXCR4/SDF-1 α . The study has revealed that this signaling pathway would further activate the members of the FAK family and upregulate Rho A. The FAK/RhoA signaling stimulates the ROCK, which promotes cell contractility and migration to reach lymph nodes [206].

Inflammation, another mechanism of AT1R activation, is viewed as a hallmark-facilitating characteristic of cancer [207]. The tumor microenvironment simulates an inflammation site that possesses remarkable advantages for tumor progression, metastasis, and immunosuppression [208, 209]. There is strong evidence that could confirm that RAAS activation via AT1 receptor signaling may promote an inflammatory TME [210–212]. Downstream activation of the AT1 receptor in breast cancer cells, triggers CBM/NF-KB-mediated inflammatory response by releasing inflammatory cytokines and growth factors including IL-1β, IL-8, IL6, and VEGF to the microenvironment [213, 214]. Another possible relationship between Ang signaling via the AT1 and TME that can be addressed is Ang II-AT1R-mediated activation of NADPH oxidase (NOX) complexes leading to ROS generation and oxidative stress [112, 207, 215-217] which is involved in inflammation and angiogenesis in TME [111]. Immune and stromal cellsin TME also contain immune cells and stromal cells that are capable of expression of RAAS components [218-221]. Tumorassociated macrophages (TAMs) are the most abundant immune cells that reside in breast tumors [222]. These intratumoral immune cells contribute to the formation of immunosuppressive microenvironment [223, 224]. Ang II induces TAM infiltration into TME. In a study by Nakumara et al.the abrogation of Ang II effect via AT1R by receptor blockade or AGT (Angiotensinogen)silenced breast tumor cells decreased the infiltration of macrophage and fibroblasts into tumor regions and further improved tumor response to immune-checkpoint inhibitors such as programmed cell death protein 1 (anti-PD-1) [143, 199, 225].

AT2R pathway

There is not much known about the role of the AT2 receptor in breast cancer. AT2 and AT1 receptor coexpression in breast tumors on one hand and the fact that both of these receptors are the target of the local Ang II on the other hand, suggests the interplay roles of these receptors [226-228]. AT2 receptor is expressed at low levels in normal breast ducts, however; its expression significantly increases under pathological conditions such as breast hyperplasia, Ductal Carcinoma In Situ (DCIS), and aggressive breast cancer [229]. On the other hand, studies report that there is no correlation between the expression of AT2R and clinical and pathological characteristics in breast cancer including proliferation and angiogenesis [226]. Another study revealed the possible role of AT2 receptor activation in migration and metastasis inhibition in breast cancer cells by shutting down CAV1/Rab5/ Rac-1 signaling pathway [138].

ACE2/Ang 1–7/ mas receptor pathway

ACE2/Ang 1–7/ Mas Receptor is another arm in the RAAS that is recognized with the counter-regulatory role for Ang II/AT1R [230]. It is demonstrated that Ang 1–7, counteract Ang II-induced metastatic functions in breast cancer cells. Accordingly, VEGF and MMP-9 expression and activation are abolished by Ang 1–7 effect through Mas receptor signaling. It is further indicated that Ang

II-induced ERK1/2 activation which also promotes EMT in cancer is inhibited by Ang 1–7 [231]. Additionally, down-regulation of ACE2/Ang 1-7/MasR, induce metastasis by activating the SOCE (Store-operated calcium Entry) and PAK/NF-ĸB/Snail1 pathways [165]. The role of ACE2 as a potential antitumor component in the RAAS family has been reported in breast cancer. It is suggested that ACE2 inhibits angiogenesis by suppressing the VEGFa/VEGFR2/ERK pathway in breast cancer [232]. A pan-cancer analysis studied the correlation of immunological features of ACE2 in breast cancer. The study uncovered that ACE2 expression in high levels is associated with an inflamed TME or so-called immunehot tumors. ACE2 expression is positively related to the levels of immunomodulators and infiltrating levels of TIICs (Tumor-Infiltrating Immune Cells) in breast cancer TME. In fact, it is suggested that the product of ACE2, Ang 1-7 exhibits anti-tumor effects by remodeling TME and sensitizes the tumor to respond to chemotherapy and anti PD-1 immunotherapy [233]. Another study demonstrated that in breast cancer cells the production of Ang 1–7 is significantly lower in comparison to healthy breast tissue, whereas the expression of the Mas receptor was a few times higher than in normal cells [234]. Considering the multiple benefits of anti-cancer effects and the protective functions of Ang 1-7/MasR signaling, the suggestion of balancing Mas receptor activity with agonist substances may be a potential therapeutic option in breast cancer [235].

In clinical perspective, the question of inhibiting the RAAS by current ARBs and ACEIs as a therapeutic potential target in breast cancer still remains a topic of debate. There is a growing body of evidence in pre-clinical and clinical studies which shows the positive role of RAAS inhibitors in cancer treatment intervention [236] One reason that make RAAS an attractive candidate as a therapeutic strategy for treatment of breast cancer, is the desmoplastic reaction in breast tumors. Several studies demonstrated that RAAS inhibitors can normalize the fibrotic stroma in cancer [237]. In a study by Chaugan et al. data indicated that Losartan, is a dual inhibitor of collagen and hyaluronan synthesis. The data suggested that losartan decompress vessels in desmoplastic tumors by reducing solid stress and antimatrix effects. This effect further enhance the drug delivery and effectiveness of chemotherapy. [77, 238-240].

A recent meta-analysis by Xie et al., which emphasized on the association between RAAS inhibitors and breast cancer demonstrated that long-term use of RAS inhibitors, reduces the risk of breast cancer compared with non-users [241]. However the results of another meta-analysis did not show association with disease free and Overall survival [242]. The heterogeneity in terms of tumor stage, the hormone receptor status and other tumor characteristics may affect the data analysis to report contrary results [237]. Nonetheless, there are clinical trials on different types of cancer including breast cancer to explore the potential role of RAAS inhibitors as adjunctive treatment options in breast cancer [243].

RAAS Components in Colorectal Cancer (CRC)

RAAS receptors and mediators have been identified in normal colon tissue. Localization of AT1R and AT2R determined by immunohistochemistry has shown that AT1 receptors are found in surface epithelium, crypt bases, and lamina propria, whereas AT2R s are mainly in the mesenchymal cells of the lamina propria [244]. The physiologic functions of these receptors are different based on Ang II-mediated activation. AT2R stimulation plays a role in water and sodium absorption from the colon. AT1 receptor-mediated pathway leads to sodium reabsorption, sodium secretion, and colon motility [245, 246].

Components of RAAS have been demonstrated to have a close relationship with the progression and metastasis of CRC. CRC presentation is often associated with advanced tumor stage and liver metastasis [247]. AT1R expression is found to play a pro-tumoral role and has a correlation with liver metastasis and pathological tumor stage [248, 249]. In AT2R knockdown CMT93 cells followed by Ang II dose-dependent treatment, proliferation, invasion, and VEGF secretion were observed which contributed to AT1R stimulation. The same study elucidated that AT2R activation via Ang II treatment in low concentrations is associated with increased VEGF secretion, whereas the high concentration of Ang II resulted in antitumoral effects by inhibition of cell growth and metastasis [249]. Loss of expression of AT2R in advanced and aggressive CRC cells may also be contributed to CRC progression [249, 250]. A study by Namsolleck et al. demonstrated that LP2, the cyclic Ang (1-7) analog which is a specific agonist of AT2 receptor, inhibits the growth of patient-derived xenografts of CRC in mice via Pi3K/AKT/mTOR signaling which results in apoptosis via CDKs. This modified peptide of Ang (1-7) with the N-terminal D-lysin and high specificity, stability and excellent pharmacokinetics, showed antitumor activity against CRC in mice models which also acted synergistically with 5-FU and Erlotinib. [251].

A study on patients with colon adenocarcinoma has shown that Mas Receptor was markedly upregulated in comparison to the non-neoplastic region [250, 253]. However, treatment of human colon adenocarcinoma expressing MasR with Ang 1–7 exhibited no change in cell cycle events and proliferation [253]. The increase of MasR in cancer development in CRC in comparison to normal tissues suggests a potential role for this receptor that needs to be elucidated.

An in vitro study by Oura et al. indicated that Telmisatan can inhibit the proliferation of HCC and induce cell cycle arrest [254]. The use of an AT1R inhibitor (Irbesartan) and an ACE inhibitor (Captopril) in a mice model of CRC liver metastasis further elucidated the role of Ang II/AT1R signaling in the induction of metastasis [169]. In fact, the blockade of AT1R has shown to promote antitumor effects in KCs in the early stages of tumor growth [169, 255]. Furthermore, the study of RAAS inhibition by an Ang II receptor blocker, Valsaratan, in CRC mouse models demonstrated CRC cell growth suppression and inhibition of cell migration by perturbation of MMP-2 and MMP-9 [256]. Clinical data analysis also represents an association between the use of RAAS inhibitors and better outcomes in CRC [173, 257-259]. A retrospective analysis of clinicopathological data of CRC patients exhibited that ACEI/ARB treatment may reduce tumor recurrence in left-sided CRC and early-stage CRC [260]. However further investigation and prospective clinical trials is required to establish the clinical significance of ACEIs and ARBs in CRC treatment and recurrence.

RAAS components in gastric cancer (GC)

Localization of AT1R and AT2R in the epithelium and lamina propria is suggested to be contributed to the functional impact of these elements in physiological conditions. AGT and Renin are both expressed by resident mesenchymal cells in lamina propria which contribute to the formation of the principal mediator, Ang II, in the tissue [261] [262].

The association of RAAS components with gastric cancer has been reported previously [263]. The most studied RAAS axis in gastric cancer is the ACE/Ang II/ AT1R which is found to elaborate on many mechanisms involved in cancer progression. The expression of Ang II, AT1, and the ACE activity in gastric cancer patients is upregulated in comparison to healthy tissues [264]. Ang II is reported to have a role by its impact on the induction of MMP-2 and MMP-9 expression in human gastric cancer cells which contribute to tumor invasion and metastasis [204]. Ang II also has the potential to increase proliferation and impair apoptosis by ERK1/2 and NF-κB activation and overexpression. Furthermore, AT1 receptor blockade confirms the anti-proliferative and tumor growth inhibition role of Ang II/AT1R-induced signaling in gastric cancer [265]. In another study, Candesartan significantly suppresses tumor proliferation and fibrotic changes in mice subcutaneous xenograft model of gastric cell line with human peritoneal mesothelial cells (HPMC). This is suggested to be by TGF- β 1-induced EMT change and stromal fibrosis by Ang II/ATR signaling which is suppressed by blockade of AT1 receptor. [266].

In a study on GC patients, the local expression of AT2R is reported [267]. However, its role in GC has been controversial. A study on human GC cell lines demonstrated that in vitro blockade of the AT2 receptor reduced invasive abilities of the tumor cells with no effect on cell number [268]. In another study on gastric cancer patients, it is demonstrated that the AT2 expression along with AT1 and ACE was increased compared to healthy tissues [264]. In the contrary, in another study, AT2R expression has been demonstrated to be significantly downregulated in GC patients. Moreover, activation of AT2R decreases migration and invasion of cancerous cells in an intraperitoneal carcinomatosis xenograft mouse model. It also decreases tumor formation and increases mouse survival in an in vivo model [269]. Thus, further investigation on the role of AT2R in GC requires to be determined considering the counter-regulatory effect on the AT1 receptor which is implied in other cancer cells.

RAAS components in pancreatic cancer

[270]. The expression of RAAS components has been found in different cell types of the pancreas including pancreatic ducts, endothelial, acinar, and stellate cells [270–272]. It has been demonstrated that RAAS components have roles in the regulation of many physiological mechanisms and are responsive to pathophysiological conditions including pancreatic cancer [273–275].

It is suggested that RAAS components play an important role in pancreatic cancer growth and progression [276, 277]. Over-expression of AT1R mRNA and protein has been observed in pancreatic cancer tissue compared to normal tissues. This study also demonstrated that AT1R selective antagonist (L-158,809) suppresses the growth of human pancreatic cancer which suggests the role of Ang II and AT1R in pancreatic cancer [277]. In a study on miR-410 role in pancreatic cancer, it was demonstrated that AT1R mRNA is the direct target of this microRNA which also suppresses AT1R expression [278]. Furthermore, it was suggested that miR-410 suppresses angiogenesis partly through inhibiting the ERK1/2 pathway via AT1R suppression. It is also demonstrated that AT1R knockdown results in suppression of cell proliferation and invasion in pancreatic cancer cells [278].

Inflammation is one of the key steps in the development of pancreatic cancer. Studies have found that fibrosis is involved in the transformation from pancreatitis to pancreatic cancer and further tumor growth and metastasis [279, 280]. Pancreatic stellate cells are the key mediators of fibrosis [281]The role of Ang II in pancreatic inflammation and fibrosis has been investigated in WBN/Kob rats. It was demonstrated that Ang II stimulates up-regulation of α -SMA and TGF- β which are the known fibrosis markers that are expressed by activated pancreatic stellate cells [276, 282]. ROS accumulation is enhanced following chronic inflammation and pancreatic fibrosis compared to normal tissues. Given that Ang II mediates ROS generation [283], oxidative stress induced by Ang II signaling promotes pancreatic fibrosis [276].

Pancreatic ductal adenocarcinoma (PDAC) is the most common and also most lethal pancreatic cancer type [284] The study on human PDAC specimens indicated that AT1R expression is significantly higher than in normal pancreas, whereas AT2R expression level in neoplastic ductal epithelium was slightly lower than in normal pancreas [285]. This study also demonstrated that selective AT2R agonist treatment using murine PDAC grafts attenuates the growth of tumor in volume and weight by inducing apoptosis in PDAC cells [285]. This suggests the potential benefit of AT2R agonists in combination with AT1R antagonists in PDAC treatment.

The expression of the ACE2/Ang 1–7/MasR axis is confirmed in the pancreas of an animal model of Severe Acute Pancreatitis (SAP) [286]. The study on in vitro model of pancreatitis demonstrated that the ACE2/Ang 1–7/MasR axis significantly suppresses the p38 MAPK/ NF- κ B signaling pathway in mouse acinar cells which inhibits pancreatitis [287]. Localization and expression of ACE2 in human pancreatic cells particularly in beta cells, have also been reported [288]. Although the protective and anti-inflammatory effects of the ACE2/Ang 1–7/ MasR axis have been recognized in rodents and in vitro studies of the pancreas [287, 289–291], further investigation of this axis is needed to shed light on its exact role in human pancreatic cancer.

Common ACE inhibitors and ARBs have emerged as promising therapeutic strategies in PDAC. In a study by Zhou et al. using a high-throughput screening platform of the Gemcitabine-resistant PDOs (patient-derived organoids), reported that Ibesartan, reduces resistance to chemotherapy. In vitro and in vivo studies demonstrated that ibesartan sensitizes PDAC tumors to chemotherapy. The mechanism underlying gemcitabinesensitivity is found to be by suppressing c-Jun expression via repressing the activation of Hippo/YAP1/TAZ signaling pathway. It is revealed that tumoral c-Jun significantly enhances tumoral iron metabolism and promotes cancer cell stemness in PDAC which can be inhibited by ibesartan [292].

Several clinical trials have investigated the role of losartan in combination with chemotherapy for PDAC. A phase II trial found that adding losartan to gemcitabine and nab-paclitaxel improved progression-free survival and overall survival compared to chemotherapy alone in patients with locally advanced PDAC. Another retrospective study of PDAC patients undergoing resection indicated that RAAS inhibitor users compare to non-users have a prolonged overall survival [293]. Losartan was found to be an Extra cellular matrix (ECM) modulator by inhibition of TGF β 1 expression, collagen I and hylouronan synthesis in cancer associated fibroblasts [276], while losartan and other RAAS inhibitors have shown promises in preclinical and clinical studies, more research is needed to fully understand their potential as a treatment option [294].

RAAS components in Hepatocellular Carcinoma (HCC)

Liver cancer comprises different types of hepatic neoplasms with hepatocellular carcinoma (HCC) as the most common type of liver cancer, representing more than 80% of the cases [295]. The pivotal role of RAAS in HCC development and other liver diseases such as liver fibrosis has been documented [296, 297]. Based on findings, local RAAS components are all present in the liver tissues and in its different cell types including hepatocytes, cholangiocytes, and hepatic Kupffer cells [252, 298, 299]. Local RAAS function in the liver is crucial under physiological and pathophysiological conditions as it plays a role in cellular functions including metabolic processes, regulation of cell proliferation, angiogenesis, and apoptosis [300–302].

Many studies have shown that Ang II/AT1R activation promotes cell proliferation, angiogenesis, inflammation, fibrosis, and ECM remodeling in HCC which is associated with cancer progression [303-305]. A study on HepG2 cells demonstrated that Ang II induces cell proliferation by AT1R/Raf/ERK1/2 signaling pathway. In support, cell treatment with Candesartan (AT1R blocker), Sorafenib (Raf inhibitor) and ERK inhibitor PD98059 indicated significant inhibition of cell proliferation [306]. Another study on HCC cell lines evaluates the downstream effects of Ang II/AT1R signaling in HCC. Their data proposed that the AT1/PKC/NF-κB pathway enhances the proliferation and inflammation induced by Ang II in HCC cells. Based on the results, stimulation of AT1R activates Protein Kinase C (PKC) signaling which plays an important role in cell proliferation and migration of HCC cells [307]. It is further demonstrated that the PKC plays a key role in NF-κB signaling pathway [303]. NF-KB signaling maintains the inflammatory response of HCC cells and also contributes to cancer progression and survival as well as enhancing cell proliferation [303, 308]. The role of RAAS in HCC was highlighted by targeting the AT1R and by mean of microRNA-152 (miR-152). The results suggested that miRNA-152 expression resulted in down-regulation of AT1R, expression of vimentin, N-cadherin and the number of invaded and migrating cells [309].

Ang II/AT1R activation also induces angiogenesis by activating signaling pathways including AT1/JAK2/ STAT3/SOCS3 [310] which confirms the results from a study on subcutaneous xenograft tumors derived from HCC cells in nude mice treated with Candesartan. In this experiment, Ang II promoted VEGF through AT1R activation. Moreover, microvessel density (MVD) as an angiogenic parameter of tumors is positively correlated with AT1R and VEGF expression. Candesartan is able to significantly reverse these effects and exhibit anti-angiogenic effects [94]. An In vivo study experimenting the role of Ang-II/AT1R signaling in metastasis revealed that Ang II upregulates VCAM-1 via the p38/MAPK pathway in HCC cells which is related to adhesion to endothelial cells and promotes metastasis [311].

Most HCCs are developed in the context of liver fibrosis, with 90% of the cases developing on the background of liver cirrhosis [312, 313]. In this regard, Hepatic Stellate Cells (HSCs) play a key role [314]. The activation of HSCs along with extracellular deposition are the key steps in liver fibrosis [315]. RAAS is one of the main mediators of HSCs activation [316, 317]. Ang II is known to be the central effector in activating the HSCs to produce pro-fibrotic markers such as TGF-B1 and promote inflammation and oxidative stress [316, 318]. In vivo investigation of RAAS inhibition by an ARB (Candesartan) and an ACEI (Perindopril) on pig serum-induced liver fibrosis model significantly attenuated fibrosis development [319]. In many studies, RAAS blockade by AT1R inhibitors or ACEIs led to reduced liver fibrosis by inhibition of HCSs through suppression of AT1R/TGF-β/Smad and AT1R/TGF-β/MAPK signaling pathways which further decrease the generation of ECM [320-322]. Supporting data suggested that using losartan, enhances anti-PD-1-triggered HCC regression in a mouse model of HCC [323].

The role of AT2R in various types of cancer has been controversial. AT2R expression in HCC cell lines is found to inhibit proliferation via down-regulation of CDK4 and cyclinD1. It is also demonstrated that AT2R induces apoptosis via MAPK pathway, Caspase-3 and Caspase-8 signaling. On the contrary, an in vivo study indicated that while high levels of AT2 receptor induce apoptosis, moderate expression of this receptor promotes cell proliferation and increases tumor growth. This implicates that the level of AT2R expression mediates different effects on HCC cells [324].

ACE2, in another arm of the RAAS was found to have prognostic value in HCC. The immunohistochemical analysis of ACE2 expression in an HCC microarray revealed that ACE2 expression was down-regulated in HCC tissues compared to normal liver. Secondly, in vitro and in vivo experiments showed that ACE2 overexpression inhibits HCC tumor growth through blocking the aerobic glycosis in HCC via Ang (1–7)/Mas Receptor axis with the downstream signaling cascade of p-SHP2/ ROS/HIF1 α axis [325]. The beneficial effects of the Ang 1–7/MasR axis have been previously reported in different types of cancer cells [326, 327]. Ang 1–7 inhibits proliferation and induces apoptosis by caspase-3 activity in vitro and in vivo. Another antitumor impact of Ang 1–7/MasR in HCC is the antiangiogenic effect which is mediated by down-regulation of VEGF-A secretion. Ang 1–7 is also proposed to play an important role in the expression of RAAS receptors. Ang 1–7 has been shown to down-regulate AT1R and upregulate AT2R along with MasR expression in vivo and in cultured H22 cells. It is suggested that Ang 1–7 may exert its antitumor effects by both MasR and AT2R activation and antagonizing AT1R effects [328].

RAAS components in esophagus cancer

Previous studies confirmed the presence of local RAAS in human esophageal mucosa. Ang II receptors and ACE are expressed in the human esophageal epithelium. The physiological actions of RAAS in esophageal mucosa are suggested to be by regulation of the ion transport [329] and also the esophagus contraction [330]. The role of RAAS the in pathophysiology of many esophagus malignancies such as esophageal adenocarcinoma and Esophageal Squamous Cell Carcinoma (ESCC) has been demonstrated [331, 332].

It is demonstrated that AT1R overexpression is significantly associated with higher T classification and overall worse prognosis. In vitro and In vivo studies indicated that ESCC cell proliferation is stimulated by Ang II in a dose-dependent manner. The mechanism underlying this effect is suggested to be via Ang II/AT1R signaling and further activates the mTOR pathway [332, 333]. In support, findings suggested that Telmisartan inhibited human esophageal adenocarcinoma (EAC) cell proliferation and tumor growth by inducing cell-cycle arrest via the AMPK α /mTOR pathway in EAC cells [332]. In vivo and In vitro studies also demonstrated the role of ACE inhibitors and ARBs in the suppression of angiogenesis by inhibition of VEGF expression [334, 335]. However, RAAS inhibition by ARBs and ACEIs as a therapeutic strategy in oesophgeal cancer has not shown prognostic effects. The retrospective study of patients with EAC and ESCC found no difference in overall survival and diseasefree survival between users of RAAS inhibitor and nonusers [336, 337].

The study of AT2R expression in ESCC cells indicated that overexpression of AT2R is associated with high T classification and AT1R expression. Moreover, the use of an AT2R antagonist, PD123319, exhibited no effect on ESCC cell proliferation [333]. Hence, the role of AT2R in esophageal cancer is still not well recognized.

The Ang 1–7/MasR axis is also investigated in Esophageal cancer. Findings suggested the antiproliferative effects of Ang 1–7/MasR signaling in vitro [338]. The study of the clinical impact of MasR on ESCC patients receiving curative esophagectomy demonstrated low MasR expression being correlated with a higher recurrence rate, poor disease-free, and overall survival. [338]. It is suggested that Ang 1–7/MasR axis could be both recognized as a diagnostic tool and an adjuvant therapy.

Conclusion

RAAS as a fundamental system is proven to have a potential role in proliferation signaling, cell migration, angiogenesis, invasion, and metastasis. Expression of RAAS receptors has been identified in various tumors, proposing these new-in receptors in cancer as novel markers for the early detection of malignancies. Additionally, Ang II, as one of the major peptides of RAAS, is demonstrated to prompt cancer metastasis by affecting and upregulating metastatic hallmarks. Thus, the use of drugs that affect RAAS may be proposed as an adjuvant treatment option in cancer. Prophylactic use of these agents may also aid in the prevention of tumor formation.

Considering the somewhat safe profile of RAAS inhibitors propose the use of RAAS modulators as a promising strategy in preventing and curing cancer as well as metastasis.

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References

- Kilmister EJ, Tan ST. The role of the renin–angiotensin system in the cancer stem cell niche. J Histochem Cytochemistry. 2021;69(12):835–47.
- Luan Z, Liu B, Shi L. Angiotensin II-induced micro RNA-21 culprit for nonsmall-cell lung adenocarcinoma. Drug Dev Res. 2019;80(8):1031–9.
- Nehme A, et al. An update on the tissue renin angiotensin system and its role in physiology and pathology. J Cardiovasc Dev Disease. 2019;6(2):14.
- Sun H et al. Do renin–angiotensin system inhibitors influence the recurrence, metastasis, and survival in cancer patients? Evidence from a meta-analysis including 55 studies. Medicine, 2017. 96(13).
- Haznedaroglu I, Malkan U. Local bone marrow renin-angiotensin system in the genesis of leukemia and other malignancies. Eur Rev Med Pharmacol Sci. 2016;20(19):4089–111.
- Hanif K, Bid HK, Konwar R. Reinventing the ACE inhibitors: some old and new implications of ACE inhibition. Hypertens Res. 2010;33(1):11–21.
- Dolley-Hitze T, et al. Angiotensin-2 receptors (AT1-R and AT2-R), new prognostic factors for renal clear-cell carcinoma? Br J Cancer. 2010;103(11):1698–705.
- 8. Afsar B, et al. Renin-angiotensin system and cancer: epidemiology, cell signaling, genetics and epigenetics. Clin Transl Oncol. 2021;23:682–96.
- Ager El, Neo J, Christophi C. The renin–angiotensin system and malignancy. Carcinogenesis. 2008;29(9):1675–84.
- Santos RA, et al. Angiotensin-converting enzyme 2, angiotensin-(1–7) and mas: new players of the renin–angiotensin system. J Endocrinol. 2013;216(2):R1–R17.
- de Miranda FS, et al. Breast cancer and the renin-angiotensin system (RAS): therapeutic approaches and related metabolic diseases. Mol Cell Endocrinol. 2021;528:111245.
- Kobori H, et al. The intrarenal renin-angiotensin system: from physiology to the pathobiology of hypertension and kidney disease. Pharmacol Rev. 2007;59(3):251–87.
- Lavoie JL, Sigmund CD. Minireview: overview of the renin-angiotensin system—an endocrine and paracrine system. Endocrinology. 2003;144(6):2179–83.
- 14. Passos-Silva DG, Verano-Braga T, Santos RA. Angiotensin-(1–7): beyond the cardio-renal actions. Clin Sci. 2013;124(7):443–56.
- 15. Ferreira A, Santos R. Cardiovascular actions of angiotensin-(1–7). Braz J Med Biol Res. 2005;38:499–507.
- Ferrario CM, Varagic J. The ANG-(1–7)/ACE2/mas axis in the regulation of nephron function. Am J Physiology-Renal Physiol. 2010;298(6):F1297–305.
- Nguyen G, et al. Specific receptor binding of renin on human mesangial cells in culture increases plasminogen activator inhibitor-1 antigen. Kidney Int. 1996;50(6):1897–903.
- Bader M. Spotlight on renin: the second life of the (pro) renin receptor. J renin-angiotensin-aldosterone Syst. 2007;8(4):205–8.
- 19. Ocaranza MP, Jalil JE. Protective role of the ACE2/Ang-(1–9) axis in cardiovascular remodeling. International journal of hypertension, 2012. 2012.
- 20. Flores-Munoz M, et al. Angiotensin-(1–9) attenuates cardiac fibrosis in the stroke-prone spontaneously hypertensive rat via the angiotensin type 2 receptor. Hypertension. 2012;59(2):300–7.
- 21. Lu H, et al. Structure and functions of angiotensinogen. Hypertens Res. 2016;39(7):492–500.
- 22. Fukamizu A, et al. Chimeric renin-angiotensin system demonstrates sustained increase in blood pressure of transgenic mice carrying both human renin and human angiotensinogen genes. J Biol Chem. 1993;268(16):11617–21.
- Yang G, et al. Functional expression of the human angiotensinogen gene in transgenic mice. J Biol Chem. 1994;269(51):32497–502.
- 24. Ferrario CM, Iyer SN. Angiotensin-(1–7): a bioactive fragment of the reninangiotensin system. Regul Pept. 1998;78(1–3):13–8.
- Rice GI, et al. Evaluation of angiotensin-converting enzyme (ACE), its homologue ACE2 and neprilysin in angiotensin peptide metabolism. Biochem J. 2004;383(1):45–51.
- 26. Patel VB, et al. Recombinant human ACE2 and the angiotensin 1–7 axis as potential new therapies for heart failure. Can J Cardiol. 2017;33(7):943–6.
- 27. Patel VB, et al. Role of the ACE2/angiotensin 1–7 axis of the renin–angiotensin system in heart failure. Circul Res. 2016;118(8):1313–26.
- Yugandhar VG, Clark MA. Angiotensin III: a physiological relevant peptide of the renin angiotensin system. Peptides. 2013;46:26–32.
- Smith N, et al. What's new in the renin-angiotensin system? Cell Mol Life Sci CMLS. 2004;61(21):2695–703.
- Jankowski V, et al. Mass-spectrometric identification of a novel angiotensin peptide in human plasma. Arterioscler Thromb Vasc Biol. 2007;27(2):297–302.

- 31. Lautner RQ, et al. Discovery and characterization of alamandine: a novel component of the renin–angiotensin system. Circul Res. 2013;112(8):1104–11.
- Hrenak J, Paulis L, Simko F. Angiotensin A/Alamandine/MrgD axis: another clue to understanding cardiovascular pathophysiology. Int J Mol Sci. 2016;17(7):1098.
- Rabelo LA, Alenina N, Bader M. ACE2–angiotensin-(1–7)–Mas axis and oxidative stress in cardiovascular disease. Hypertens Res. 2011;34(2):154–60.
- Tikellis C, Thomas M. Angiotensin-converting enzyme 2 (ACE2) is a key modulator of the renin angiotensin system in health and disease. International journal of peptides, 2012. 2012.
- Iwata M, et al. Targeting the ACE2–Ang-(1–7) pathway in cardiac fibroblasts to treat cardiac remodeling and heart failure. J Mol Cell Cardiol. 2011;51(4):542–7.
- Santos RAS, et al. The renin-angiotensin system: going beyond the classical paradigms. American Journal of Physiology-Heart and Circulatory Physiology; 2019.
- Iwai M, Horiuchi M. Devil and angel in the renin–angiotensin system: ACE– angiotensin II–AT1 receptor axis vs. ACE2–angiotensin-(1–7)–Mas receptor axis. Hypertens Res. 2009;32(7):533–6.
- Alenina N, et al. Genetically altered animal models for mas and angiotensin-(1–7). Exp Physiol. 2008;93(5):528–37.
- Sampaio WO, et al. Angiotensin-(1–7) through receptor mas mediates endothelial nitric oxide synthase activation via akt-dependent pathways. Hypertension. 2007;49(1):185–92.
- Santos RA et al. Angiotensin-(1–7) is an endogenous ligand for the G proteincoupled receptor Mas. Proceedings of the National Academy of Sciences, 2003. 100(14): p. 8258–8263.
- Tallant EA, Ferrario CM, Gallagher PE. Angiotensin-(1–7) inhibits growth of cardiac myocytes through activation of the mas receptor. Am J Physiol Heart Circ Physiol. 2005;289(4):H1560–6.
- 42. Hammer A, et al. Role of the receptor mas in macrophage-mediated inflammation in vivo. Proc Natl Acad Sci. 2016;113(49):14109–14.
- Santos RAS et al. The ACE2/angiotensin-(1–7)/MAS axis of the renin-angiotensin system: focus on angiotensin-(1–7). Physiological reviews, 2017.
- 44. Xu P, et al. Endothelial dysfunction and elevated blood pressure in MAS genedeleted mice. Hypertension. 2008;51(2):574–80.
- Trerattanavong K, Chen JS. Biochemistry, renin, in StatPearls [Internet]. Stat-Pearls Publishing; 2022.
- Vargas RAV, Millán JMV, Bonilla EF. Renin–angiotensin system: Basic and clinical aspects—A general perspective. Endocrinología, Diabetes y Nutrición (English ed.), 2022. 69(1): p. 52–62.
- 47. Re RN. Cellular biology of the renin-angiotensin systems. Arch Intern Med. 1984;144(10):2037–41.
- Wang J, et al. The (pro) renin receptor: a novel biomarker and potential therapeutic target for various cancers. Cell Communication and Signaling. 2020;18(1):1–13.
- Ichihara A, Yatabe MS. The (pro) renin receptor in health and disease. Nat Rev Nephrol. 2019;15(11):693–712.
- Nguyen G, et al. Pivotal role of the renin/prorenin receptor in angiotensin II production and cellular responses to renin. J Clin Investig. 2002;109(11):1417–27.
- 51. Burcklé C, Bader M. Prorenin and its ancient receptor. 2006, Am Heart Assoc. p. 549–51.
- Cruciat C-M, et al. Requirement of prorenin receptor and vacuolar H+-ATPase-mediated acidification for wnt signaling. Science. 2010;327(5964):459–63.
- 53. Ohba K, et al. Expression of (pro) renin receptor in breast cancers and its effect on cancercell proliferation. Biomed Res. 2014;35(2):117–26.
- 54. Kouchi M, et al. (Pro) renin receptor is crucial for glioma development via the Wnt/ β -catenin signaling pathway. J Neurosurg. 2017;127(4):819–28.
- Shibayama Y, et al. (Pro) renin receptor is crucial for Wnt/β-catenindependent genesis of pancreatic ductal adenocarcinoma. Sci Rep. 2015;5(1):1–9.
- Arundhathi A, et al. Prorenin receptor acts as a potential molecular target for pancreatic ductal adenocarcinoma diagnosis. Oncotarget. 2016;7(34):55437.
- 57. Yamamoto H, et al. Increased expression of (pro) renin receptor in aldosterone-producing adenomas. Peptides. 2013;49:68–73.
- Wang J, et al. (pro) renin receptor promotes colorectal cancer through the Wnt/beta-catenin signalling pathway despite constitutive pathway component mutations. Br J Cancer. 2019;120(2):229–37.
- Turner AJ, Hooper NM. The angiotensin–converting enzyme gene family: genomics and pharmacology. Trends Pharmacol Sci. 2002;23(4):177–83.

- Donoghue M, et al. A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1–9. Circul Res. 2000;87(5):e1–e9.
- 62. Wang K, Gheblawi M, Oudit GY. Angiotensin converting enzyme 2: a doubleedged sword. Circulation. 2020;142(5):426–8.
- Kambayashi Y, et al. Molecular cloning of a novel angiotensin II receptor isoform involved in phosphotyrosine phosphatase inhibition. J Biol Chem. 1993;268(33):24543–6.
- Mukoyama M, et al. Expression cloning of type 2 angiotensin II receptor reveals a unique class of seven-transmembrane receptors. J Biol Chem. 1993;268(33):24539–42.
- 65. Carey RM. Cardiovascular and renal regulation by the angiotensin type 2 receptor: the AT2 receptor comes of age. Hypertension. 2005;45(5):840–4.
- 66. Reudelhuber TL. The continuing saga of the AT2 receptor: a case of the good, the bad, and the innocuous. Hypertension. 2005;46(6):1261–2.
- Steckelings UM, et al. The angiotensin AT2 receptor: from a binding site to a Novel Therapeutic Target. Pharmacol Rev. 2022;74(4):1051–135.
- Bosnyak S, et al. Relative affinity of angiotensin peptides and novel ligands at AT1 and AT2 receptors. Clin Sci. 2011;121(7):297–303.
- Matavelli LC, Siragy HM. AT2 receptor activities and pathophysiological implications. J Cardiovasc Pharmacol. 2015;65(3):226.
- Namsolleck P, et al. AT2 receptor and tissue injury: therapeutic implications. Curr Hypertens Rep. 2014;16(2):1–10.
- 71. Endo Y et al. Function of angiotensin II type 2 receptor in the postglomerular efferent arteriole. Kidney Int Supplement, 1997(63).
- 72. Jones A, et al. Genetic variants of angiotensin II receptors and cardiovascular risk in hypertension. Hypertension. 2003;42(4):500–6.
- Gembardt F, et al. Angiotensin metabolites can stimulate receptors of the Mas-related genes family. Mol Cell Biochem. 2008;319(1):115–23.
- 74. Villela DC, Passos-Silva DG, Santos RA. Alamandine: a new member of the angiotensin family. Curr Opin Nephrol Hypertens. 2014;23(2):130–4.
- Qaradakhi T, Apostolopoulos V, Zulli A. Angiotensin (1–7) and Alamandine: similarities and differences. Pharmacol Res. 2016;111:820–6.
- Sevá Pessôa B, et al. Key developments in renin–angiotensin–aldosterone system inhibition. Nat Rev Nephrol. 2013;9(1):26–36.
- Chauhan VP, et al. Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. Nat Commun. 2013;4(1):2516.
- Afsar B, et al. Renin-angiotensin system and cancer: epidemiology, cell signaling, genetics and epigenetics. Clin Transl Oncol. 2021;23(4):682–96.
- Fujita M, et al. Angiotensin type 1a receptor signaling-dependent induction of vascular endothelial growth factor in stroma is relevant to tumor-associated angiogenesis and tumor growth. Carcinogenesis. 2005;26(2):271–9.
- Cheng N, et al. Targeting tumor-associated macrophages as an antitumor strategy. Biochem Pharmacol. 2021;183:114354.
- Acconcia F. The network of angiotensin receptors in breast cancer. Cells. 2020;9(6):1336.
- Delforce SJ, et al. Expression of renin–angiotensin system (RAS) components in endometrial cancer. Endocr Connections. 2017;6(1):9.
- Deshayes F, Nahmias C. Angiotensin receptors: a new role in cancer? Trends in Endocrinology & Metabolism. 2005;16(7):293–9.
- Yang X, et al. Angiotensin II promotes smooth muscle cell proliferation and migration through release of heparin-binding epidermal growth factor and activation of EGF-receptor pathway. Volume 20. Molecules & Cells (Springer Science & Business Media BV); 2005. 2.
- Uemura H, et al. Antiproliferative activity of angiotensin II receptor blocker through cross-talk between stromal and epithelial prostate cancer cells. Mol Cancer Ther. 2005;4(11):1699–709.
- Uemura H, et al. Angiotensin II receptor blocker shows antiproliferative activity in prostate cancer cells: a possibility of tyrosine kinase inhibitor of growth factor. Mol Cancer Ther. 2003;2(11):1139–47.
- Kinoshita J, et al. Local angiotensin II-generation in human gastric cancer: correlation with tumor progression through the activation of ERK1/2, NF-κB and survivin. Int J Oncol. 2009;34(6):1573–82.
- Ohashi H, et al. Phosphatidylinositol 3-kinase/Akt regulates angiotensin Il-induced inhibition of apoptosis in microvascular endothelial cells by governing survivin expression and suppression of caspase-3 activity. Circul Res. 2004;94(6):785–93.

- Tharaux P-L, et al. Angiotensin II activates collagen I gene through a mechanism involving the MAP/ER kinase pathway. Hypertension. 2000;36(3):330–6.
- Greco S, et al. Angiotensin II activates extracellular signal regulated kinases via protein kinase C and epidermal growth factor receptor in breast cancer cells. J Cell Physiol. 2003;196(2):370–7.
- Fujiyama S, et al. Angiotensin AT1 and AT2 receptors differentially regulate angiopoietin-2 and vascular endothelial growth factor expression and angiogenesis by modulating heparin binding–epidermal growth factor (EGF)–mediated EGF receptor transactivation. Circul Res. 2001;88(1):22–9.
- Anandanadesan R, et al. Angiotensin II induces vascular endothelial growth factor in pancreatic cancer cells through an angiotensin II type 1 receptor and ERK1/2 signaling. J Gastrointest Surg. 2008;12(1):57–66.
- 93. Kosugi M, et al. Effect of angiotensin II type 1 receptor antagonist on tumor growth and angiogenesis in a xenograft model of human bladder cancer. Hum Cell. 2007;20(1):1–9.
- Fan F, et al. Candesartan attenuates angiogenesis in hepatocellular carcinoma via downregulating AT1R/VEGF pathway. Biomed Pharmacother. 2016;83:704–11.
- Suganuma T, et al. Functional expression of the angiotensin II type1 receptor in human ovarian carcinoma cells and its blockade therapy resulting in suppression of tumor invasion, angiogenesis, and peritoneal dissemination. Clin Cancer Res. 2005;11(7):2686–94.
- 96. Laghlam D, Jozwiak M, Nguyen LS. Renin–angiotensin–aldosterone system and immunomodulation: a state-of-the-art review. Cells. 2021;10(7):1767.
- Zhao Y, et al. Ang II–AT1R increases cell migration through PI3K/AKT and NF-κB pathways in breast cancer. J Cell Physiol. 2014;229(11):1855–62.
- Bose SK, et al. Angiotensin II up-regulates PAX2 oncogene expression and activity in prostate cancer via the angiotensin II type I receptor. Prostate. 2009;69(12):1334–42.
- 99. Teranishi Ji, et al. Evaluation of role of angiotensin III and aminopeptidases in prostate cancer cells. Prostate. 2008;68(15):1666–73.
- 100. Perini MV, et al. Evaluating the benefits of renin-angiotensin system inhibitors as cancer treatments. Pharmacol Ther. 2020;211:107527.
- 101. Takahashi M, et al. Angiotensin II and tumor necrosis factor-α synergistically promote monocyte chemoattractant protein-1 expression: roles of NF-κB, p38, and reactive oxygen species. Am J Physiol Heart Circ Physiol. 2008;294(6):H2879–88.
- 102. Hoesel B, Schmid JA. The complexity of NF-kB signaling in inflammation and cancer. Mol Cancer. 2013;12(1):1–15.
- 103. Ishikane S, Takahashi-Yanaga F. The role of angiotensin II in cancer metastasis: potential of renin-angiotensin system blockade as a treatment for cancer metastasis. Biochem Pharmacol. 2018;151:96–103.
- 104. Rasha F, et al. Mechanisms linking the renin-angiotensin system, obesity, and breast cancer. Endocrine-related Cancer. 2019;26(12):R653–72.
- Gelosa P et al. Repurposing of drugs approved for cardiovascular diseases: opportunity or mirage? Biochemical pharmacology, 2020. 177: p. 113895.
- Buharalioglu CK, et al. Angiotensin II-induced process of angiogenesis is mediated by spleen tyrosine kinase via VEGF receptor-1 phosphorylation. Am J Physiol Heart Circ Physiol. 2011;301(3):H1043–55.
- 107. Daemen M, et al. Angiotensin II induces smooth muscle cell proliferation in the normal and injured rat arterial wall. Circul Res. 1991;68(2):450–6.
- Muscella A, et al. PKC-ζ is required for angiotensin II-induced activation of ERK and synthesis of C-FOS in MCF-7 cells. J Cell Physiol. 2003;197(1):61–8.
- Isobe A, et al. Dual repressive effect of angiotensin II-type 1 receptor blocker telmisartan on angiotensin II-induced and estradiol-induced uterine leiomyoma cell proliferation. Hum Reprod. 2008;23(2):440–6.
- 110. Itabashi H, et al. Angiotensin II and epidermal growth factor receptor cross-talk mediated by a disintegrin and metalloprotease accelerates tumor cell proliferation of hepatocellular carcinoma cell lines. Hepatol Res. 2008;38(6):601–13.
- 111. George AJ, Thomas WG, Hannan RD. The renin–angiotensin system and cancer: old dog, new tricks. Nat Rev Cancer. 2010;10(11):745–59.
- Mehta PK, Griendling KK. Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. Am J Physiology-Cell Physiol. 2007;292(1):C82–C97.
- 113. Li X-Y, Sun J-F, Hu S-Q. The renin-angiotensin system blockers as adjunctive therapy for cancer: a meta-analysis of survival outcome. Volume 21. European Review for Medical & Pharmacological Sciences; 2017. 6.
- 114. Mohammad AH, et al. V-ATPase-associated prorenin receptor is upregulated in prostate cancer after PTEN loss. Oncotarget. 2019;10(48):4923.

- Wang J, et al. The (pro) renin receptor: a novel biomarker and potential therapeutic target for various cancers. Cell Communication and Signaling. 2020;18:1–13.
- 117. Krop M, et al. The (pro) renin receptor. A decade of research: what have we learned? Pflügers. Archiv-European J Physiol. 2013;465:87–97.
- 118. Huang Y, et al. Renin increases mesangial cell transforming growth factor-β1 and matrix proteins through receptor-mediated, angiotensin II-independent mechanisms. Kidney Int. 2006;69(1):105–13.
- 119. Huang Y, et al. Renin-stimulated TGF-β1 expression is regulated by a mitogenactivated protein kinase in mesangial cells. Kidney Int. 2007;72(1):45–52.
- Peng H, et al. (Pro) renin receptor mediates both angiotensin II-dependent and-independent oxidative stress in neuronal cells. PLoS ONE. 2013;8(3):e58339.
- 121. Howe LR, Brown AM. Wnt signaling and breast cancer. Cancer Biol Ther. 2004;3(1):36–41.
- 122. Kurayoshi M, et al. Expression of Wnt-5a is correlated with aggressiveness of gastric cancer by stimulating cell migration and invasion. Cancer Res. 2006;66(21):10439–48.
- 123. Gupta S, et al. FZD4 as a mediator of ERG Oncogene–Induced WNT signaling and epithelial-to-mesenchymal transition in human prostate Cancer CellsFZD4 as a mediator of ERG-Induced WNT signaling. Cancer Res. 2010;70(17):6735–45.
- 124. de Sousa EMF, et al. Targeting wnt signaling in colon cancer stem cells. Clin Cancer Res. 2011;17(4):647–53.
- 125. El Wakil A, Lalli E. The Wnt/beta-catenin pathway in adrenocortical development and cancer. Mol Cell Endocrinol. 2011;332(1–2):32–7.
- 126. Khoshghamat N, et al. The therapeutic potential of renin-angiotensin system inhibitors in the treatment of pancreatic cancer. Life Sci. 2021;270:119118.
- 127. Catarata MJ, et al. Renin-angiotensin system in lung tumor and microenvironment interactions. Cancers. 2020;12(6):1457.
- 128. Datzmann T, et al. Systematic review and meta-analysis of randomised controlled clinical trial evidence refutes relationship between pharmacotherapy with angiotensin-receptor blockers and an increased risk of cancer. Eur J Intern Med. 2019;64:1–9.
- 129. Pei N, et al. Angiotensin II type 2 receptor promotes apoptosis and inhibits angiogenesis in bladder cancer. J Experimental Clin Cancer Res. 2017;36(1):1–12.
- 130. Machado RD, Santos RA, Andrade SP. Opposing actions of angiotensins on angiogenesis. Life Sci. 1999;66(1):67–76.
- 131. Benndorf R, et al. Angiotensin II type 2 receptor inhibits vascular endothelial growth factor–induced migration and in vitro tube formation of human endothelial cells. Circul Res. 2003;93(5):438–47.
- 132. Qi Y, et al. Moderate cardiac-selective overexpression of angiotensin II type 2 receptor protects cardiac functions from ischaemic injury. Exp Physiol. 2012;97(1):89–101.
- 133. Sun L, et al. Angiotensin II induces apoptosis in intestinal epithelial cells through the AT2 receptor, GATA-6 and the bax pathway. Biochem Biophys Res Commun. 2012;424(4):663–8.
- 134. Pickel L, et al. Over-expression of angiotensin II type 2 receptor gene induces cell death in lung adenocarcinoma cells. Cancer Biol Ther. 2010;9(4):277–85.
- 135. Kawabata A, et al. Intratracheal administration of a nanoparticle-based therapy with the angiotensin II type 2 receptor gene attenuates lung cancer growth. Cancer Res. 2012;72(8):2057–67.
- 136. Pei N, et al. Angiotensin II type 2 receptor promotes apoptosis and inhibits angiogenesis in bladder cancer. J Experimental Clin Cancer Res. 2017;36:1–12.
- Li H, et al. Angiotensin type 2 receptor–mediated apoptosis of human prostate cancer cellsAT2R-Induced apoptosis of prostate Cancer cells. Mol Cancer Ther. 2009;8(12):3255–65.
- Martínez-Meza S, et al. AT2 receptor mediated activation of the Tyrosine phosphatase PTP1B blocks Caveolin-1 enhanced Migration, Invasion and Metastasis of Cancer cells. Cancers. 2019;11(9):1299.
- 139. Murphy AM, Wong AL, Bezuhly M. Modulation of angiotensin II signaling in the prevention of fibrosis. Fibrogenesis & Tissue Repair. 2015;8:1–7.
- 140. Simões e, Silva A, et al. ACE2, angiotensin-(1-7) and M as receptor axis in inflammation and fibrosis. Br J Pharmacol. 2013;169(3):477–92.
- 141. Chen Y-H, et al. The clinical impact of angiotensin-(1–7)/mitochondrial assembly receptor axis in esophageal squamous cell carcinoma patients receiving curative esophagectomy. J Formos Med Assoc. 2020;119(1):310–8.

- Lin Y-T, et al. Pre-treatment with angiotensin-(1–7) inhibits tumor growth via autophagy by downregulating PI3K/Akt/mTOR signaling in human nasopharyngeal carcinoma xenografts. J Mol Med. 2018;96(12):1407–18.
- 143. Nakamura K, et al. Involvement of local renin-angiotensin system in immunosuppression of tumor microenvironment. Cancer Sci. 2018;109(1):54–64.
- 144. Domińska K, et al. Angiotensin 1–7 modulates molecular and cellular processes central to the pathogenesis of prostate cancer. Sci Rep. 2018;8(1):1–12.
- 145. Xu J, et al. The ACE2/angiotensin-(1–7)/mas receptor axis: pleiotropic roles in cancer. Front Physiol. 2017;8:276.
- Gallagher PE, Tallant EA. Inhibition of human lung cancer cell growth by angiotensin-(1–7). Carcinogenesis. 2004;25(11):2045–52.
- 147. Lin Y-T, et al. Pre-treatment with angiotensin-(1–7) inhibits tumor growth via autophagy by downregulating PI3K/Akt/mTOR signaling in human nasopharyngeal carcinoma xenografts. J Mol Med. 2018;96:1407–18.
- Cambados N, et al. Angiotensin-(1–7) counteracts the transforming effects triggered by angiotensin II in breast cancer cells. Oncotarget. 2017;8(51):88475.
- Nouet S, et al. Trans-inactivation of receptor tyrosine kinases by novel angiotensin II AT2 receptor-interacting protein, ATIP. J Biol Chem. 2004;279(28):28989–97.
- Rodrigues-Ferreira S, et al. 8p22 MTUS1 gene product ATIP3 is a novel anti-mitotic protein underexpressed in invasive breast carcinoma of poor prognosis. PLoS ONE. 2009;4(10):e7239.
- 151. Zhu P, et al. Expression and function of Mas-related G protein-coupled receptor D and its ligand alamandine in retina. Mol Neurobiol. 2020;57:513–27.
- Soltani Hekmat A, Javanmardi K. Alamandine: Potential Protective Effects in SARS-CoV-2 Patients. Journal of the Renin-Angiotensin-Aldosterone System, 2021. 2021; p. 1–9.
- Uchiyama T, et al. Alamandine reduces leptin expression through the c-Src/ p38 MAP kinase pathway in adipose tissue. PLoS ONE. 2017;12(6):e0178769.
- 154. Qaradakhi T, et al. Alamandine reverses hyperhomocysteinemia-induced vascular dysfunction via PKA-dependent mechanisms. Cardiovasc Ther. 2017;35(6):e12306.
- 155. Paz Ocaranza M, et al. Counter-regulatory renin–angiotensin system in cardiovascular disease. Nat Reviews Cardiol. 2020;17(2):116–29.
- Jesus ICGd, et al. Alamandine acts via MrgD to induce AMPK/NO activation against ANG II hypertrophy in cardiomyocytes. Am J Physiology-Cell Physiol. 2018;314(6):C702–11.
- Li P, et al. Alamandine attenuates sepsis-associated cardiac dysfunction via inhibiting MAPKs signaling pathways. Life Sci. 2018;206:106–16.
- Song X, Feng J, Yang R. Alamandine protects rat from myocardial ischemiareperfusion injury by activating JNK and inhibiting NF-κB. Eur Rev Med Pharmacol Sci. 2019;23(15):6718–26.
- 159. da Silva FA, et al. Phosphoproteomic studies of alamandine signaling in CHO-MrgD and human pancreatic carcinoma cells: an antiproliferative effect is unveiled. Proteomics. 2022;22(17):2100255.
- Levin A, Stevens PE. Summary of KDIGO 2012 CKD Guideline: behind the scenes, need for guidance, and a framework for moving forward. Kidney Int. 2014;85(1):49–61.
- 161. James PA, et al. 2014 evidence-based guideline for the management of high blood pressure in adults: report from the panel members appointed to the Eighth Joint National Committee (JNC 8). JAMA. 2014;311(5):507–20.
- 162. O'gara PT, et al. 2013 ACCF/AHA guideline for the management of STelevation myocardial infarction: executive summary: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. Circulation. 2013;127(4):529–55.
- 163. Cronin-Fenton D. Angiotensin converting enzyme inhibitors and lung cancer. British Medical Journal Publishing Group; 2018.
- 164. Shen J, et al. Renin–angiotensin system blockade for the risk of cancer and death. J Renin-Angiotensin-Aldosterone Syst. 2016;17(3):1470320316656679.
- 165. Yu C, et al. Downregulation of ACE2/Ang-(1–7)/Mas axis promotes breast cancer metastasis by enhancing store-operated calcium entry. Cancer Lett. 2016;376(2):268–77.
- 166. Luo Y, et al. Expression of MAS 1 in breast cancer. Cancer Sci. 2015;106(9):1240–8.
- 167. Diop-Frimpong B et al. Losartan inhibits collagen I synthesis and improves the distribution and efficacy of nanotherapeutics in tumors. Proceedings of the National Academy of Sciences, 2011. 108(7): p. 2909–2914.
- Rhodes DR et al. AGTR1 overexpression defines a subset of breast cancer and confers sensitivity to losartan, an AGTR1 antagonist. Proceedings of the National Academy of Sciences, 2009. 106(25): p. 10284–10289.

- Alhusban A, et al. Clinically relevant doses of candesartan inhibit growth of prostate tumor xenografts in vivo through modulation of tumor angiogenesis. J Pharmacol Exp Ther. 2014;350(3):635–45.
- 171. Ishiguro H, et al. Regulation of prostate cancer cell growth and PSA expression by angiotensin II receptor blocker with peroxisome proliferator-activated receptor gamma ligand like action. Prostate. 2007;67(9):924–32.
- 172. Takahashi S, et al. Therapeutic targeting of angiotensin II receptor type 1 to regulate androgen receptor in prostate cancer. Prostate. 2012;72(14):1559–72.
- 173. Morris ZS, et al. Increased tumor response to neoadjuvant therapy among rectal cancer patients taking angiotensin-converting enzyme inhibitors or angiotensin receptor blockers. Cancer. 2016;122(16):2487–95.
- Li X, Sun J, Hu S. The renin-angiotensin system blockers as adjunctive therapy for cancer: a meta-analysis of survival outcome. Eur Rev Med Pharmacol Sci. 2017;21(6):1375–83.
- 175. Stewart C, Ralyea C, Lockwood S. Ovarian cancer: an integrated review. Seminars in oncology nursing. 2019. Elsevier.
- 176. Kreienbring K et al. Predictive and prognostic value of sPRR in patients with primary epithelial ovarian cancer. Analytical cellular pathology, 2016. 2016.
- Beyazit F, et al. Assessment of serum angiotensin-converting enzyme in patients with epithelial ovarian cancer. Arch Gynecol Obstet. 2015;292:415–20.
- 178. Park Y-A, et al. Dual targeting of angiotensin receptors (AGTR1 and AGTR2) in epithelial ovarian carcinoma. Gynecol Oncol. 2014;135(1):108–17.
- Salvi A, et al. PAX8 modulates the tumor microenvironment of high grade serous ovarian cancer through changes in the secretome. Neoplasia. 2023;36:100866.
- 180. Hardy LR, et al. Proteomic analysis reveals a role for PAX8 in peritoneal colonization of high grade serous ovarian cancer that can be targeted with micelle encapsulated thiostrepton. Oncogene. 2019;38(32):6003–16.
- Almutlaq M, et al. The effect of local renin angiotensin system in the common types of cancer. Front Endocrinol. 2021;12:736361.
- Lengyel E. Ovarian cancer development and metastasis. Am J Pathol. 2010;177(3):1053–64.
- 183. Zhang Q, et al. Angiotensin II promotes ovarian cancer spheroid formation and metastasis by upregulation of lipid desaturation and suppression of endoplasmic reticulum stress. J Experimental Clin Cancer Res. 2019;38:1–18.
- 184. Zhao Y et al. Losartan treatment enhances chemotherapy efficacy and reduces ascites in ovarian cancer models by normalizing the tumor stroma. Proceedings of the National Academy of Sciences, 2019. 116(6): p. 2210–2219.
- Li J, et al. Adenovirus-mediated angiotensin II type 2 receptor overexpression inhibits tumor growth of prostate cancer in vivo. J Cancer. 2016;7(2):184.
- 186. Ito Y, et al. Chemopreventive effects of angiotensin II receptor type 2 agonist on prostate carcinogenesis by the down-regulation of the androgen receptor. Oncotarget. 2018;9(17):13859.
- Domińska K, et al. Interaction between angiotensin II and relaxin 2 in the progress of growth and spread of prostate cancer cells. Int J Oncol. 2016;48(6):2619–28.
- Kowalska K, et al. Coexpression of CAV-1, AT1-R and FOXM1 in prostate and breast cancer and normal cell lines and their influence on metastatic properties. Acta Biochim Pol. 2016;63(3):493–9.
- Domińska K, et al. A common effect of angiotensin II and relaxin 2 on the PNT1A normal prostate epithelial cell line. J Physiol Biochem. 2016;72:381–92.
- 190. Pai P-Y, et al. Long term antihypertensive drug use and prostate cancer risk: a 9-year population-based cohort analysis. Int J Cardiol. 2015;193:1–7.
- 191. Woo Y, Jung YJ. Angiotensin II receptor blockers induce autophagy in prostate cancer cells. Oncol Lett. 2017;13(5):3579–85.
- 192. Domińska K, et al. Effects of testosterone and 17β–estradiol on angiotensin– induced changes in tyrosine kinase activity in the androgen–independent human prostate cancer cell line, DU145. Int J Mol Med. 2017;40(5):1573–81.
- Domińska K, et al. The opposite effects of angiotensin 1–9 and angiotensin 3–7 in prostate epithelial cells. Biochem Biophys Res Commun. 2019;519(4):868–73.
- 194. Domińska K, et al. The impact of Ang-(1–9) and Ang-(3–7) on the biological properties of prostate cancer cells by modulation of inflammatory and steroidogenesis pathway genes. Int J Mol Sci. 2020;21(17):6227.
- Vinson GP, Ho MM, Puddefoot JR. The distribution of angiotensin II type 1 receptors, and the tissue renin-angiotensin systems. Mol Med Today. 1995;1(1):35–9.

- 196. Tahmasebi M, et al. Localisation of renin-angiotensin system (RAS) components in breast. Br J Cancer. 2006;95(1):67–74.
- 197. Inwang ER, et al. Angiotensin II type 1 receptor expression in human breast tissues. Br J Cancer. 1997;75(9):1279–83.
- 198. Puddefoot JR, et al. The role of angiotensin II in the regulation of breast cancer cell adhesion and invasion. Endocr Relat Cancer. 2006;13(3):895–903.
- 199. Xie G, et al. Local angiotensin II contributes to tumor resistance to checkpoint immunotherapy. J Immunother Cancer. 2018;6(1):88.
- 200. Deshayes F, Nahmias C. Angiotensin receptors: a new role in cancer? Trends Endocrinol Metab. 2005;16(7):293–9.
- Zhao Y, et al. Angiotensin II/angiotensin II type I receptor (AT1R) signaling promotes MCF-7 breast cancer cells survival via PI3-kinase/Akt pathway. J Cell Physiol. 2010;225(1):168–73.
- 202. Egami K, et al. Role of host angiotensin II type 1 receptor in tumor angiogenesis and growth. J Clin Invest. 2003;112(1):67–75.
- Herr D, et al. Potential role of renin–angiotensin-system for tumor angiogenesis in receptor negative breast cancer. Gynecol Oncol. 2008;109(3):418–25.
- Huang W, et al. Angiotensin II type 1 receptor expression in human gastric cancer and induces MMP2 and MMP9 expression in MKN-28 cells. Dig Dis Sci. 2008;53(1):163–8.
- 205. Rodrigues-Ferreira S, et al. Angiotensin II facilitates breast cancer cell migration and metastasis. PLoS ONE. 2012;7(4):e35667.
- 206. Ma Y, et al. AGTR1 promotes lymph node metastasis in breast cancer by upregulating CXCR4/SDF-1α and inducing cell migration and invasion. Aging. 2019;11(12):3969–92.
- 207. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74.
- 208. Schwartsburd PM. Chronic inflammation as inductor of pro-cancer microenvironment: pathogenesis of dysregulated feedback control. Cancer Metastasis Rev. 2003;22(1):95–102.
- 209. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? Lancet. 2001;357(9255):539–45.
- 210. Pagé EL, et al. Induction of hypoxia-inducible factor-1alpha by transcriptional and translational mechanisms. J Biol Chem. 2002;277(50):48403–9.
- 211. Husain K, et al. Inflammation, oxidative stress and renin angiotensin system in atherosclerosis. World J Biol Chem. 2015;6(3):209–17.
- 212. Rasha F, et al. Mechanisms linking the renin-angiotensin system, obesity, and breast cancer. Endocr Relat Cancer. 2019;26(12):R653–r672.
- 213. Ekambaram P, et al. The CARMA3-Bcl10-MALT1 signalosome drives NFkB activation and promotes aggressiveness in angiotensin II receptor-positive breast Cancer. Cancer Res. 2018;78(5):1225–40.
- Liu F et al. Key Factor Regulating Inflammatory Microenvironment, Metastasis, and Resistance in Breast Cancer: Interleukin-1 Signaling. Mediators of Inflammation, 2021. 2021: p. 7785890.
- Hunyady L, Catt KJ. Pleiotropic AT1 receptor signaling pathways mediating physiological and pathogenic actions of angiotensin II. Mol Endocrinol. 2006;20(5):953–70.
- 216. Garrido AM, Griendling KK. NADPH oxidases and angiotensin II receptor signaling. Mol Cell Endocrinol. 2009;302(2):148–58.
- 217. Brown NS, Bicknell R. Hypoxia and oxidative stress in breast cancer oxidative stress - its effects on the growth, metastatic potential and response to therapy of breast cancer. Breast Cancer Res. 2001;3(5):323.
- 218. Salgado R, et al. The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: recommendations by an International TILs Working Group 2014. Ann Oncol. 2015;26(2):259–71.
- 219. Wu L et al. Tumor-Associated Neutrophils in Cancer: going pro. Cancers (Basel), 2019. 11(4).
- 220. Qiu SQ, et al. Tumor-associated macrophages in breast cancer: innocent bystander or important player? Cancer Treat Rev. 2018;70:178–89.
- 221. Tower H, Ruppert M, Britt K. The Immune Microenvironment of breast Cancer progression. Cancers (Basel), 2019. 11(9).
- Williams CB, Yeh ES, Soloff AC. Tumor-associated macrophages: unwitting accomplices in breast cancer malignancy. NPJ Breast Cancer. 2016;2:15025.
- 223. Joyce JA, Fearon DT.T cell exclusion, immune privilege, and the tumor microenvironment. Science. 2015;348(6230):74–80.
- 224. Noy R, Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. Immunity. 2014;41(1):49–61.
- Li JJ, Tsang JY, Tse GM. Tumor microenvironment in breast Cancer-updates on therapeutic implications and pathologic Assessment. Cancers (Basel), 2021. 13(16).

- 226. Arrieta O, et al. Association between AT1 and AT2 angiotensin II receptor expression with cell proliferation and angiogenesis in operable breast cancer. Tumor Biology. 2015;36(7):5627–34.
- 227. Smith GR, Missailidis S. Cancer, inflammation and the AT1 and AT2 receptors. J Inflamm (Lond). 2004;1(1):3.
- 228. Wegman-Ostrosky T, et al. The renin-angiotensin system meets the hallmarks of cancer. J Renin Angiotensin Aldosterone Syst. 2015;16(2):227–33.
- 229. De Paepe B, et al. Increased angiotensin II type-2 receptor density in hyperplasia, DCIS and invasive carcinoma of the breast is paralleled with increased iNOS expression. Histochem Cell Biol. 2002;117(1):13–9.
- Santos RA, Ferreira AJ, Simões ESAC. Recent advances in the angiotensin-converting enzyme 2-angiotensin(1–7)-Mas axis. Exp Physiol. 2008;93(5):519–27.
- Cambados N, et al. Angiotensin-(1–7) counteracts the transforming effects triggered by angiotensin II in breast cancer cells. Oncotarget. 2017;8(51):88475–87.
- 232. Zhang Q, et al. ACE2 inhibits breast cancer angiogenesis via suppressing the VEGFa/VEGFR2/ERK pathway. J Exp Clin Cancer Res. 2019;38(1):173.
- 233. Mei J, et al. Angiotensin-converting enzyme 2 identifies immuno-hot tumors suggesting angiotensin-(1–7) as a sensitizer for chemotherapy and immuno-therapy in breast cancer. Biol Proced Online. 2022;24(1):15.
- 234. Bujak-Gizycka B et al. Angiotensin 1–7 formation in breast tissue is attenuated in breast cancer - a study on the metabolism of angiotensinogen in breast cancer cell lines. J Physiol Pharmacol, 2019. 70(4).
- Rodrigues-Ferreira S, Nahmias C. G-protein coupled receptors of the reninangiotensin system: new targets against breast cancer? Front Pharmacol. 2015;6:24.
- Yang J, et al. The role of the renin-angiotensin system inhibitors in malignancy: a review. Am J Cancer Res. 2021;11(3):884–97.
- 237. Pinter M, Jain RK. Targeting the renin-angiotensin system to improve cancer treatment: implications for immunotherapy. Sci Transl Med, 2017. 9(410).
- 238. Raskov, H., et al., The Matrix Reloaded—The Role of the Extracellular Matrix in Cancer. Cancers, 2023. 15(7): p. 2057.
- 239. Xia T, et al. Losartan loaded liposomes improve the antitumor efficacy of liposomal paclitaxel modified with pH sensitive peptides by inhibition of collagen in breast cancer. Pharm Dev Technol. 2018;23(1):13–21.
- 240. Zhao Q et al. Enhanced therapeutic efficacy of combining Losartan and Chemo-Immunotherapy for Triple negative breast Cancer. Front Immunol, 2022. 13.
- 241. Xie Y et al. Association between antihypertensive medication use and breast Cancer: a systematic review and Meta-analysis. Front Pharmacol, 2021. 12.
- 242. Raimondi S, et al. Use of beta-blockers, angiotensin-converting enzyme inhibitors and angiotensin receptor blockers and breast cancer survival: systematic review and meta-analysis. Int J Cancer. 2016;139(1):212–9.
- 243. Murphy JE, et al. Total neoadjuvant therapy with FOLFIRINOX in combination with losartan followed by chemoradiotherapy for locally advanced pancreatic cancer: a phase 2 clinical trial. JAMA Oncol. 2019;5(7):1020–7.
- 244. Hirasawa K, et al. Immunohistochemical localization of angiotensin II receptor and local renin-angiotensin system in human colonic mucosa. J Histochem Cytochem. 2002;50(2):275–82.
- Mastropaolo M, et al. Activation of angiotensin II type 1 receptors and contractile activity in human sigmoid colon in vitro. Acta Physiol. 2015;215(1):37–45.
- Mastropaolo M, et al. Angiotensin II contractile effects in mouse colon: role for pre-and post-junctional AT 1A receptors. Acta Physiol. 2013;207(2):337–45.
- Valderrama-Treviño AI, et al. Hepatic metastasis from Colorectal Cancer. Euroasian J Hepatogastroenterol. 2017;7(2):166–75.
- 248. Shimomoto T, et al. Diabetes-associated angiotensin activation enhances liver metastasis of colon cancer. Clin Exp Metastasis. 2012;29(8):915–25.
- Zhou L, et al. Role of two types of angiotensin II receptors in colorectal carcinoma progression. Pathobiology. 2014;81(4):169–75.
- Beitia M, et al. Altered expression of renin-angiotensin system receptors throughout colorectal adenoma-adenocarcinoma sequence. Int J Med Sci. 2019;16(6):813–21.
- 251. Namsolleck P, de Vries L, Moll GN. LP2, a cyclic angiotensin-(1 7) analog extended with an N-terminal D-lysine, impairs growth of patient-derived xenografts of colorectal carcinoma in mice. Peptides. 2023;160:170920.
- Leung PS, et al. Expression and localization of AT1 receptors in hepatic kupffer cells: its potential role in regulating a fibrogenic response. Regul Pept. 2003;116(1):61–9.
- 253. Bernardi S, et al. Characterization and significance of ACE2 and mas receptor in human colon adenocarcinoma. J Renin Angiotensin Aldosterone Syst. 2012;13(1):202–9.

- 254. Oura K, et al. Telmisartan inhibits hepatocellular carcinoma cell proliferation in vitro by inducing cell cycle arrest. Oncol Rep. 2017;38(5):2825–35.
- 255. Wen SW, et al. The renin angiotensin system regulates Kupffer cells in colorectal liver metastases. Cancer Biol Ther. 2013;14(8):720–7.
- Asgharzadeh F, et al. Inhibition of angiotensin pathway via valsartan reduces tumor growth in models of colorectal cancer. Toxicol Appl Pharmacol. 2022;440:115951.
- Balkrishnan R, et al. Associations between initiating antihypertensive regimens on stage I–III colorectal cancer outcomes: a Medicare SEER cohort analysis. Cancer Med. 2021;10(15):5347–57.
- 258. Bueno V, Forones NM, Pawelec G. Alternative Chemotherapies: angiotensinconverting enzyme inhibitors reduce myeloid-derived suppressor cells to benefit older patients with colorectal Cancer. FBL, 2023. 28(1).
- 259. Engineer DR, et al. Exposure to ACEI/ARB and β-blockers is associated with improved survival and decreased tumor progression and hospitalizations in patients with advanced colon cancer. Translational Oncol. 2013;6(5):539–45.
- 260. Ozawa T, et al. Angiotensin I-converting enzyme inhibitors/angiotensin II receptor blockers may reduce tumor recurrence in left-sided and early colorectal cancers. Int J Colorectal Dis. 2019;34(10):1731–9.
- 261. Chow L, et al. Gastric acid secretion after blockade of angiotensin AT1 receptors in the na(+)-depleted rat. Eur J Pharmacol. 1995;294(1):309–17.
- Hallersund P, et al. The expression of renin-angiotensin system components in the human gastric mucosa. J Renin Angiotensin Aldosterone Syst. 2011;12(1):54–64.
- Carl-McGrath S, et al. Expression of the local angiotensin II system in gastric cancer may facilitate lymphatic invasion and nodal spread. Cancer Biol Ther. 2007;6(8):1218–26.
- 264. Huang MM, et al. Angiotensin II promotes the progression of human gastric cancer. Mol Med Rep. 2014;9(3):1056–60.
- 265. Kinoshita J, et al. Local angiotensin II-generation in human gastric cancer: correlation with tumor progression through the activation of ERK1/2, NFkappaB and survivin. Int J Oncol. 2009;34(6):1573–82.
- Okazaki M, et al. The angiotensin II type 1 receptor blocker candesartan suppresses proliferation and fibrosis in gastric cancer. Cancer Lett. 2014;355(1):46–53.
- 267. Röcken C, et al. The angiotensin II/angiotensin II receptor system correlates with nodal spread in intestinal type gastric cancer. Cancer Epidemiol Biomarkers Prev. 2007;16(6):1206–12.
- Carl-McGrath S, et al. Expression of the local angiotensin II system in gastric cancer may facilitate lymphatic invasion and nodal spread. Cancer Biol Ther. 2007;6(8):1229–37.
- Sandoval-Borquez AM, et al. Reduced angiotensin II type 2 receptor expression is Associated with gastric Cancer progression and enhanced gastric Cancer Cell Migration/Invasion. FASEB J. 2022;36:1.
- Lam KY, Leung PS. Regulation and expression of a renin-angiotensin system in human pancreas and pancreatic endocrine tumours. Eur J Endocrinol. 2002;146(4):567–72.
- 271. Leung PS. The physiology of a local renin-angiotensin system in the pancreas. J Physiol. 2007;580(Pt 1):31–7.
- Chappell MC, Jacobsen DW, Tallant EA. Characterization of angiotensin II receptor subtypes in pancreatic acinar AR42J cells. Peptides. 1995;16(4):741–7.
- Tahmasebi M, et al. The tissue renin-angiotensin system in human pancreas. J Endocrinol. 1999;161(2):317–22.
- Leung PS, Chappell MC. A local pancreatic renin-angiotensin system: endocrine and exocrine roles. Int J Biochem Cell Biol. 2003;35(6):838–46.
- 275. Leung PS. The peptide hormone angiotensin II: its new functions in tissues and organs. Curr Protein Pept Sci. 2004;5(4):267–73.
- 276. Sakurai T, et al. Involvement of angiotensin II and reactive oxygen species in pancreatic fibrosis. Pancreatology. 2011;11(2):7–13.
- 277. Fujimoto Y, et al. Angiotensin II type 1 receptor expression in human pancreatic cancer and growth inhibition by angiotensin II type 1 receptor antagonist. FEBS Lett. 2001;495(3):197–200.
- Guo R, et al. MicroRNA-410 functions as a tumor suppressor by targeting angiotensin II type 1 receptor in pancreatic cancer. IUBMB Life. 2015;67(1):42–53.
- 279. Huang C, Iovanna J, Santofimia-Castaño P. Targeting fibrosis: the Bridge that connects pancreatitis and pancreatic Cancer. Int J Mol Sci, 2021. 22(9).
- 280. Habisch H, et al. Interaction of stellate cells with pancreatic carcinoma cells. Cancers (Basel). 2010;2(3):1661–82.
- 281. Cancer P, Microenvironment T, Grippo MH PJ, editors. Pancreatic stellate cells and fibrosis. Trivandrum:Transworld Research Network;: India; 2012.

- 283. Sachse A, Wolf G. Angiotensin II–Induced reactive oxygen species and the kidney. J Am Soc Nephrol. 2007;18(9):2439–46.
- Sarantis P, et al. Pancreatic ductal adenocarcinoma: treatment hurdles, tumor microenvironment and immunotherapy. World J Gastrointest Oncol. 2020;12(2):173–81.
- 285. Ishiguro S, et al. Involvement of angiotensin II type 2 receptor (AT2R) signaling in human pancreatic ductal adenocarcinoma (PDAC): a novel AT2R agonist effectively attenuates growth of PDAC grafts in mice. Cancer Biol Ther. 2015;16(2):307–16.
- Wang Y, et al. Severe acute pancreatitis is associated with upregulation of the ACE2-angiotensin-(1–7)-Mas axis and promotes increased circulating angiotensin-(1–7). Pancreatology. 2012;12(5):451–7.
- 387. Yu X, et al. Angiotensin-converting enzyme 2-angiotensin (1–7)-Mas axis prevents pancreatic acinar cell inflammatory response via inhibition of the p38 mitogen-activated protein kinase/nuclear factor-κB pathway. Int J Mol Med. 2018;41(1):409–20.
- Liu F, et al. ACE2 expression in Pancreas May cause pancreatic damage after SARS-CoV-2 infection. Clin Gastroenterol Hepatol. 2020;18(9):2128–2130e2.
- Wang J, et al. The ACE2-Angiotensin-(1–7)-Mas Axis protects against pancreatic cell damage in Cell Culture. Pancreas. 2015;44(2):266–72.
- 290. Lu CL, et al. The angiotensin-converting enzyme 2/angiotensin (1–7)/Mas axis protects the function of pancreatic β cells by improving the function of islet microvascular endothelial cells. Int J Mol Med. 2014;34(5):1293–300.
- 291. Zhang F, et al. Antioxidant effect of angiotensin (1–7) in the protection of pancreatic β cell function. Mol Med Rep. 2016;14(3):1963–9.
- 292. Zhou T, et al. Irbesartan overcomes gemcitabine resistance in pancreatic cancer by suppressing stemness and iron metabolism via inhibition of the Hippo/YAP1/c-Jun axis. J Experimental Clin Cancer Res. 2023;42(1):1–27.
- 293. KANG MY, HWANG HK. Use of angiotensin inhibitor associated with longer survival in patients with pancreatic cancer undergoing curative resection. Annals of Hepato-Biliary-Pancreatic Surgery. 2022;26(1):S265–5.
- 294. Lopez CD, et al. Updates to NeoOPTIMIZE: an open-label, phase II trial and biomarker discovery platform to assess the efficacy of adaptive switching of modified FOLFIRINOX (mFFX) or gemcitabine/nab-paclitaxel (GA) as a neoadjuvant strategy for patients with resectable/borderline resectable and locally advanced unresectable pancreatic ductal adenocarcinoma (PDAC). J Clin Oncol. 2023;41(4suppl):TPS776–6.
- 295. Farazi PA, DePinho RA. Hepatocellular carcinoma pathogenesis: from genes to environment. Nat Rev Cancer. 2006;6(9):674–87.
- 296. Ye G, et al. The association of renin-angiotensin system genes with the progression of hepatocellular carcinoma. Biochem Biophys Res Commun. 2015;459(1):18–23.
- 297. Bataller R, et al. Liver fibrogenesis: a new role for the renin-angiotensin system. Antioxid Redox Signal. 2005;7(9–10):1346–55.
- 298. Hayden MR, et al. Possible mechanisms of local tissue renin-angiotensin system activation in the Cardiorenal metabolic syndrome and type 2 diabetes Mellitus. Cardiorenal Med. 2011;1(3):193–210.
- 299. Zhang X, et al. Tetramethylpyrazine inhibits angiotensin II-induced activation of hepatic stellate cells associated with interference of platelet-derived growth factor β receptor pathways. Febs j. 2014;281(12):2754–68.
- 300. Warner FJ, et al. Liver fibrosis: a balance of ACEs? Clin Sci. 2007;113(3):109-18.
- 301. de Moreira S, et al. The role of renin-angiotensin system modulation on treatment and prevention of liver diseases. Peptides. 2014;62:189–96.
- Koh SL, et al. Blockade of the renin-angiotensin system improves the early stages of liver regeneration and liver function. J Surg Res. 2013;179(1):66–71.
- 303. Ji Y, et al. Angiotensin II enhances proliferation and inflammation through AT1/PKC/NF-κB signaling pathway in Hepatocellular Carcinoma cells. Cell Physiol Biochem. 2016;39(1):13–32.
- Yoshiji H, et al. Suppression of renin-angiotensin system attenuates hepatocarcinogenesis via angiogenesis inhibition in rats. Anticancer Res. 2005;25(5):3335–40.
- Yoshiji H, et al. Impact of renin-angiotensin system in hepatocellular carcinoma. Curr Cancer Drug Targets. 2011;11(4):431–41.
- Qi R, et al. The AT1/Raf/ERK1/2 signaling pathway is involved in angiotensin II-enhanced proliferation of hepatic carcinoma cells. Neoplasma. 2019;66(1):83–91.
- Wu TT, et al. Reduction of PKC alpha decreases cell proliferation, migration, and invasion of human malignant hepatocellular carcinoma. J Cell Biochem. 2008;103(1):9–20.

- Wang X, et al. Regulation of phorbol ester-mediated TRAF1 induction in human colon cancer cells through a PKC/RAF/ERK/NF-kappaB-dependent pathway. Oncogene. 2004;23(10):1885–95.
- 309. Quan Y, et al. [miR-152 inhibits the epithelial-mesenchymal transition and renin-angiotensin system of human hepatocellular carcinoma cells by downregulating AGTR1]. Xi bao yu fen zi mian yi xue za zhi = Chinese. J Cell Mol Immunol. 2022;38(9):819–24.
- Ji Y, et al. Angiotensin II induces angiogenic factors production partly via AT1/JAK2/STAT3/SOCS3 signaling pathway in MHCC97H cells. Cell Physiol Biochem. 2012;29(5–6):863–74.
- 311. Feng LH, et al. Irbesartan inhibits metastasis by interrupting the adherence of tumor cell to endothelial cell induced by angiotensin II in hepatocellular carcinoma. Ann Transl Med. 2021;9(3):207.
- Sakurai T, Kudo M. Molecular Link between Liver Fibrosis and Hepatocellular Carcinoma. Liver Cancer. 2013;2(3–4):365–6.
- O'Rourke JM, et al. Carcinogenesis on the background of liver fibrosis: implications for the management of hepatocellular cancer. World J Gastroenterol. 2018;24(39):4436–47.
- Dhar D, et al. Mechanisms of liver fibrosis and its role in liver cancer. Experimental Biology and Medicine. 2020;245(2):96–108.
- Dhar D, et al. Mechanisms of liver fibrosis and its role in liver cancer. Exp Biol Med (Maywood). 2020;245(2):96–108.
- 316. Bataller R, et al. Angiotensin II induces contraction and proliferation of human hepatic stellate cells. Gastroenterology. 2000;118(6):1149–56.
- 317. Granzow M, et al. Angiotensin-II type 1 receptor-mediated Janus kinase 2 activation induces liver fibrosis. Hepatology. 2014;60(1):334–48.
- Bataller R, et al. Prolonged infusion of angiotensin II into normal rats induces stellate cell activation and proinflammatory events in liver. Am J Physiol Gastrointest Liver Physiol. 2003;285(3):G642–51.
- Yoshiji H, et al. Angiotensin-II type 1 receptor interaction is a major regulator for liver fibrosis development in rats. Hepatology. 2001;34(4 Pt 1):745–50.
- 320. Afroze SH, et al. Activation of the renin-angiotensin system stimulates biliary hyperplasia during cholestasis induced by extrahepatic bile duct ligation. Am J Physiol Gastrointest Liver Physiol. 2015;308(8):G691–701.
- 321. Caja L et al. TGF- β and the tissue microenvironment: relevance in fibrosis and Cancer. Int J Mol Sci, 2018. 19(5).
- 322. Zhang H-F, et al. The mechanisms of renin–angiotensin system in hepatocellular carcinoma: from the perspective of liver fibrosis, HCC cell proliferation, metastasis and angiogenesis, and corresponding protection measures. Biomed Pharmacother. 2021;141:111868.
- 323. Gu L et al. Angiotensin II receptor inhibition ameliorates liver fibrosis and enhances hepatocellular carcinoma infiltration by effector T cells. Proceedings of the National Academy of Sciences, 2023. 120(19): p. e2300706120.
- 324. Du H, et al. Effects of angiotensin II type 2 receptor overexpression on the growth of hepatocellular carcinoma cells in vitro and in vivo. PLoS ONE. 2013;8(12):e83754.
- 325. Dong F, et al. ACE2 negatively regulates the Warburg effect and suppresses hepatocellular carcinoma progression via reducing ROS-HIF1α activity. Int J Biol Sci. 2023;19(8):2613–29.
- 326. Soto-Pantoja DR, et al. Angiotensin-(1–7) inhibits tumor angiogenesis in human lung cancer xenografts with a reduction in vascular endothelial growth factor. Mol Cancer Ther. 2009;8(6):1676–83.
- 327. Krishnan B, et al. Angiotensin-(1–7) attenuates metastatic prostate cancer and reduces osteoclastogenesis. Prostate. 2013;73(1):71–82.
- 328. Liu Y, et al. Angiotensin-(1–7) suppresses Hepatocellular Carcinoma Growth and Angiogenesis via Complex interactions of angiotensin II type 1 receptor, angiotensin II type 2 receptor and mas receptor. Mol Med. 2015;21(1):626–36.
- Casselbrant A, et al. Angiotensin II receptors are expressed and functional in human esophageal mucosa. Am J Physiol Gastrointest Liver Physiol. 2009;297(5):G1019–27.
- 330. Casselbrant A, et al. Actions by angiotensin II on esophageal contractility in humans. Gastroenterology. 2007;132(1):249–60.
- Chen Y-H, et al. Prognostic impact of renin-angiotensin system blockade in esophageal squamous cell carcinoma. J Renin-Angiotensin-Aldosterone Syst. 2014;16(4):1185–92.
- 332. Fujihara S, et al. The angiotensin II type 1 receptor antagonist telmisartan inhibits cell proliferation and tumor growth of esophageal adenocarcinoma via the AMPKα/mTOR pathway in vitro and in vivo. Oncotarget. 2017;8(5):8536–49.
- 333. Li SH, et al. Angiotensin II type I receptor (AT1R) is an independent prognosticator of esophageal squamous cell carcinoma and promotes cells proliferation via mTOR activation. Oncotarget. 2016;7(41):67150–65.

- Chen YH, et al. Prognostic impact of renin-angiotensin system blockade in esophageal squamous cell carcinoma. J Renin Angiotensin Aldosterone Syst. 2015;16(4):1185–92.
- Wang Q, et al. Angiotensin-l converting enzyme inhibitors suppress angiogenesis and growth of esophageal carcinoma xenografts. Dis Esophagus. 2012;25(8):757–63.
- 336. Wang X, et al. Impact of angiotensin system inhibitors on esophageal cancer survival. Surg Open Sci. 2021;3:34–8.
- Schiffmann LM, et al. Tumor Microenvironment of Esophageal Cancer. Cancers. 2021;13(18):4678.
- 338. Chen YH, et al. The clinical impact of angiotensin-(1–7)/mitochondrial assembly receptor axis in esophageal squamous cell carcinoma patients receiving curative esophagectomy. J Formos Med Assoc. 2020;119(1 Pt 2):310–8.
- Ino K, et al. Angiotensin II type 1 receptor expression in ovarian cancer and its correlation with tumour angiogenesis and patient survival. Br J Cancer. 2006;94(4):552–60.
- Domińska K, et al. The influence of angiotensin peptides on survival and motility of human high-grade serous ovarian cancer cells in serum starvation conditions. Int J Mol Sci. 2021;23(1):52.
- Piastowska-Ciesielska AW, et al. Effect of an angiotensin II type 1 receptor blocker on caveolin-1 expression in prostate cancer cells. Archives of Medical Science. 2013;9(4):739–44.
- 342. Uemura H, et al. Renin-angiotensin system is an important factor in hormone refractory prostate cancer. Prostate. 2006;66(8):822–30.
- Krishnan B, et al. Angiotensin-(1-7) attenuates metastatic prostate cancer and reduces osteoclastogenesis. Prostate. 2013;73(1):71–82.
- 344. Krishnan B, et al. Angiotensin-(1-7) reduces proliferation and angiogenesis of human prostate cancer xenografts with a decrease in angiogenic factors and an increase in sFIt-1. Prostate. 2013;73(1):60–70.
- 345. Chen X, et al. Angiotensin II type 1 receptor antagonists inhibit cell proliferation and angiogenesis in breast cancer. Cancer Lett. 2013;328(2):318–24.
- Du N, et al. Angiotensin II receptor type 1 blockers suppress the cell proliferation effects of angiotensin II in breast cancer cells by inhibiting AT1R signaling. Oncol Rep. 2012;27(6):1893–903.
- 347. Bakhtiari E, et al. Angiotensin II receptor antagonist olmesartan and NFkappaB inhibitor as cytotoxic and apoptotic agents in MCF-7 human cell line. J Chemother. 2016;28(4):314–20.
- Coulson R, et al. The angiotensin receptor blocker, Losartan, inhibits mammary tumor development and progression to invasive carcinoma. Oncotarget. 2017;8(12):18640–56.
- 349. Namazi S, et al. The role of captopril and losartan in prevention and regression of tamoxifen-induced resistance of breast cancer cell line MCF-7: an in vitro study. Biomed Pharmacother. 2014;68(5):565–71.
- 350. Rasha F, et al. Renin angiotensin system inhibition attenuates adipocytebreast cancer cell interactions. Exp Cell Res. 2020;394(1):112114.
- 351. Napoleone E, et al. Inhibition of the renin-angiotensin system downregulates tissue factor and vascular endothelial growth factor in human breast carcinoma cells. Thromb Res. 2012;129(6):736–42.
- 352. Patel SS, Nakka S. Protective effect of Perindopril on Tumor Progression and Angiogenesis in animal model of breast Cancer. Anticancer Agents Med Chem. 2017;17(7):955–60.
- 353. Ni H, et al. Antihypertensive drug use and breast cancer risk: a meta-analysis of observational studies. Oncotarget. 2017;8(37):62545–60.
- Goldvaser H, et al. The association between angiotensin receptor blocker usage and breast Cancer characteristics. Oncology. 2016;91(4):217–23.
- 355. Riddiough GE, et al. Captopril, a renin-angiotensin system inhibitor, attenuates features of Tumor Invasion and Down-Regulates C-Myc expression in a mouse model of Colorectal Cancer Liver Metastasis. Cancers. 2021;13(11):2734.
- 356. Tabatabai E, et al. Inhibition of angiotensin II type 1 receptor by candesartan reduces tumor growth and ameliorates fibrosis in colorectal cancer. Excli j. 2021;20:863–78.

- 357. Asgharzadeh F et al. Inhibition of angiotensin II type I pathway reduced tumor growth and ameliorates fibrosis/inflammation associated with colorectal cancer. 2020.
- Hashemzehi M, et al. Angiotensin receptor blocker Losartan inhibits tumor growth of colorectal cancer. Excli j. 2021;20:506–21.
- 359. Cheung KS, et al. ACE (angiotensin-Converting enzyme) Inhibitors/Angiotensin receptor blockers are Associated with Lower Colorectal Cancer risk: a territory-wide study with propensity score analysis. Hypertension. 2020;76(3):968–75.
- Huang W, et al. Angiotensin II type 1 receptor antagonist suppress angiogenesis and growth of gastric Cancer xenografts. Dig Dis Sci. 2008;53(5):1206–10.
- 361. Lee S-H, et al. Renin-angiotensin-aldosterone system inhibitors and risk of Cancer: a Population-Based Cohort Study using a Common Data Model. Diagnostics. 2022;12(2):263.
- Kumar V, et al. Noninvasive Assessment of Losartan-Induced increase in functional microvasculature and drug delivery in pancreatic ductal adenocarcinoma. Transl Oncol. 2016;9(5):431–7.
- Kim S, et al. Antitumor effect of angiotensin II type 1 receptor blocker losartan for orthotopic rat pancreatic adenocarcinoma. Pancreas. 2014;43(6):886–90.
- 364. Chen X, et al. Cancer-Associated fibroblast-targeted delivery of Captopril to overcome penetration obstacles for enhanced pancreatic Cancer therapy. ACS Appl Bio Mater. 2022;5(7):3544–53.
- 365. Yamana Y, et al. MicroRNA profiles following telmisartan treatment in pancreatic ductal adenocarcinoma cells. J Cancer Res Ther. 2022;18(Suppl 2):S305–12.
- 366. Han S, et al. Abstract B70: combination of AT1R blockade with CD40 activation provides enhanced therapeutic efficacy for mouse pancreatic adenocarcinoma. Cancer Immunol Res. 2015;3(10Supplement):B70–0.
- 367. Liu H, et al. Use of angiotensin system inhibitors is Associated with Immune activation and longer survival in Nonmetastatic Pancreatic Ductal Adenocarcinoma. Clin Cancer Res. 2017;23(19):5959–69.
- Nakai Y, et al. Inhibition of renin–angiotensin system affects prognosis of advanced pancreatic cancer receiving gemcitabine. Br J Cancer. 2010;103(11):1644–8.
- Liu H, et al. The use of angiotensin system inhibitors correlates with longer survival in resected pancreatic adenocarcinoma patients. HPB. 2023;25(3):320–9.
- 370. Keith SW, et al. Angiotensin blockade therapy and survival in pancreatic cancer: a population study. BMC Cancer. 2022;22(1):150.
- Tamaki Y, et al. Angiotensin II type 1 receptor antagonist prevents hepatic carcinoma in rats with nonalcoholic steatohepatitis. J Gastroenterol. 2013;48(4):491–503.
- 372. Saber S, et al. Perindopril, fosinopril and losartan inhibited the progression of diethylnitrosamine-induced hepatocellular carcinoma in mice via the inactivation of nuclear transcription factor kappa-B. Toxicol Lett. 2018;295:32–40.
- 373. Ahmadian E, et al. Novel angiotensin receptor blocker, azilsartan induces oxidative stress and NFkB-mediated apoptosis in hepatocellular carcinoma cell line HepG2. Biomed Pharmacother. 2018;99:939–46.
- 374. Gu L et al. Angiotensin II receptor inhibition ameliorates liver fibrosis and enhances hepatocellular carcinoma infiltration by effector T cells. bioRxiv, 2023.
- Sjöberg T, García Rodríguez LA, Lindblad M. Angiotensin-converting enzyme inhibitors and risk of esophageal and gastric cancer: a nested case-control study. Clin Gastroenterol Hepatol. 2007;5(10):1160–1166e1.

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