

PRIMARY RESEARCH

Open Access

Wnt5a promotes migration of human osteosarcoma cells by triggering a phosphatidylinositol-3 kinase/Akt signals

Ailiang Zhang^{*}, Shuanghua He, Xiaoliang Sun, Lianghua Ding, Xinnan Bao and Neng Wang

Abstract

Wnt5a is classified as a non-transforming Wnt family member and plays complicated roles in oncogenesis and cancer metastasis. However, Wnt5a signaling in osteosarcoma progression remains poorly defined. In this study, we found that Wnt5a stimulated the migration of human osteosarcoma cells (MG-63), with the maximal effect at 100 ng/ml, via enhancing phosphorylation of phosphatidylinositol-3 kinase (PI3K)/Akt. PI3K and Akt showed visible signs of basal phosphorylation and elevated phosphorylation at 15 min after stimulation with Wnt5a. Pharmaceutical inhibition of PI3K with LY294002 significantly blocked the Wnt5a-induced activation of Akt (p-Ser473) and decreased Wnt5a-induced cell migration. Akt siRNA remarkably inhibited Wnt5a-induced cell migration. Additionally, Wnt5a does not alter the total expression and phosphorylation of β -catenin in MG-63 cells. Taken together, we demonstrated for the first time that Wnt5a promoted osteosarcoma cell migration via the PI3K/Akt signaling pathway. These findings could provide a rationale for designing new therapy targeting osteosarcoma metastasis.

Keywords: Wnt5a, PI3K, Akt, Osteosarcoma, Migration

Introduction

Osteosarcoma, characterized by a high malignant and metastatic potential, principally affects children and adolescents [1]. Progression of disease is inexorable and response to therapy can be unrewarding: fewer than 50% of patients live beyond 10 years, and there are no reliable predictors to guide the choice or intensity of therapy [1,2]. Several improvements in understanding the molecular pathology of metastatic osteosarcoma have been achieved in the last several years [1]. However, the molecular mechanisms underlying this malignancy are still largely unknown. For this reason, elucidating the signaling pathways involved in the metastatic cascade has become a key goal for developing novel effective therapeutics aimed at reducing osteosarcoma mortality rates.

Wnt signaling regulates several developmental and oncogenic processes in both insects and vertebrates [3]. Signals triggered by Wnt are classified into two groups, a canonical β -catenin pathway and non-canonical pathways. In the canonical pathway, Wnt signals are mediated by

disheveled (Dvl), which inhibits glycogen synthase kinase-3 β (GSK-3 β) activity [4]. The accumulated β -catenin enters the nucleus, forms complexes as a co-factor with Tcf/Lef transcription factors, and then triggers transcription of a set of target genes, which ultimately leads to regulation of cell proliferation and cell fate as well as cell transformation [5]. Compared to the canonical pathway, the non-canonical Wnt pathways are not fully understood, especially in mammals, while its contributions to cell polarity in *Drosophila* (a planar cell polarity pathway, a PCP pathway) and convergent extension movement in *Xenopus* and zebrafish are demonstrated [6-9]. Wnt5a has been originally classified into the non-canonical Wnt group and reported to trigger Ca²⁺ pathways that subsequently activate protein kinase C (PKC) and Ca²⁺/Calmodulin kinase II [10,11]. Wnt5a can activate other intracellular protein kinases such as extracellular signal-regulated kinase (ERK), Akt and c-Jun N-terminal kinase (JNK) [12,13].

It has been reported that trimeric G protein mediated activation of phospholipase C and phosphodiesterase is involved in Wnt-induced gene expression [14]. Phosphatidylinositol-3 kinases (PI3Ks) are a family of enzymes involved in cellular functions including cell growth,

^{*} Correspondence: Ailiangzhang07@163.com
Department of Orthopaedics, Changzhou No. 1 People's Hospital,
Changzhou, Jiangsu 213003, PR China

proliferation, differentiation, motility, survival and intracellular trafficking, all of which can potentially influence the development of cancer. The serine/threonine kinase Akt is a major effector of the PI3K pathway and is activated by many polypeptide growth factors [15]. In this study, we hypothesized that the PI3K/Akt signaling pathway might mediate Wnt5a-induced osteosarcoma cell migration.

Materials and methods

Cell culture

Human osteosarcoma cell line MG-63 purchased from Cells Resource Center of Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences (Shanghai, China) and were cultured in Dulbecco-modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS).

Small interfering RNA (siRNA)

For gene knockdown, siRNA duplexes specific for Akt (Cell Signaling) were transfected into MG-63 cells by using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA) in serum-free OPTI-MEM according to the manufacturer's instructions. Knockdown efficiency was evaluated 48 h after transfection by measuring protein levels in cell lysates through using immunoblotting.

Immunoblotting analysis

Subconfluent cells were washed twice with PBS, and then lysed with ice-cold RIPA lysis buffer (50 mmol/L Tris, 150 mmol/L NaCl, 1% Triton X-100, 1% sodium deoxycholate, 0.1% SDS, 1 mmol/L sodium orthovanadate, 1 mmol/L sodium fluoride, 1 mmol/L EDTA, 1 mmol/L PMSE, and 1% cocktail of protease inhibitors) (pH7.4). The lysates were then clarified by centrifuging at 12,000 g for 20 min at 4°C. The protein extracts were separated by SDS-PAGE. The immunoblotting procedure was performed as described [16] and the following antibodies were used: mouse anti- β -actin antibody (KangChen Bio-tech, Shanghai, China), rabbit anti-Akt antibody, rabbit anti-phospho-Akt (p-Ser473) antibody, rabbit anti-PI3K p85 antibody, rabbit anti-phospho-PI3K p85 (Tyr458) (Cell Signaling Technology, Danvers, MA), mouse anti- β -catenin antibody, rabbit anti-phospho- β -catenin (p-Ser33) antibody (Santa Cruz Biotechnology, Santa Cruz, CA). Protein bands were detected by incubating with horseradish peroxidase-conjugated antibodies (Santa Cruz Biotechnology, Santa Cruz, CA) and visualized with ECL reagent (Thermo Scientific, Rockford, IL).

Wound healing assay

MG-63 cells were plated onto 96-well cell culture clusters (Costar) and grown to confluence, and then serum-starved for 24 h. The monolayer cells were scratched

manually with a plastic pipette tip, and after two washes with PBS, the wounded cellular monolayer was allowed to heal for 10 h in DMEM containing 100 ng/ml recombinant Wnt5a (rWnt5a) (R&D Systems). Photographs of central wound edges per condition were taken at time 0 and at the indicated time points using digital camera (Nikon, Tokyo, Japan).

Cell migration assays

Cell migration was assessed in a modified Boyden chamber (Costar), in which two chambers were separated by a polycarbonate membrane (pore diameter, 8.0 μ m). MG-63 cells were grown to subconfluence in tissue culture plates and then detached; thereafter, they were centrifuged and rendered into single cell suspensions in serum-free culture medium supplemented with 5 μ g/mL BSA. The suspensions containing 5×10^4 cells were added to wells with a membrane placed in the bottom. Medium containing indicated Wnt5a was added to the upper and lower compartment of the Boyden chamber. The cells were allowed to migrate for the indicated periods of time at 37°C in this assay. Thereafter, the medium was discarded, stationary cells were removed with a cotton-tipped applicator and the membranes were cut out of the chamber and stained with 0.5% crystal violet. The response was evaluated in a light microscope by counting the number of cells that had migrated into the membrane.

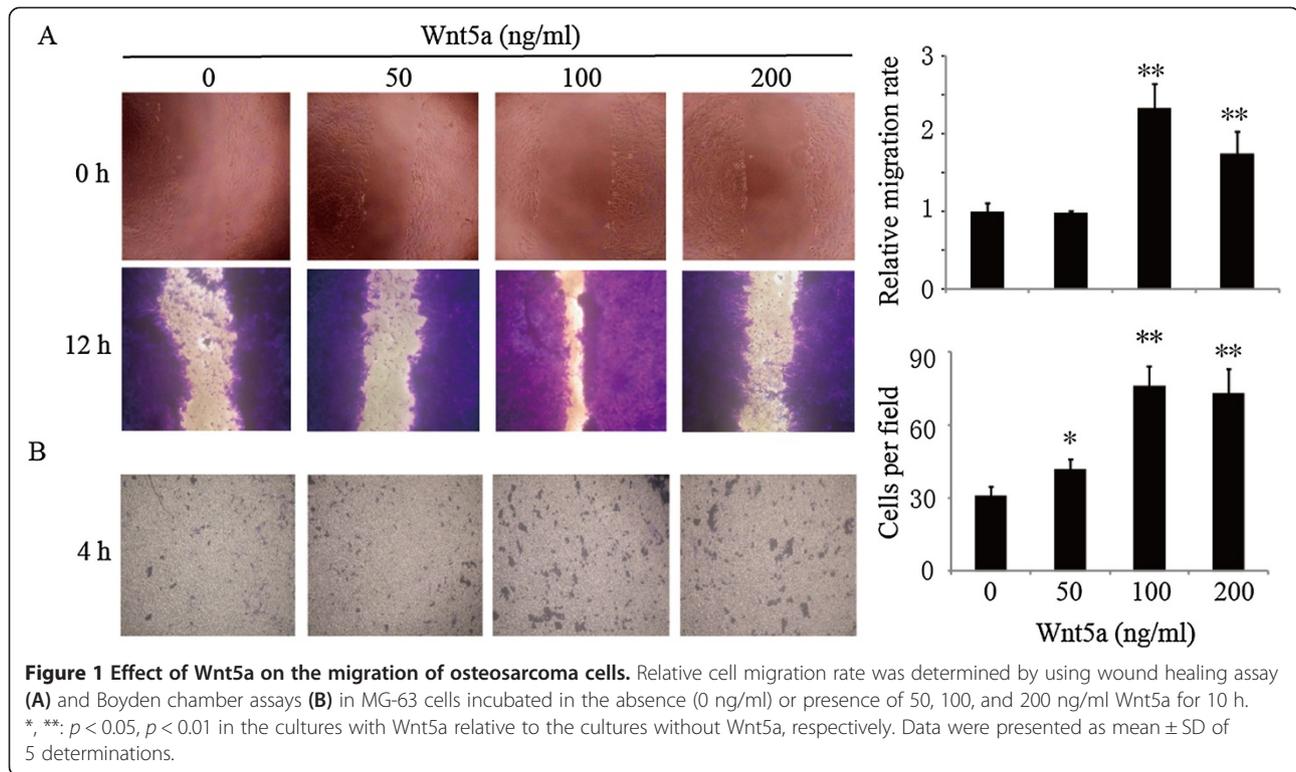
Statistical analysis

All experiments here were repeated at least three times, with independent treatments, each of which showed essentially the same results. The data were analyzed using Student's *t*-test by SPSS statistical software package. All the results were expressed as mean \pm SD. For all analyses a two-sided *p* value of less than 0.05 was deemed statistically significant. This study had been approved by institutional ethics committee of Changzhou No. 1 People's Hospital.

Results

Wnt5a stimulates osteosarcoma cell migration in vitro

To assess the effect of Wnt5a on osteosarcoma cell migration, we treated MG-63 cells with different doses of recombinant Wnt5a (rWnt5a), and measured the migration rate by wound healing assays and Boyden chamber assays. We found that Wnt5a had a potent stimulatory effect on MG-63 cell migration (Figure 1A, 1B). An approximately 2-fold increase in cell migration was observed in cells treated with 100 ng/ml rWnt5a (Figure 1A, 1B). Nevertheless, low concentration of Wnt5a (50 ng/ml) has no stimulative effect on MG-63 cell migration (Figure 1A). Accordingly, 100 ng/ml rWnt5a was used for the remaining studies hereafter to

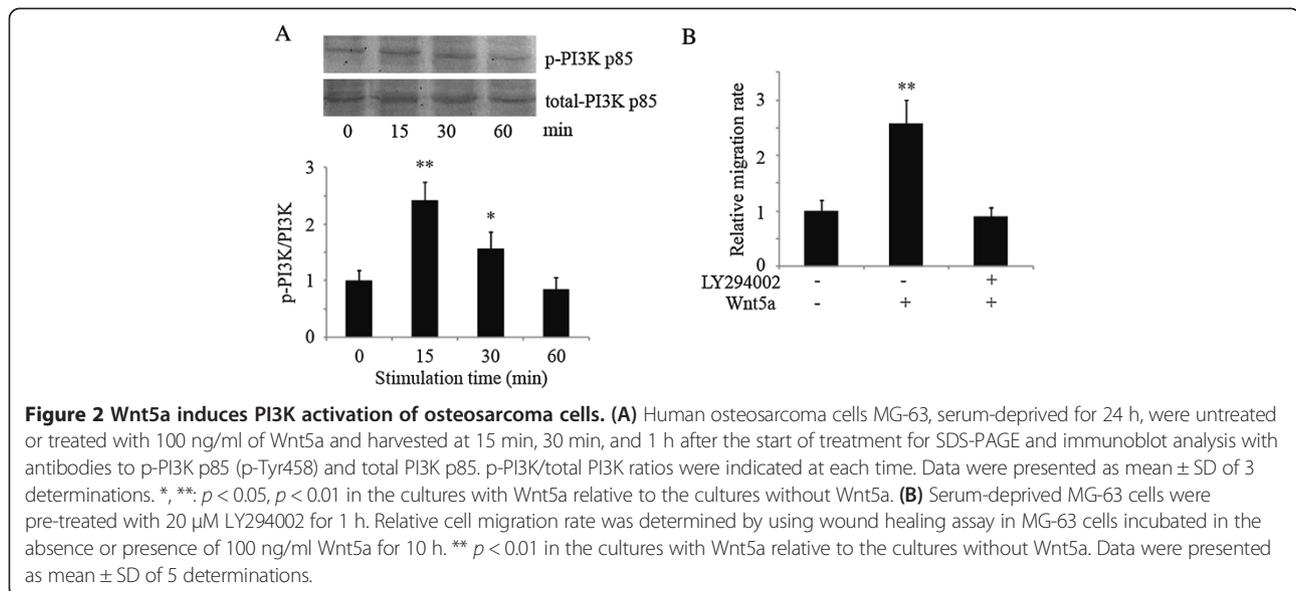


identify the mechanism that accounts for the changes in the migration of MG-63 cells.

Wnt5a induces PI3K and Akt phosphorylations

Wnt5a-triggered signals in human osteosarcoma cells have remained completely unknown. To address the question, we first tried to identify the downstream signals triggered by Wnt5a in MG-63 cells. We first detected the

phosphorylated-PI3K p85 (p-Tyr458), which represents the PI3K activation state. Human osteosarcoma cells, serum-starved for 24 h, were treated with 100 ng/ml of rWnt5a. The cells were harvested at 15 min, 30 min and 1 h after the start of Wnt5a treatment, followed by SDS-PAGE and immunoblot analyses. PI3K showed visible signs of basal phosphorylation and elevated phosphorylation at 15 min after stimulation with rWnt5a and



continued to be elevated at least until 30 min after the start of treatment with Wnt5a (Figure 2A).

The most established activator of Akt is PI3K, therefore we sought to determine whether Akt activation was triggered by Wnt5a. The same assays were performed to detect the phosphorylated-Akt (p-Ser473), which represents the Akt activation state. Akt also showed visible signs of basal phosphorylation and elevated phosphorylation at 15 min after stimulation with rWnt5a and continued to be elevated at least until 1 h after the start of treatment with Wnt5a (Figure 3A).

Wnt5a promotes cell migration via PI3K pathway

The finding that Wnt5a could induce PI3K/Akt phosphorylation in MG-63 cells prompted us to determine whether PI3K/Akt activation was required for Wnt5a-mediated cell migration. Wnt5a-induced cell migration was largely abolished by pre-treatment with 20 μ M LY294002, the PI3K-specific inhibitor (Figure 2B), suggesting that PI3K activation is required for Wnt5a-induced MG-63 cell migration.

To demonstrate the involvement of PI3K in Wnt5a-induced activation of Akt, we tested the effect of LY294002 for Akt activation. Human osteosarcoma cells, serum-starved for 24 h and pretreated with 20 μ M LY294002 for 1 h, were incubated with 100 ng/ml of Wnt5a. The cells were harvested 15 min after the start of Wnt5a treatment and the cell lysates were subjected to SDS-PAGE and immunoblot analysis. The Wnt5a-induced activation of Akt was mostly blocked by pretreatment of LY294002 (Figure 3B). These data indicate that PI3K mediates Wnt5a-induced activation of Akt.

Akt activation regulates osteosarcoma cell migration

To analyze the role of endogenous Akt activation on Wnt5a-induced cell migration, we knocked down Akt expression by using siRNA, which reduced the protein level of Akt by approximately 60%, as assessed by immunoblotting (Figure 4A) and significantly reduced Wnt5a-induced migration of MG-63 cells (Figure 4B). Taken together, these experiments demonstrated that Akt activation was required for Wnt5a-induced MG-63 cell migration.

Wnt5a does not alter the total expression and phosphorylation of β -catenin

Mikels and Nusse reported that purified Wnt5a inhibits Wnt3a protein-induced canonical Wnt signaling in a dose-dependent manner, not by influencing β -catenin levels but by downregulating β -catenin-induced reporter gene expression in HEK293 cells or mouse L cells [17]. To assess the effect of Wnt5a on the total expression and phosphorylation of β -catenin, we treated MG-63 cells with different doses of rWnt5a, and measured the total expression and phosphorylation by immunoblotting assays. We found that the total expression and phosphorylation of β -catenin had not been altered under Wnt5a stimulation for less than 1 hour (Figure 5).

Discussion

Wnt5a is a prototypic ligand that activates a β -catenin independent pathway in Wnt signaling [3]. Mice with the disrupted Wnt5a gene also exhibit various developmental abnormalities such as dwarfism, facial abnormalities, shortened limbs and tails, dysmorphic ribs and

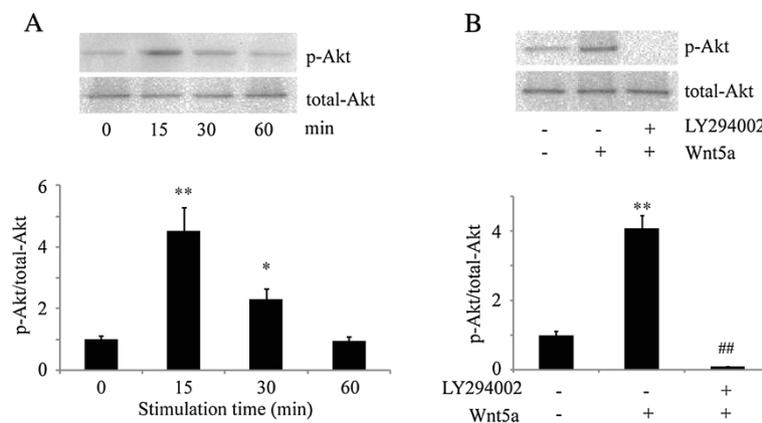
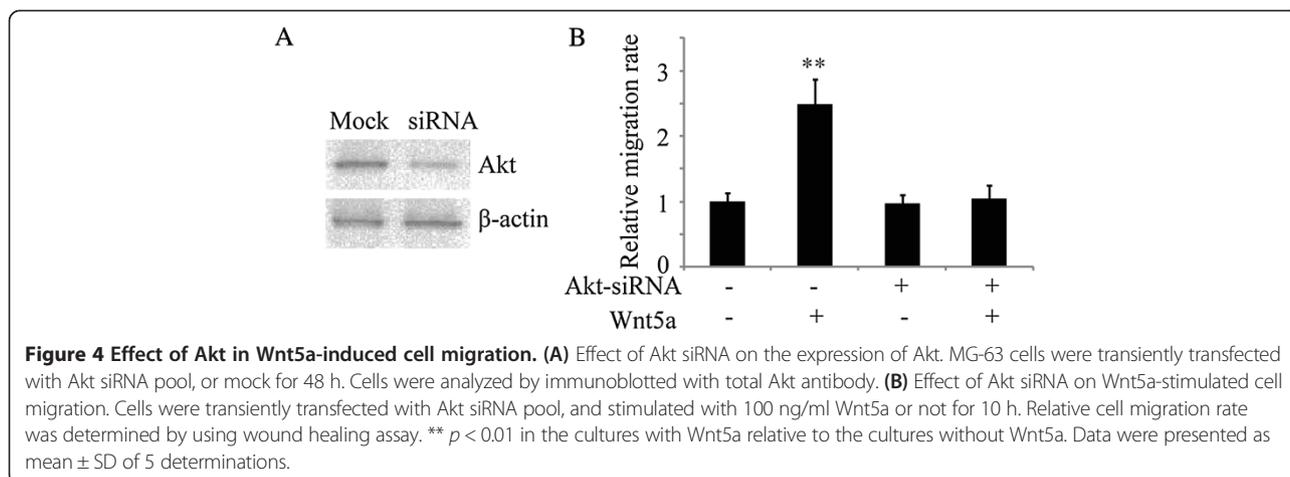


Figure 3 Wnt5a promotes cell migration via Akt pathway. (A) Human osteosarcoma cells MG-63, serum-deprived for 24 h, were untreated or treated with 100 ng/ml of Wnt5a and harvested at 15 min, 30 min, and 1 h after the start of treatment for SDS-PAGE and immunoblot analysis with antibodies to p-Akt (p-Ser473) and total Akt. p-Akt/total Akt ratios were indicated at each time. Data were presented as mean \pm SD of 3 determinations. *, **: $p < 0.05$, $p < 0.01$ in the cultures with Wnt5a relative to the cultures without Wnt5a. (B) An equal number of MG-63 cells were pre-treated with 20 μ M LY294002 (PI3K inhibitor) for 1 h, then incubated with 100 ng/ml Wnt5a for 15 min. After stimulation, cells were analyzed by immunoblotted with p-Akt (p-Ser473) or total Akt antibodies. Data were presented as mean \pm SD of 3 determinations. **, ##: $p < 0.01$, $p < 0.01$, in the cultures with Wnt5a relative to the cultures without Wnt5a.

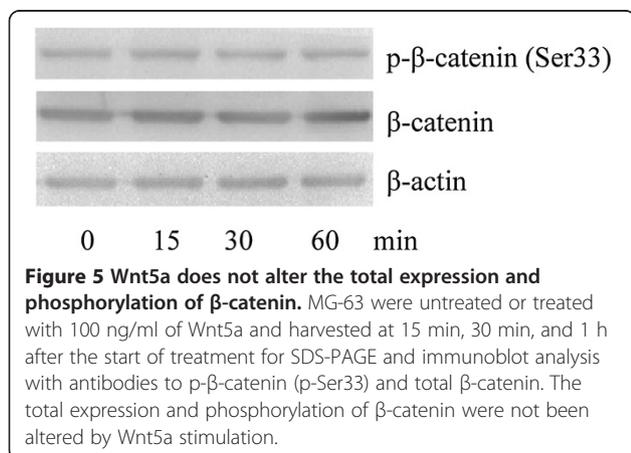


vertebrae, absence of the genital tubercle, and abnormal distal lung morphogenesis, indicating that it plays a crucial role in the development of various organs [18]. Wnt5a has been demonstrated to exert differential effects on cancer development [5]. Wnt5a can promote cancer progression and metastasis in malignant melanoma, breast cancer, and gastric cancer [19-23]. However, some evidence supports the hypothesis that Wnt5a acts as a tumor suppressor in certain experimental systems [24-28]. The first observation in the present study was that Wnt5a induced the migration of MG-63 osteosarcoma cells. Our finding suggest that Wnt5a acts as a migratory stimulator in osteosarcoma cells.

Kinases show the comprehensive functions triggered by Wnt5a in a variety of cells. It has been reported that Wnt5a may promote tumor progression through inducing actin reorganization and increasing cell motility via activating the protein kinase C (PKC) [19,29]. Wnt5a activates extracellular regulated protein kinase (ERK) in endothelial cells [30]. Wnt5a prevents apoptosis caused by serum-deprivation in osteoblasts by activating both

Akt and ERK [12], and promotes cell adhesion in mammary epithelial cells in a PI3K/Akt-dependent manner [31]. Furthermore, Wnt5a triggers Akt phosphorylation via PI3K, but not ERK or PKC phosphorylation in human dermal fibroblasts [14]. These tanglesome viewpoints of kinases in Wnt5a signaling indicate that phosphorylated signaling molecules triggered by Wnt5a in these cases determine the diverse roles of these kinases. In this study, we demonstrated that Wnt5a promoted MG-63 osteosarcoma cell migration by activating PI3K/Akt. We found that Wnt5a induced PI3K (p-Tyr458) and Akt phosphorylation (p-Ser473) rapidly and transiently in MG-63 cells. Blocking Akt signaling by chemical inhibition of PI3K or Akt siRNA completely abolished Wnt5a-induced cell migration, indicating that PI3K/Akt activation participated in the regulation of the MG-63 cell migration.

The activation of PI3K/Akt signaling is thought to play a central role in cell viability-promoting phenotypes such as inhibition of cell death and cell cycle progression [14]. It has been shown that the PI3K/Akt signaling could be activated not only by Wnt5a but also by Wnt3a. Wnt3a prevents the serum deprivation-triggered apoptosis of osteoblasts by activating the canonical β -catenin pathway, ERK-, and/or PI3K-mediated uncharacterized pathways [12]. In this study, we have showed that Wnt5a promotes cell migration of human osteosarcoma cells in the PI3K/Akt-dependent manner. These results indicated that PI3K/Akt played crucial and complicated roles in canonical and non-canonical Wnt signalings in various cell and tissue types. Besides mentions above, PI3K/Akt signaling presents wide crosstalks referred to instances in which one or more components of one signal transduction pathway affect another. High-mobility group box 1 (HMGB1) activates the FAK/PI3K/mTOR signaling cascade and regulates tumor-associated cell migration through the interaction with BTB domain [32]. CXCR4 drives the metastatic phenotype in breast cancer through



induction of CXCR2, and activation of MEK and PI3K pathways [33]. Akt signaling is involved in fucoidan-induced inhibition of growth and migration of human bladder cancer cells [34]. These studies provide us a complicated and crossed network around the PI3K/Akt signaling.

In summary, we present the first direct evidence here that Wnt5a promotes osteosarcoma cell migration via PI3K/Akt signaling. These findings elucidate a molecular pathway linking Wnt5a signaling to PI3K/Akt in cell motility. This result will contribute to further understanding of biological roles of Wnt5a/PI3K/Akt in cell migration of osteosarcoma and other cancers.

Competing interest

The author declares that they have no competing interest.

Authors' contributions

AZ and SH participated in the design of this study, and they both carried out the study. XS collected important background information, together with LD, and performed the statistical analysis. XB and NW participated in the design and helped to write the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by grant from the Science & Technology Bureau of Changzhou (No.CJ20130029).

Received: 10 July 2013 Accepted: 7 February 2014

Published: 14 February 2014

References

1. Marulanda GA, Henderson ER, Johnson DA, Letson GD, Cheong D: Orthopedic surgery options for the treatment of primary osteosarcoma. *Cancer Control* 2008, **15**(1):13–20.
2. Mirabello L, Troisi RJ, Savage SA: Osteosarcoma incidence and survival rates from 1973 to 2004: data from the surveillance, epidemiology, and end results program. *Cancer* 2009, **115**(7):1531–1543.
3. Logan CY, Nusse R: The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol* 2004, **20**:781–810.
4. Widelitz R: Wnt signaling through canonical and non-canonical pathways: recent progress. *Growth Factors* 2005, **23**(2):111–116.
5. Klaus A, Birchmeier W: Wnt signalling and its impact on development and cancer. *Nat Rev Cancer* 2008, **8**(5):387–398.
6. Moon RT, Campbell RM, Christian JL, McGrew LL, Shih J, Fraser S: Xwnt-5A: a maternal Wnt that affects morphogenetic movements after overexpression in embryos of *Xenopus laevis*. *Development* 1993, **119**(1):97–111.
7. Wallingford JB, Rowing BA, Vogeli KM, Rothbacher U, Fraser SE, Harland RM: Dishevelled controls cell polarity during *Xenopus* gastrulation. *Nature* 2000, **405**(6782):81–85.
8. Heisenberg CP, Tada M, Rauch GJ, Saude L, Concha ML, Geisler R, Stemple DL, Smith JC, Wilson SW: Silberblick/Wnt11 mediates convergent extension movements during zebrafish gastrulation. *Nature* 2000, **405**(6782):76–81.
9. Tada M, Smith JC: Xwnt11 is a target of *Xenopus* Brachyury: regulation of gastrulation movements via Dishevelled, but not through the canonical Wnt pathway. *Development* 2000, **127**(10):2227–2238.
10. Veeman MT, Axelrod JD, Moon RT: A second canon. Functions and mechanisms of beta-catenin-independent Wnt signaling. *Dev Cell* 2003, **5**(3):367–377.
11. Kuhl M, Sheldahl LC, Park M, Miller JR, Moon RT: The Wnt/Ca²⁺ pathway: a new vertebrate Wnt signaling pathway takes shape. *Trends Genet* 2000, **16**(7):279–283.
12. Almeida M, Han L, Bellido T, Manolagas SC, Kousteni S: Wnt proteins prevent apoptosis of both uncommitted osteoblast progenitors and differentiated osteoblasts by beta-catenin-dependent and -independent signaling cascades involving Src/ERK and phosphatidylinositol 3-kinase/AKT. *J Biol Chem* 2005, **280**(50):41342–41351.
13. Yamanaka H, Moriguchi T, Masuyama N, Kusakabe M, Hanafusa H, Takada R, Takada S, Nishida E: JNK functions in the non-canonical Wnt pathway to regulate convergent extension movements in vertebrates. *EMBO Rep* 2002, **3**(1):69–75.
14. Kawasaki A, Torii K, Yamashita Y, Nishizawa K, Kanekura K, Katada M, Ito M, Nishimoto I, Terashita K, Aiso S, et al: Wnt5a promotes adhesion of human dermal fibroblasts by triggering a phosphatidylinositol-3 kinase/Akt signal. *Cell Signal* 2007, **19**(12):2498–2506.
15. Datta SR, Brunet A, Greenberg ME: Cellular survival: a play in three Akts. *Genes Dev* 1999, **13**(22):2905–2927.
16. Zhang A, Zhang J, Sun P, Yao C, Su C, Sui T, Huang H, Cao X, Ge Y: EIF2alpha and caspase-12 activation are involved in oxygen-glucose-serum deprivation/restoration-induced apoptosis of spinal cord astrocytes. *Neurosci Lett* 2010, **478**(1):32–36.
17. Mikels AJ, Nusse R: Purified Wnt5a protein activates or inhibits beta-catenin-TCF signaling depending on receptor context. *PLoS Biol* 2006, **4**(4):e115.
18. Yamaguchi TP, Bradley A, McMahon AP, Jones S: A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo. *Development* 1999, **126**(6):1211–1223.
19. Weeraratna AT, Jiang Y, Hostetter G, Rosenblatt K, Duray P, Bittner M, Trent JM: Wnt5a signaling directly affects cell motility and invasion of metastatic melanoma. *Cancer Cell* 2002, **1**(3):279–288.
20. Zhu Y, Tian Y, Du J, Hu Z, Yang L, Liu J, Gu L: Dvl2-dependent activation of Daam1 and RhoA regulates Wnt5a-induced breast cancer cell migration. *PLoS One* 2012, **7**(5):e37823.
21. Zhu Y, Shen T, Liu J, Zheng J, Zhang Y, Xu R, Sun C, Du J, Chen Y, Gu L: Rab35 is required for Wnt5a/Dvl2-induced Rac1 activation and cell migration in MCF-7 breast cancer cells. *Cell Signal* 2013, **25**(5):1075–1085.
22. Kurayoshi M, Oue N, Yamamoto H, Kishida M, Inoue A, Asahara T, Yasui W, Kikuchi A: Expression of Wnt-5a is correlated with aggressiveness of gastric cancer by stimulating cell migration and invasion. *Cancer Res* 2006, **66**(21):10439–10448.
23. Bittner M, Meltzer P, Chen Y, Jiang Y, Seftor E, Hendrix M, Radmacher M, Simon R, Yakhini Z, Ben-Dor A, et al: Molecular classification of cutaneous malignant melanoma by gene expression profiling. *Nature* 2000, **406**(6795):536–540.
24. Kremenevskaja N, von Wasielewski R, Rao AS, Schoff C, Andersson T, Brabant G: Wnt-5a has tumor suppressor activity in thyroid carcinoma. *Oncogene* 2005, **24**(13):2144–2154.
25. Dejmeek J, Dejmeek A, Safholm A, Sjolander A, Andersson T: Wnt-5a protein expression in primary duodenal B colon cancers identifies a subgroup of patients with good prognosis. *Cancer Res* 2005, **65**(20):9142–9146.
26. Liu XH, Pan MH, Lu ZF, Wu B, Rao Q, Zhou ZY, Zhou XJ: Expression of Wnt-5a and its clinicopathological significance in hepatocellular carcinoma. *Dig Liver Dis* 2008, **40**(7):560–567.
27. Safholm A, Tuomela J, Rosenkvist J, Dejmeek J, Harkonen P, Andersson T: The Wnt-5a-derived hexapeptide Foxy-5 inhibits breast cancer metastasis in vivo by targeting cell motility. *Clin Cancer Res* 2008, **14**(20):6556–6563.
28. Jonsson M, Dejmeek J, Bendahl PO, Andersson T: Loss of Wnt-5a protein is associated with early relapse in invasive ductal breast carcinomas. *Cancer Res* 2002, **62**(2):409–416.
29. Witze ES, Litman ES, Argast GM, Moon RT, Ahn NG: Wnt5a control of cell polarity and directional movement by polarized redistribution of adhesion receptors. *Science* 2008, **320**(5874):365–369.
30. Mascakauchan TN, Agalliu D, Vorontchikhina M, Ahn A, Parmalee NL, Li CM, Khoo A, Tycko B, Brown AM, Kitajewski J: Wnt5a signaling induces proliferation and survival of endothelial cells in vitro and expression of MMP-1 and Tie-2. *Mol Biol Cell* 2006, **17**(12):5163–5172.
31. Dejmeek J, Dib K, Jonsson M, Andersson T: Wnt-5a and G-protein signaling are required for collagen-induced DDR1 receptor activation and normal mammary cell adhesion. *Int J Canc Suppl J Int Canc Suppl* 2003, **103**(3):344–351.
32. Ko YB, Kim BR, Nam SL, Yang JB, Park SY, Rho SB: High-mobility group box 1 (HMGB1) protein regulates tumor-associated cell migration through the interaction with BTB domain. *Cell Signal* 2014, **26**(4):777–783.

33. Sobolik T, Su YJ, Wells S, Ayers GD, Cook RS, Richmond A: **CXCR4 drives the metastatic phenotype in breast cancer through induction of CXCR2, and activation of MEK and PI3K pathways.** *Mol Biol Cell* 2014:mbc.E13-07-0360.
34. Cho TM, Kim WJ, Moon SK: **AKT signaling is involved in fucoidan-induced inhibition of growth and migration of human bladder cancer cells.** *Food Chem Toxicol* 2014, **64**:344–352.

doi:10.1186/1475-2867-14-15

Cite this article as: Zhang *et al.*: Wnt5a promotes migration of human osteosarcoma cells by triggering a phosphatidylinositol-3 kinase/Akt signals. *Cancer Cell International* 2014 **14**:15.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

