

REVIEW

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New insights into M1/M2 macrophages: key modulators in cancer progression

Jiuyang Liu^{1†}, Xiafei Geng^{2†}, Jinxuan Hou^{1*} and Gaosong Wu^{1*} 

Abstract

Infiltration of macrophages in and around tumor nest represents one of the most crucial hallmarks during tumor progression. The mutual interactions with tumor cells and stromal microenvironment contribute to phenotypically polarization of tumor associated macrophages. Macrophages consist of at least two subgroups, M1 and M2. M1 phenotype macrophages are tumor-resistant due to intrinsic phagocytosis and enhanced antitumor inflammatory reactions. Contrastingly, M2 are endowed with a repertoire of tumor-promoting capabilities involving immuno-suppression, angiogenesis and neovascularization, as well as stromal activation and remodeling. The functional signature of M2 incorporates location-related, mutually connected, and cascade-like reactions, thereby accelerating paces of tumor aggressiveness and metastasis. In this review, mechanisms underlying the distinct functional characterization of M1 and M2 macrophages are demonstrated to make sense of M1 and M2 as key regulators during cancer progression.

Keywords: Tumor-associated macrophages, Cancer, Metastasis, Polarization

Introduction

Tumor, to some extent, could be defined as a systemic disease of immuno-imbalance [1]. Malignant tumor is characterized by uncontrolled cell proliferation due to the unbalance between the mutations of oncogenes and tumor suppressor genes [2]. Once the mutation has been detected by host immune system, various lymphocytic infiltrates would accumulate in and around tumor zones to harbor intrinsic and adaptive immunities [3]. In this case, the tumoral immune microenvironment plays crucial roles during the multi-stage processes of tumorigenesis and progression.

Within the immune microenvironment, resident and recruited macrophages act as first lines of immunoregulatory functions and key modulators during tumor progression. In response to tumor-derived or

microenvironmental signals, macrophages undergo phenotypically polarization to a universe of activation states [4]. Extremely, the macrophages plasticity could be summarized as M1 (classically activated) and M2 (alternatively activated) phenotypes, for better understandings of their distinct cellular and molecular mechanisms [5]. Both of M1 and M2 macrophages are involved in the affection of tumor-related inflammatory [6], whereas M2 is prone to promote angiogenesis and neovascularization, as well as stromal activation and remodeling [7, 8], thereby impacting cancer progression positively and patient' prognosis negatively [9]. Consequently, the interplay between host immune system and tumor cells could be representatively indicated from the aspect of M1/M2 macrophages.

For long momentum studies have emphasized the significance of M1/M2 macrophages. Some questions remain to be debated. Whether phenotypical polarizations explain pleiotropic but opposed activities of TAM? To what extent M1 and M2 differ considering functional properties under the framework of tumor microenvironment? Do spatial locations of macrophages especially M2 contribute largely to tumor progression? After describing

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the origination and differentiation of macrophages, this review would focus on the distinct functional characteristics of M1/M2 in terms of three major aspects, including immunoediting, angiogenesis and neovascularization, and stromal orchestration.

Origination and differentiation of macrophages

Origination and maturation of macrophages

Macrophages populated within tissues are grouped resident macrophages and recruited macrophages. For long resident macrophages are proved to be replaced by migrated circulating peripheral blood monocytes, which acted as precursors of macrophages [5]. Recent studies have found a fact that resident macrophages are endowed with self-renewal capacity especially under inflammatory conditions. Meanwhile, these studies imply a new origination from hematopoiesis in the yolk sac before birth [10]. Recruited macrophages are mainly derived from monocytes in circulation and bone marrow derived cells (BMDC) [11]. Homeostatic control of monocyte/macrophage development is majorly driven by colony stimulating factor-1 (CSF-1, also known as M-CSF) [12]. New evidence has identified the cytokine IL-34 as a new CSF-1R ligand that influences maturation of macrophages, which is restricted to the epidermis and central nervous system [13].

Differentiation and phenotypes of macrophages

In normal tissues, macrophages could be differentiated into microglial cells in the brain, Kupffer cells in the liver, and Langerhans cells in the skin [14, 15]. In the similar manner, the phenotypes of macrophages within tumor tissues (tumor associated macrophages, TAM) also represent of innate and adaptive immune responses. Tumor related inflammation represents one of the complex hallmarks of cancer especially due to an enrichment with monocyte-derived macrophages [16]. Theoretically, in response to various stimuli, TAM could be differentiated into distinct functional subsets, extremely into M1 phenotype macrophages by Th1 (IFN- γ , TNF α , and LPS et al.), and into M2 by Th2 (IL-4, IL-10, TGF β 1, and PGE2 et al.) cytokines and immunocomplexes [17] (Fig. 1). M1 phenotype macrophages are characterized by the expression of HLA-DR and CD197, whereas M2 is typical for the high expression of CD163, CD209, CD206, and CCL2 et al. [18, 19]. In addition, potential roles of IRF4 in M1-like polarization [20] and Trib1 in tissue-resident M2-like macrophages [21] have been demonstrated, which further confirms that macrophages consist of more than two subgroups. Underlying mechanisms of TAM polarization has been demonstrated to be correlated with several major signaling pathways, including C-Jun N-terminal kinase (JNK) signaling pathway, PI3K/

Akt signaling pathway, Notch signaling pathway, JAK/STAT signaling pathway, et al. [22].

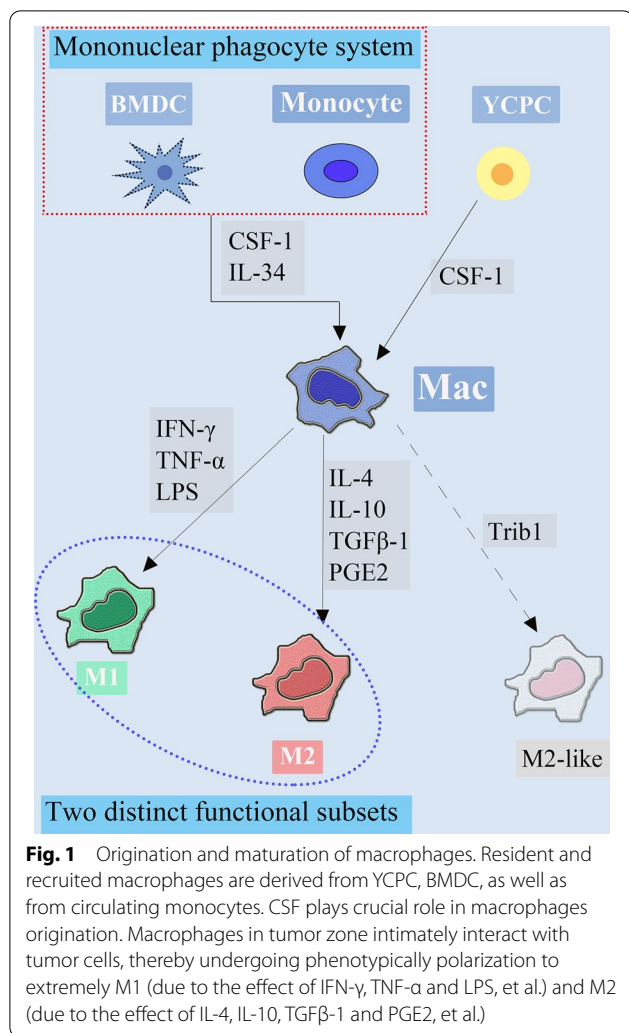
Pleiotropic but opposed activities of TAM

Herein, we would focus on the location-related distinct functional properties instead of these pathways. In normal tissues, macrophages contribute to maintaining tissue homeostasis through phases-depending mechanisms. In the initiation phase of host immune response, macrophages could generally be served as tissue sentinels via phagocytosis. Then the damaged cells and debris could be eliminated by macrophages in the phase of full inflammation. In the recovery phase, macrophages participate in the re-establishment of tissue integrity [23]. As known, tumor is partially a disease of homeostatic imbalance. Some mechanisms involved in keeping normal tissues homeostasis might also be utilized by TAM to acquire different functional phenotypes, thereafter influencing adaptive immunity towards different directions [24].

In this perspective, momentum TAM accumulated in and around tumor organs to monitor the abnormal tissue architecture, cellular compositions, and functional states [25], whereas TAM could finally be trained to perform key homeostatic functions in the maintenance of tumoral growth and progression [26]. Reasons could be offered by the issue of heterogeneity in the M1/M2. M1 phenotype macrophages are tumor-resistant due to intrinsic phagocytosis and enhanced antitumor inflammatory reactions. Contrastingly, M2 are endowed with a repertoire of tumor-promoting capabilities involving immunosuppression, angiogenesis and neovascularization, as well as stromal activation and remodeling [27]. Functional properties of M1 and M2 would be discussed later in this review. Due to ignorance of distinct polarization of TAM, researches on exact prognostic values of TAM during cancer progression still failed to reach a consensus and remained contradictory [28]. In addition, histological distribution of TAM within cancer tissue has been analyzed and studied [29, 30], further proven to be related with corresponding functional potentials and overall influence on cancer outcomes [31, 32].

Major functional properties of M1 phenotype

M1 phenotype macrophages have intrinsic function to trap, phagocytose, and lyse tumor cells [33]. In addition, enhanced tumor antigen presenting ability of M1 would promote other leukocytes cytotoxic functions. For instance, CD8+ T cells and NK cells could be strengthened by immuno-stimulatory cytokines (IL-6, IL-12, TNF et al.) from M1 phenotype macrophages [34]. As a result, tumor cells apoptosis would be induced at this stage. Tumor stem cells have fewer immunogenic antigens, but more vigorous proliferative and differentiative abilities.



Considering the heterogeneity and tumor-intrinsic mechanisms of immune escape [35], tumor stem cells might use M1 phenotype macrophages as a natural filter to avoid being destroyed and survived to next stage [36] (Fig. 2a).

Major functional properties of M2 phenotype *Immuno-modulation basing on immunoediting theory*

In contrast to M1, most macrophages are switched to M2 phenotype followed by the interactions with tumor cells, which is prone to show immunosuppression potentials. Contrary, M2 act as immune-suppressor in tumor nest. On one hand, the pro-inflammatory potential of M2 phenotype macrophages are dramatically descended, due to weakened tumor antigen presenting ability [37] and the secretions of suppressor factors such as IL-12 [38]. On the other hand, M2 phenotype macrophages are educated to be tumor-promoting by releasing growth factors like PDGF, TGF β 1, HGF, and bFGF et al. [4, 39], forming a

positive feed-back loop in accordance with cytokines and factors of tumor cells (IL-4, IL-6, IL-10, MDE, TGF β 1, PGE2, et al.) [40] (Fig. 2b). In fact, all macrophages are crucial for tumor proliferation at primary tumor nest, and intrigue subsequent tumor metastasis. Therefore, tumor progression could be hypothesized as stages of immune elimination, equilibrium and escape basing on M1/M2 ratios and reactions according to immunoediting theory [41].

Angiogenesis and neovascularization

For long it has been a consensus that angiogenesis and neovascularization are essential for tumor growth and metastasis [42]. Many studies have shown that M2 phenotype macrophages are crucial to support the evolution of tumor-related vasculature, a process in which neo-vessel sprout from existing blood vessels or by proliferation, motility and accumulation of vascular endothelial cells [43]. Recent studies have emphasized the significant correlation between M2 and tumor neo-vessels with focus on perivascular and peri-necrotic areas [44]. Additionally, morphological studies have demonstrated a disorganized and collapsed characteristic of tumor neo-vessels [45]. This reason, combined with over-rapid growth of tumor cells, coordinately contribute to hypoxia zones, especially in tumor center and peri-necrotic regions. Hypoxia in these areas induces the expression of inflammatory molecules (IL-4, IL-10 et al.), thus promoting the recruitment of macrophages followed by conversion to the M2 phenotype [46]. In this case, M2 participates in a “angiogenesis cascade”, which is capable to affect the onset and maintenance of the angiogenic process, including degradation of the extracellular matrix, endothelial cell proliferation and migration, and neovascularization [47]. Molecular mechanisms underlying each phase could resulted from a variety of pro-angiogenic cytokines and growth factors, including VEGF, TNF α , IL-8, and bFGF, as well as angiogenesis-regulating enzymes like MMP and COX-2 [48] (Fig. 2c).

Stromal orchestration

Emerging evidence has also illustrated the significance of tumor stromal transformation in tumor progression. Herein, M2 is discussed in terms of intimate correlations with other stromal components. Actually, stromal cells around incipient tumors have a futile attempt to “destroy and repair” tumor tissues [49]. However, M2 could assist tumor cells to gain a hallmark to activate and remodel stromal features to support tumors [50].

For instance, fibroblasts differentiated from mesenchymal cells are important to tissue-repair, whereas transformed to cancer-associated fibroblasts (CAFs). In turn, CAFs also promote tumor growth and metastasis

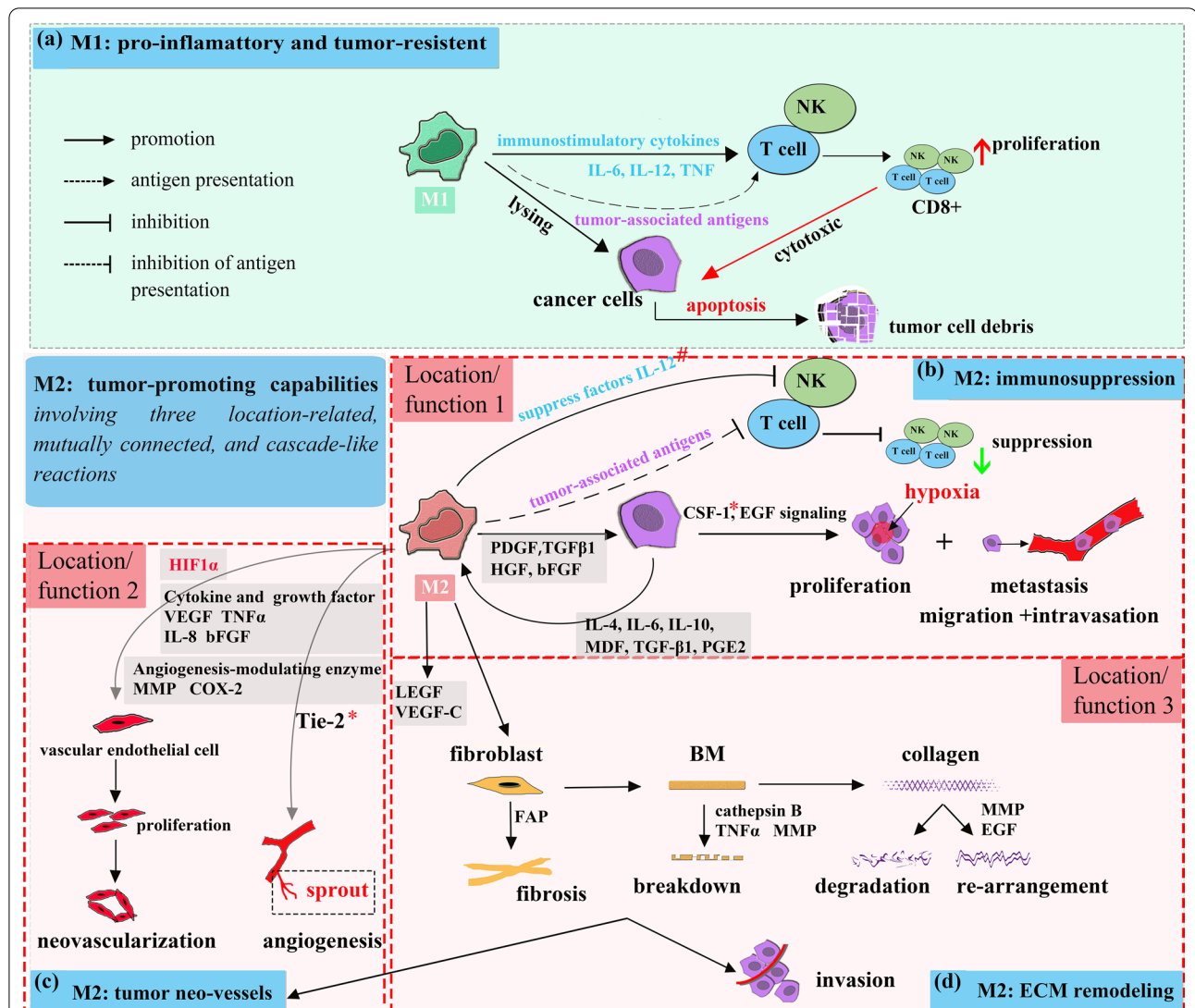


Fig. 2 Functional characterization of M1 and M2 macrophages. **a** Generally, M1 is tumor-resistant by directly lysing tumor cells after phagocytosis, and pro-inflammatory by enhanced tumor antigen-presenting ability or by indirectly promote the proliferation of immune cells like CD8+ T cells and NK cells (due to the effect of IL-6, IL-12 and TNF, et al.). **b-d** Whist and however, M2 is tumor-promoting through a repertoire of mechanisms, typically summarized as immunosuppression, tumor angiogenesis and neo-vascularization, and stromal activation and remodeling. **b** In contrast to M1-related immune response, M2 obstacles host immune states (Function 1). In tumor center (defined as Location 1) growth factors (including PDGF, TGFβ, HGF, and bFGF et al.) secreted by M2 would induce proliferation and metastasis of tumor cells. As a feed-back loop, cytokines and factors (including IL-4, IL-6, IL-10, MDF, TGF-β1 and PGE2 et al.) secreted by tumor cells enhance this effect in turn. **c** In avascular and peri-necrotic areas (defined as Location 2), HIF1α induced by hypoxia or low oxygen tension, in accordance with cytokines and factors (including VEGF, TNFα, IL-8 and bFGF et al.), and angiogenesis-modulating enzymes (including MMP and COX-2 et al.) would promote neo-vascularization and induce angiogenesis (Function 2). **d** In stromal areas (defined as Location 3), M2 actively impact on CAFs differentiation, BM breakdown, and collagen degradation and re-arrangement (Function 3). These combined stromal remodeling signatures would correspondingly induce tumor neo-vessels formation and maturation, as well as tumor invasion capability. The distinct tumor-related potentials of M1/M2 should be further investigated from known mechanisms illustrated in this figure

by recruiting and reprogramming macrophages into M2 phenotype [51]. In addition, M2-related enzymes promote ECM digestion and deposition, as well as the proteolysis of basement membrane (BM) and collagens surrounding tumor nests [52]. In this case, intrinsic

defensive abilities of collagens are dramatically reduced. The principal factors accounted for this transformation include cathepsin B, TNFα, MMPs et al. [53]. Subsequently, degraded BM and collagen fragments may serve as chemotactic stimuli to circulating

monocytes. These recruited macrophages, especially M2, could further induce the degradation and re-arrangement, enhance the stiffness of collagens, and co-orchestrate angiogenesis [54]. Thus, all these have uncovered a positive feed-back transformation displayed among M2 and other representative stromal constitutions (Fig. 2d).

New insights into distribution pattern of macrophages

The specialization of macrophages in particular micro-environments explains their heterogeneity. In addition, the heterogeneous functional properties of macrophages would result from their location in tumor tissues [55]. There are three typical locations including tumor center, invasive front (the interface between tumor cells and stroma), and tumor stroma [56]. It has been documented that macrophages seem to be “hijacked” to perform distinct functions according to the location-related signals [22]. Briefly, M2 phenotype macrophages would be preferentially endowed with potentials to promote the motility of cancer cells in invasion areas [57], to promote metastasis in stromal and perivascular areas [31], and to stimulate angiogenesis in avascular and peri-necrotic hypoxic areas [54]. In other words, the distribution pattern of macrophages might be correlated with different cancer progression mechanisms.

Notably, we have recently found a timely novel result that the distribution pattern of macrophages could be an independent prognostic factor in gastric cancer [30]. According to relative macrophages densities in tumor nest or tumor stroma, gastric cases could be divided into nest-dominant pattern and stroma-dominant pattern. As a phenomenon, stroma-dominant pattern cases tended to have poorer survival and higher malignancy. In theory, accumulation of macrophages in tumor stroma might participate more actively in the process of stroma activation [58] and ECM remodeling, together with other stromal components including lysyl oxidase, MMP9, type IV collagen, which has been emphasized by Peng et al. [59]. In addition, Yang et al. [60] preliminarily elucidated the prognostic value of CD163+/CD68+ ratio in colorectal cancer invasive front. To make sense of macrophages polarization in different distribution patterns, studies could be conducted around macrophages phenotypes and ratios, tumor locations and biological functions in the future.

Conclusions

In summary, M1 and M2 macrophages are functionally distinct and key modulators in host immune system against tumors. In contrast to M1, M2 mechanisms incorporate location-related, mutually connected, and cascade-like reactions. A better understanding of macrophage polarization, especially in distinct locations

could make sense for tumor progression and guide therapy in the future.

Abbreviations

BM: Basement membrane; BMDC: Bone marrow derived cell; bFGF: Basic fibroblast growth factor; CAFs: Cancer-associated fibroblasts; COX: Cyclooxygenase; CSF: Colony stimulation factor; ECM: Extracellular matrix; EGF: Epidermal growth factor; IFN- γ : Interferon- γ ; IL: Interleukin; LEGF: Lymphatic endothelial growth factor; LPS: Lipopolysaccharide; Mac: Macrophages; MDF: Myocardial depressant factor; MMP: Matrix metallo proteinases; NK: Natural killer cell; PGE2: Prostaglandin E2; TAM: Tumor associated macrophages; TNF- α : Tumor necrosis factor- α ; VEGF: Vascular endothelial growth factor; YCPC: Yolk sac progenitor cell.

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Authors' contributions

GW and JL participated in the design of the review. JL and XG supervised the study and wrote the manuscript. HJ improved language editing and illustrations. All authors read and approved the final manuscript.

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Availability of data and materials

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Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Written informed consent for publication was obtained from all authors.

Competing interests

The authors declare no conflicts of interest.

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References

- Shi X, Tang Y, Sun X, et al. Interleukin33 induced immune tolerance is associated with the imbalance of memory and naïve T lymphocyte subsets. *Mol Med Rep*. 2016;14:4837–43.
- Morris LG, Chan TA. Therapeutic targeting of tumor suppressor genes. *Cancer*. 2015;121:1357–68.
- Burugu S, Asleh-Aburaya K, Nielsen TO. Immune infiltrates in the breast cancer microenvironment: detection, characterization and clinical implication. *Breast Cancer*. 2017;24:3–15.
- Murray PJ, Allen JE, Biswas SK, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity*. 2014;41:14–20.

5. Italiani P, Boraschi D. From monocytes to M1/M2 macrophages: phenotypical vs. functional differentiation. *Front Immunol*. 2014;5:514.
6. Ries CH, Cannarile MA, Hoves S, et al. Targeting tumor-associated macrophages with anti-CSF-1R antibody reveals a strategy for cancer therapy. *Cancer Cell*. 2014;25:846–59.
7. Ruffell B, Coussens LM. Macrophages and therapeutic resistance in cancer. *Cancer Cell*. 2015;27:462–72.
8. Afik R, Zigmund E, Vugman M, et al. Tumor macrophages are pivotal constructors of tumor collagenous matrix. *J Exp Med*. 2016;213:2315–31.
9. Tiainen S, Tumelius R, Rilla K, et al. High numbers of macrophages, especially M2-like (CD163-positive), correlate with hyaluronan accumulation and poor outcome in breast cancer. *Histopathology*. 2015;66:873–83.
10. Sieweke MH, Allen JE. Beyond stem cells: self-renewal of differentiated macrophages. *Science*. 2013;342(6161):1242974.
11. Mildner A, Schmidt H, Nitsche M, et al. Microglia in the adult brain arise from Ly-6ChiCCR2+ monocytes only under defined host conditions. *Nat Neurosci*. 2007;10(12):1544–53.
12. Hamilton JA. Colony-stimulating factors in inflammation and autoimmunity. *Nat Rev Immunol*. 2008;8(7):533–44.
13. Wang Y, Szretter KJ, Vermi W, et al. IL-34 is a tissue-restricted ligand of CSF1R required for the development of Langerhans cells and microglia. *Nat Immunol*. 2012;13(8):753–60.
14. Horwood NJ. Macrophage polarization and bone formation: a review. *Clin Rev Allergy Immunol*. 2016;51:79–86.
15. Gomez Perdiguero E, Klapproth K, Schulz C, et al. Tissue-resident macrophages originate from yolk-sac-derived erythro-myeloid progenitors. *Nature*. 2015;518:547–51.
16. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–74.
17. Enderlin Vaz da Silva Z, Lehr HA, Velin D. In vitro and in vivo repair activities of undifferentiated and classically and alternatively activated macrophages. *Pathobiology*. 2014; 81:86–93.
18. Gao L, Wang FQ, Li HM, et al. CCL2/EGF positive feedback loop between cancer cells and macrophages promotes cell migration and invasion in head and neck squamous cell carcinoma. *Oncotarget*. 2016;7:87037–51.
19. Feriotti C, Loures FV, Frank de Araujo E, et al. Mannosyl-recognizing receptors induce an M1-like phenotype in macrophages of susceptible mice but an M2-like phenotype in mice resistant to a fungal infection. *PLoS One*. 2013;8:e54845.
20. Eguchi J, Kong X, Tenta M, et al. Interferon regulatory factor 4 regulates obesity-induced inflammation through regulation of adipose tissue macrophage polarization. *Diabetes*. 2013;62(10):3394–403.
21. Satoh T, Kidoya H, Naito H, et al. Critical role of Trib1 in differentiation of tissue-resident M2-like macrophages. *Nature*. 2013;495(7442):524–8.
22. Zhou D, Huang C, Lin Z, et al. Macrophage polarization and function with emphasis on the evolving roles of coordinated regulation of cellular signaling pathways. *Cell Signal*. 2014;26(2):192–7.
23. Lavin Y, Merad M. Macrophages: gatekeepers of tissue integrity. *Cancer Immunol Res*. 2013;1(4):201–9.
24. Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol*. 2010;22(2):231–7.
25. Egeblad M, Nakasone ES, Werb Z. Tumors as organs: complex tissues that interface with the entire organism. *Dev Cell*. 2010;18(6):884–901.
26. Colegio OR, Chu NQ, Szabo AL, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature*. 2014;513(7519):559–63.
27. Mills CD, Ley K. M1 and M2 macrophages: the chicken and the egg of immunity. *J Innate Immun*. 2014;6(6):716–26.
28. Liu JY, Yang XJ, Geng XF, et al. Prognostic significance of tumor-associated macrophages density in gastric cancer: a systemic review and meta-analysis. *Minerva Med*. 2016;107(5):314–21.
29. Christofori G. New signals from the invasive front. *Nature*. 2006;441(7092):444–50.
30. Liu JY, Peng CW, Yang GF, et al. Distribution pattern of tumor associated macrophages predicts the prognosis of gastric cancer. *Oncotarget*. 2017;8(54):92757–69.
31. Park JY, Sung JY, Lee J, et al. Polarized CD163+ tumor-associated macrophages are associated with increased angiogenesis and CXCL12 expression in gastric cancer. *Clin Res Hepatol Gastroenterol*. 2016;40:357–65.
32. Forssell J, Oberg A, Henriksson ML, et al. High macrophage infiltration along the tumor front correlates with improved survival in colon cancer. *Clin Cancer Res*. 2007;13(5):1472–9.
33. Su Z, Zhang P, Yu Y, et al. HMGBl facilitated macrophage reprogramming towards a proinflammatory M1-like phenotype in experimental autoimmune myocarditis development. *Sci Rep*. 2016;6:21884.
34. Dungan LS, McGuinness NC, Boon L, et al. Innate IFN- γ promotes development of experimental autoimmune encephalomyelitis: a role for NK cells and M1 macrophages. *Eur J Immunol*. 2014;44:2903–17.
35. Luen SJ, Savas P, Fox SB, et al. Tumour-infiltrating lymphocytes and the emerging role of immunotherapy in breast cancer. *Pathology*. 2017;49:141–55.
36. Lu H, Clauser KR, Tam WL, et al. A breast cancer stem cell niche supported by juxtacrine signalling from monocytes and macrophages. *Nat Cell Biol*. 2014;16:1105–17.
37. Zhou D, Yang K, Chen L, et al. Promising landscape for regulating macrophage polarization: epigenetic viewpoint. *Oncotarget*. 2017;8:57693–706.
38. Ruffell B, Chang-Strachan D, Chan V, et al. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. *Cancer Cell*. 2014;26:623–37.
39. Ho VW, Hofs E, Elisia I, et al. All trans retinoic acid, transforming growth factor β and prostaglandin E2 in mouse plasma synergize with basophil-secreted interleukin-4 to M2 polarize murine macrophages. *PLoS One*. 2016;11:e0168072.
40. Potas JR, Haque F, Maclean FL, et al. Interleukin-10 conjugated electrospun polycaprolactone (PCL) nanofibre scaffolds for promoting alternatively activated (M2) macrophages around the peripheral nerve in vivo. *J Immunol Methods*. 2015;420:38–49.
41. Muenst S, Läubli H, Soysal SD, et al. The immune system and cancer evasion strategies: therapeutic concepts. *J Intern Med*. 2016;279:541–62.
42. Spring H, Schüler T, Arnold B, et al. Chemokines direct endothelial progenitors into tumor neovessels. *Proc Natl Acad Sci U S A*. 2005;102:18111–6.
43. Spiller KL, Anfang RR, Spiller KJ, et al. The role of macrophage phenotype in vascularization of tissue engineering scaffolds. *Biomaterials*. 2014;35:4477–88.
44. Lewis CE, Harney AS, Pollard JW. The multifaceted role of perivascular macrophages in tumors. *Cancer Cell*. 2016;30:18–25.
45. Li GG, Guo ZZ, Ma XF, et al. The M2 macrophages induce autophagic vascular disorder and promote mouse sensitivity to urethane-related lung carcinogenesis. *Dev Comp Immunol*. 2016;59:89–98.
46. Raggi F, Pelassa S, Pierobon D, et al. Regulation of human macrophage M1-M2 polarization balance by hypoxia and the triggering receptor expressed on myeloid cells-1. *Front Immunol*. 2017;8:1097.
47. Butoi E, Gan AM, Tucureanu MM, et al. Cross-talk between macrophages and smooth muscle cells impairs collagen and metalloproteinase synthesis and promotes angiogenesis. *Biochim Biophys Acta*. 2016;1863(7 Pt A):1568–78.
48. Osinsky S, Bubnovskaya L, Ganusevich I, et al. Hypoxia, tumour-associated macrophages, microvessel density, VEGF and matrix metalloproteinases in human gastric cancer: interaction and impact on survival. *Clin Transl Oncol*. 2011;13:133–8.
49. Liu J, Geng X, Li Y. Milky spots: omental functional units and hotbeds for peritoneal cancer metastasis. *Tumour Biol*. 2016;37:5715–26.
50. Chazaud B. Macrophages: supportive cells for tissue repair and regeneration. *Immunobiology*. 2014;219:172–8.
51. Cohen N, Shani O, Raz Y, et al. Fibroblasts drive an immunosuppressive and growth-promoting microenvironment in breast cancer via secretion of Chitinase 3-like 1. *Oncogene*. 2017;36:4457–68.
52. Fang M, Yuan J, Peng C, et al. Collagen as a double-edged sword in tumor progression. *Tumour Biol*. 2014;35:2871–82.
53. Liu XQ, Kiehl R, Roskopf C, et al. Interactions among lung cancer cells, fibroblasts, and macrophages in 3D co-cultures and the impact on mmp-1 and vegf expression. *PLoS One*. 2016;11:e0156268.
54. Hu WQ, Fang M, Zhao HL, et al. Tumor invasion unit in gastric cancer revealed by QDs-based in situ molecular imaging and multispectral analysis. *Biomaterials*. 2014;35:4125–32.
55. Komohara Y, Jinushi M, Takeya M. Clinical significance of macrophage heterogeneity in human malignant tumors. *Cancer Sci*. 2014;105:1–8.

56. Llosa NJ, Cruise M, Tam A, et al. The vigorous immune microenvironment of microsatellite instable colon cancer is balanced by multiple counter-inhibitory checkpoints. *Cancer Discov.* 2015;5:43–51.
57. Lim SY, Yuzhalin AE, Gordon-Weeks AN, et al. Tumor-infiltrating monocytes/macrophages promote tumor invasion and migration by upregulating S100A8 and S100A9 expression in cancer cells. *Oncogene.* 2016;35:5735–45.
58. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell.* 2010;141:39–51.
59. Peng C, Liu J, Yang G, et al. Lysyl oxidase activates cancer stromal cells and promotes gastric cancer progression: quantum dot-based identification of biomarkers in cancer stromal cells. *Int J Nanomedicine.* 2017;13:161–74.
60. Yang C, Wei C, Wang S, et al. Elevated CD163+/CD68+ ratio at tumor invasive front is closely associated with aggressive phenotype and poor prognosis in colorectal cancer. *Int J Biol Sci.* 2019;15(5):984–98.

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