

REVIEW

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The role of Th17 cells in the pathogenesis and treatment of breast cancer

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Abstract

Breast cancer is a severe problem worldwide due to an increase in mortality and prevalence among women. Despite early diagnostic procedures as well as advanced therapies, more investigation is required to find new treatment targets. Various factors and mechanisms, such as inflammatory conditions, can play a crucial role in cancer progression. Among them, Th17 cells are identified as effective CD4⁺ T cells that play an essential role in autoimmune diseases and inflammation which may be associated with anti-tumor responses. In addition, Th17 cells are one of the main factors involved in cancer, especially breast cancer via the inflammatory process. In tumor immunity, the exact mechanism of Th17 cells is not entirely understood and seems to have a dual function in tumor development. Various studies have reported that cytokines secreted by Th17 cells are in close relation to cancer stem cells and tumor microenvironment. Therefore, they play a critical role in the growth, proliferation, and invasion of tumor cells. On the other hand, most studies have reported that T cells suppress the growth of tumor cells by the induction of immune responses. In patients with breast cancer compared to normal individuals, various studies have been reported that the Th17 population dramatically increases in peripheral blood which results in cancer progression. It seems that Th17 cells by creating inflammatory conditions through the secretion of cytokines, including IL-22, IL-17, TNF- α , IL-21, and IL-6, can significantly enhance breast cancer progression. Therefore, to identify the mechanisms and factors involved in the activation and development of Th17 cells, they can provide an essential role in preventing breast cancer progression. In the present review, the role of Th17 cells in breast cancer progression and its therapeutic potential was investigated.

Keywords: Th17, Breast cancer, Targeted therapy

Introduction

Although tremendous advances have been made to identify molecular and cellular pathways in carcinogenesis and cancer genetics, cancer is recognized as one of the main reasons for death worldwide. Breast cancer (BC) is one of the deadliest cancer among women. Notably, BC alongside colon and lung cancer, are the three prevalent malignancies worldwide [1]. Various studies globally have shown that 1 out of 8 women has BC, which 90% of all

cases are related to lifestyle and environmental factors, while 10% are related to genetic disorders [2]. In developed countries, over the next 25 years, researchers predict that BC incidence and the mortality rate will increase up to 60% [3]. Despite advances in new treatment strategies, such as chemoradiotherapy or chemotherapy combined with surgery, there is a long way to find a practical approach to treat and control BC. Therefore, further studies need to be conducted in order to find new strategies to treat cancer [4].

In tumor microenvironment, accumulated tumor cells can inhibit the immune system's responses through various mechanisms [5]. One of these mechanisms is to develop the inflammatory conditions in the

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tumor microenvironment, in which tumor cells cause inflammation by infiltration the leukocytes as well as secreting inflammatory cytokines in the tumor site [6–8]. Studies have shown that inflammatory cells and cytokines accumulated in the tumor microenvironment suppress the immune responses and contribute to tumor cells' growth and development instead of triggering an anti-tumor response [9].

Tumor-associated fibroblasts and tumor cells have been reported to create an inflammatory condition for Th17 cell recruitment [10]. Molecules such as TNF- α , IL-1 β , IL-6, IL-21, TGF- β , and IL-23 which are secreted by the immune and tumor cells, accumulated in the TME, can play a vital role in inducing Th17 development [11–13]. Several studies have reported that IL-23 promotes the development of Th17 cells, while TGF- β , IL-6, and retinoid orphan nuclear receptor (ROR) are significant factors in the differentiation of Th17 cells [12, 14–16]. However, some studies have shown that TGF- β can provide an inhibitory function in the differentiation of human Th17 cells [17, 18]. Beside producing and secreting the cytokines such as IL-22, IL-21, and IL-17 [19, 20], Th17 cells have performed an important function in inducing allergic reactions and autoimmune diseases [21]. Numerous studies have reported that Th17 cell populations have been implicated in various cancers, including liver, ovarian, breast, melanoma, and colon [22]. In tumor immunity, the exact mechanism of the Th17 cells is still unknown and seems to have a dual function in tumor development, not only by promoting but also inhibiting. On the one hand, Th17 cells enhance anti-tumor immune responses by stimulating immune cells in the TME, becoming the Th1 phenotype, producing abundant IFN- γ or inducing effector CD8⁺ T cells. In contrast, Th17 cells via secreting IL-17 and exerting immunosuppressive functions, stimulate angiogenesis and inhibit the immune system from growing, respectively, which result in tumor cells development [23]. In BC, the penetration of the Th17 population is a poor prediction factor [24]. In BC patients' peripheral blood, various studies have shown that the population of Th17 cells sharply increases compared to normal individuals, and these cells are more effective than Th1 cells to kill tumor cells [25]. Notably, in BC tissue, Th17 cells are positively related to the IL-6, IL-1 β , and IL-17 expression, and also negatively correlated with an increase in the number of metastatic lymph nodes and angiogenesis of tumor cells as well [26]. Therefore, in BC tissue, an increase in Th17 cell population could be an ideal strategy to treat BC in future investigations.

In the present study, we will discuss the Th17 role in tumorigenesis and the treatment of BC.

Breast cancer and therapeutic strategies

Despite significant advances in cancer treatment, BC is still one of the prevalent health problems. In the coming years, evidence about BC suggests that its incidence and mortality will dramatically increase [27]. Studies have shown that BC is the prevalent malignancy in women under 45-year-old [28]. Due to its complexity and invasive biological characteristics, BC is highly heterogeneous [29]. Although many studies have evaluated BC etiology, little information is available about disease etiology [30]. Numerous environmental and genetic factors significantly increase the risk of BC morbidity and its recurrence. Environmental and lifestyle factors involved in the development of BC include smoking, sedentary lifestyle, obesity, alcoholism, hormone therapy, and ionizing radiation [31–33]. Genetic factors such as family history, ethnicity, and genetic mutations can also increase the risk of BC [34]. Symptoms of BC are different from person to person. In general, BC symptoms include pain, deformity, size changes, abnormal discharge, swelling, redness, dimpling, and lump formation in the breast [35]. There are different strategies to diagnose BC such as mammography, biopsies, blood tests, genomic assays, and imaging tests [36].

Common treatments for BC include hormone therapy, chemotherapy, surgery, molecular targeted therapy, radiation, and immunotherapy [37]. Various factors such as genetics and disease stages were considered to determine the treatment methods. In the early stages of cancer, most patients undergo surgery but often use chemotherapy as the tumor cells progress and metastasize [38]. Endocrine therapies are commonly used for elderly or infirm patients with hormone-positive tumors [39]. BC is currently divided into 4 subtypes: Luminal A (HER2 -/PR +/ER +), Luminal B (HER2 +/PR +/ER + or HER2 -/PR +/ER +), HER2 overexpression (HER2 +/PR -/ER -), and triple-negative BC (TNBC) (HER2 -/PR -/ER -) [40].

Today, with the development of pharmacogenomics and immunology, immunotherapy has become an effective and ideal strategy in BC treatment [41]. In BC, immunotherapy methods such as stimulatory molecule agonist Ab, immune checkpoint therapy, bispecific Ab, and cancer vaccines are among the techniques researchers have focused (Table 1). In BC, scientists hope that new immunotherapy strategies can modify the side effects and current status of common treatments to have effective and better treatment approaches.

Th17 lymphocytes

Th17 population is a subgroup of CD4⁺ T cells that can produce and secrete various cytokines, including TNF- α , GM-CSF, IL-22, and IL-17 [18]. IL-17-produced by Th17

Table 1 Examples of immunotherapies in clinical trials for breast cancer

Method	Study	Breast Cancer Subtype	Phase	Trial id
Vaccines	CD40L vector vaccine	Breast cancer	I	NCT02140996
Vaccines	HER2 vaccine	Breast cancer	I	NCT01376505
Vaccines	HER2 DC Vaccine	Breast cancer	I/II	NCT02061332
Vaccines	Poly ICLC	TNBC	I	NCT02427581
Vaccines	Trastuzumab + E75	HER2 breast cancer	II	NCT01570036
Vaccines	Chemotherapy + DC vaccine	HER2-/ER+ (phase II) and TNBC (phase I)	I/II	NCT02018458
Bispecific antibodies	68Ga-IMP-288 + TF2	CEA + / HER2- breast cancer	I/II	NCT01730612
PD-1	Vinorelbine + Gemcitaine + Eribulin + Capecitabine + Pembrolizumab	TNBC	III	NCT02555657
PD-1	PDR001	TNBC	I/II	NCT02404441
PD-1	Carboplatin + Gemcitabine + Nivolumab + Nab-Paclitaxel	Breast cancer	I	NCT02309177
PD-1	Pembrolizumab	HER2 + breast cancer	I/II	NCT02129556
PD-1	Nivolumab + Ipilimumab + Entinostat	HER2- breast cancer	I	NCT02453620
PD-1	Poly ICLC + CDX-1401 + Pembrolizumab	TNBC	I/II	NCT02661100
PD-L1	Atezolizumab + Entinostat	TNBC	I/II	NCT02708680
PD-L1	Trastuzumab + Durvalumab	HER2 + breast cancer	I	NCT02649686
PD-L1	Atezolizumab	TNBC	II	NCT02478099
PD-L1	Placebo + Nab-Paclitaxel + Atezolizumab	TNBC	III	NCT02425891
PD-L1	Durvalumab + Paclitaxel	TNBC	I/II	NCT02628132
CTLA-4	Ipilimumab + MGA271	TNBC	I	NCT02381314
CTLA-4	Tremelimumab + MEDI4736	HER2- breast cancer	II	NCT02536794
4-1BB	Avelumab + PF-05082566	TNBC	II	NCT02554812
OX40	MEDI6469	Breast cancer	I/II	NCT01862900
LAG-3	Paclitaxel + Placebo + IMP321	Stage IV	II	NCT02614833

cells was first identified in the host immune response to *Borrelia burgdorferi* [42]. Vital factors needed for Th17 cell development, including retinoic acid-related (RAR), orphan receptor gamma (ROR- γ t), orphan receptor alpha (ROR- α), and STAT3 as a transcription factor. Also, the detection of TGF- β and IL-6 cytokines as Th17 differentiators show that Th17 cells are a separate subgroup of CD4 T cells, easily distinguished from Th1 and Th2 cells [43, 44].

About the site of Th17 cells, the lamina propria is the leading site, but it can be present in epithelial and mucosal tissues when fighting fungi, viruses, and extracellular bacteria [45]. IL-17 produced by Th17 cells can increase the secretion of inflammatory cytokines, including IL-6, TNF- α , and IL-1 β , not only by enhancing the production of chemokines such as CXCL-2 and CXCL-8 but also by secreting significant intestinal stimuli such as GM-CSF. In inflammatory tissue, GM-CSF causes the accumulation of granulocytes and granulopoiesis, which lead to inflammation [46–48]. IL-22 and IL-17 cause the secretion of antimicrobial proteins and peptides such as S100 proteins and β -defensins by keratinocytes [49]. Th17 population can play a crucial role in increasing

immune responses by enhancing the function of CD8 + T cells and B cells [50–52]. Studies have also shown that there is a direct correlation between Th17 cells and myeloid cells. Ronchi et al. have shown that the presence of pertussis toxin (PTX) during immunization causes the production of IL-1 β by myeloid cells with CD11b⁺ CCR2⁺ Gr1⁺ phenotype, thus increases the activation of Th17 cells [53]. Also, Pang and colleagues reported that myeloid-derived suppressor cell (MDSC)-derived arginase-1, miR-322-5p, and TGF- β can promote Th17 cell differentiation [54]. Recent studies have reported that Th17 cells are often correlated with various cancers such as lung, breast, prostate, colon, and melanoma [55], and also autoimmune diseases such as multiple sclerosis (MS), inflammatory bowel disease (IBD), and rheumatoid arthritis [56–60].

Plasticity, differentiation, and development of Th17

Unlike Th2 and Th1 cells, which are relatively stable, Th17 cells have a very high degree of plasticity [61]. As shown in Fig. 1, studies have shown that Th17 cells can mainly transform into TFH, Th2, TR1, Treg, and Th1

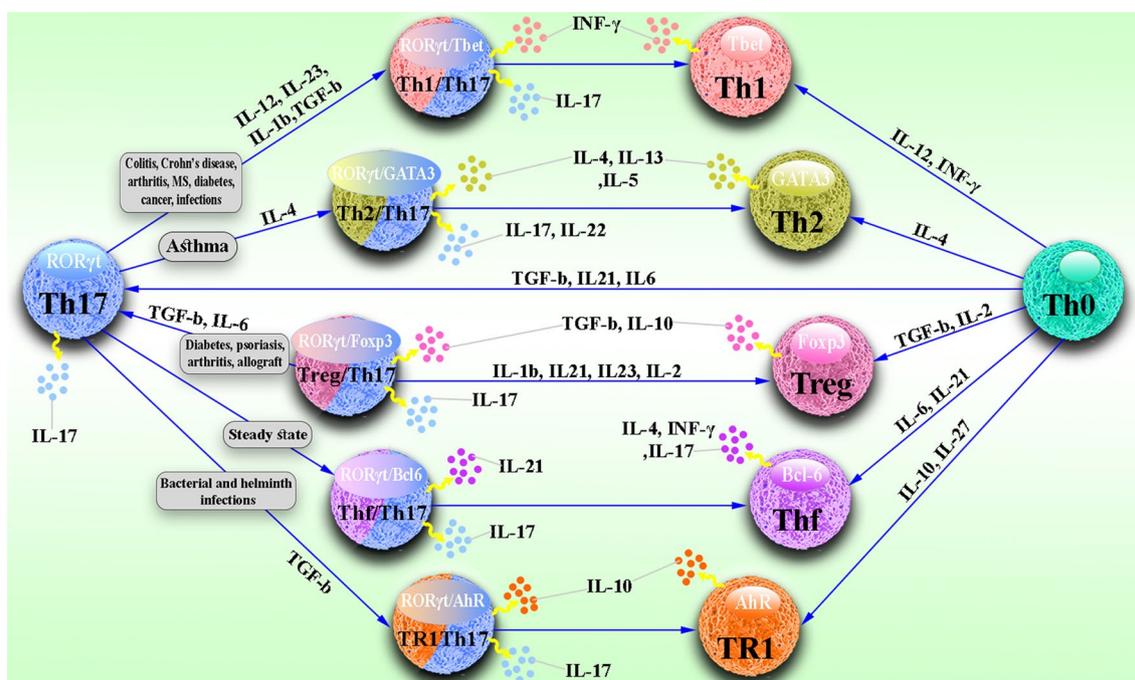


Fig. 1 Plasticity of Th17. Th17 cells can mainly transform into TFH, Th2, TR1, Treg, and Th1 cells and exhibit a variety of contrasting functions depending on environmental conditions. During infections and autoimmune diseases, Th17 cells can acquire immune inhibitory functions by becoming TR1 or Treg cells. Th17 cells also change to TFH cells at a steady-state and participate in promoting IgA-producing B cells. Also, Th17 cells acquire pathogenic activity by changing to Th2 cells during asthma or Th1 cells during infection, cancer, and autoimmune diseases

cells which exhibit a variety of contrasting functions depending on environmental conditions. Th17 cells can acquire immune inhibitory functions by changing to TR1 or Treg cells during infections and autoimmune diseases. Th17 cells also modify to TFH cells at a steady state and participate in promoting IgA-producing B cells. Most importantly, Th17 cells acquire pathogenic activity by changing to Th2 cells during asthma or Th1 cells during infection, cancer, and autoimmune diseases [23].

For almost two decades, many studies have been conducted on the molecular mechanisms involved in Th17 cell differentiation. Data have shown that various factors and cytokines engaged in this differentiation (Fig. 2) [62]. One of these cytokines is IL-23, which can play an essential role in Th17 cells differentiation by inducing STAT3 as well as inhibiting T-bet and FoxP3 [63]. In naive T cells, it has been confirmed that the expression of IL-23R is in a poor state, that is why it has no role in the early stages of Th17 differentiation from CD4⁺ T cells, but it has a vital role in Th17 activity and final differentiation [43, 64]. However, other key cytokines may be involved in Th17 differentiation, includes TGF-β, IL-21, and IL-6 cytokines. Notably, the TGF-β mechanism in the differentiation of Th17 cells is to suppress transcription factors, including GATA-3 and T-bet, needed for Th2 and

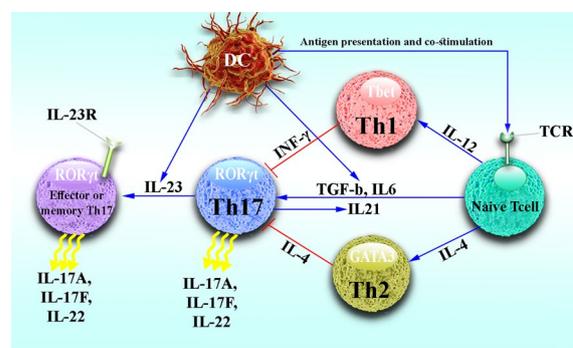


Fig. 2 Differentiation of Th17 cell. Naive T cells can be differentiated into three subsets of T helper cells by different cytokines. IL-4 and IL-12 cytokines can increase the differentiation of Th2 and Th1 cells, respectively, and the cytokines secreted by these cells suppress the differentiation of Th17 cells. In contrast, the cytokines IL-23, IL-6, TGF-β, and IL-21 enhance Th17 cell differentiation

Th1 differentiation [65–67]. However, Schumann and colleagues reported that TGF-β could inhibit the production and differentiation of Th17 cells in mice [68]. Despite the auto-proliferative role of IL-21 in Th17 differentiation, the data shows that its absence does not make any difference in Th17 differentiation [69]. Of note, IL-6

can regulate IL-21 function through the STAT3-dependent pathway by direct induction of IL-21 gene expression [70, 71]. Studies have reported that IL-6 can act as a positive enhancer of Th17 differentiation by inducing IL-21 [72, 73]. By signaling pathways such as STAT3, ROR- α , and ROR- γ t, Th17 cells are differentiated from other subtypes of T helper cells. Naive T cells produce IL-6 and TGF- β to activate the ROR- α and ROR- γ t signaling pathways, which lead to the differentiation of Th17 cells from CD4⁺ T cells [74, 75]. Studies have shown that IL-23, IL-6, and IL-21 cytokines play an essential role in expressing transcription factors which is necessary for Th17 cell differentiation by the activation of STAT3 signaling [63]. In mouse T cells, experiments have shown that mutations or lack of STAT3 expression can result in cancer and metabolic disorders in vivo [76, 77]. It was documented that it could be effective to use antibodies such as ustekinumab, canakinumab, ixekizumab, secukinumab, anakinra, and tocilizumab to suppress Th17 cell differentiation [76, 78–81].

Numerous studies have shown that cytokines required for Th1 differentiation, such as IL-12 and IFN γ , can inhibit the growth and development of Th17 cells [82]. In contrast, IFN γ induces T-bet expression, which stimulates the development of cells by binding and activating the IL-23R promoter. It should be noted that T-bet overexpression reduces IL-17 production [83, 84]. STAT4 and STAT1, as important transcription factors for Th1 differentiation, are significant inhibitors of Th17 development because IL-17 production has been reported to be increased in STAT1-deficient T cells [82]. IL-27 produced by DCs and macrophages, like IFN- γ , can inhibit Th17 cell proliferation by STAT1 activation [85]. Studies have shown that IL-4, STAT6, and GATA-3, involved in Th2 differentiation, can inhibit the development of Th17 cells [82, 86]. IL-25, belonging to the IL-17 cytokine family, can inhibit the development of Th17 by an increase in IL-13 expression. Notably, IL-13 can suppress Th17 development by inhibiting IL-23, IL-1, and IL-6 [87]. Based on previous studies, transcription factors such as STAT5 and cytokine IL-2, are essential factors for Treg cells' development, can effectively inhibit Th17 cell growth [88–90]. About another transcription factor called Ets-1, it also negatively regulates Th17 development by increasing IL-2 production. Deficiency or mutation of Ets-1 increases the IL-23R and IL-22 expression level, thereby promoting the development and differentiation of Th17 [91].

Th17 in breast cancer

This section will evaluate studies on Th17 function and its potential in BC treatment (Table 2).

The role of Th17 in the progression of breast cancer

By collecting blood samples from 55 BC patients (HER2+/PR+/ER+) and 34 normal individuals, Baharlou et al., observed that serum levels of Th17-related cytokines, including IL-17, TGF- β , and IL-6, were significantly less than healthy individuals using the ELISA method. They reported that a serum decrease in TGF- β , IL-17, and IL-6 cytokines could be prognostic markers and predictors of the clinical stage of BC. It was concluded that a reduction in serum levels of TGF- β , IL-6, and IL-17 indicated the potential suppressive effects of radiation therapy and chemotherapy on Th17 cells and TGF- β -producing tumor cells in the early stages of BC patients. It was suggested that considering the balance between Th17 and Treg cells using antibodies against Th17-related cytokines could be an ideal strategy for immune therapy in BC [92]. In a study on BALB/c mice, Du and colleagues reported that IL-17 as one of the most critical cytokines of Th17 cells was highly expressed in breast tumor tissue. To determine the level of IL-17 expression in breast tumors, the tumor tissue of mice was examined by ELISA assay on day 5 after inoculation with 4T1 and MA782 cancer cells. They observed that IL-17 increased angiogenesis, metastasis, tumor cells proliferation and growth, and BC progression rate by raising microvessel accumulation in the tumor area. In addition, they stated that injection of IL-17 into tumor-bearing mice dramatically increased tumor progression, whereas in vitro exposure to IL-17 did not contribute to the growth of tumor cells. This means that IL-17 indirectly causes tumor cells to proliferate. That is why this cytokine can be targeted in order to inhibit the development of BC [93]. Accordingly, studying the TME of 207 patients with BC (Luminal A, Luminal B, HER2, Triple-negative), Chen and coworkers examined the frequency of IL-17 secreting cells and its association with pathological and clinical features by immunohistochemical assay. Their data showed that the high level of IL-17 secretory cells was directly related to the triple-negative molecular subgroup, ER/PR negative, and high histological grade in BC. They concluded that Th17 cells could become a major pharmacological target by inhibiting the major regulator ROR- γ t, a strategy that could be promising because the predominant role of Th17 cells in several autoimmune diseases has been demonstrated. Therefore, they suggested that these cells were a poor prognostic agent in the TME of BC [56]. In another study on tumor samples of 40 patients (age: 43–73) and 4T1 tumor-bearing mice, it is reported that IL-22, another important cytokine produced by Th17 cells, increased the rate of angiogenesis, proliferation, and tumorigenesis by inducing the expression of Pin1 and MAP3K8, which is directly correlated to the BC progression, as investigated by

Table 2 Research correlated to the function of Th17 Cell in breast cancer

Cell line	Treatment	Mice/ Human	In vitro/in vivo	Result	References
-	Anti-FoxP3 and anti-IL-17 antibodies	Human	In vivo	Increasing the population of Th17 cells in breast cancer tissues enhances anti-tumor immune responses	[26]
-	Anti-IFN γ , anti-IL-13, anti-IL-17A, anti-CD8, anti-CD4, anti-CD3 and anti-CD45	Human	In vivo	Th17 is a new prognostic compound bio-marker in TNBC patients	[106]
-	-	Human	In vivo	Serum levels of Th17-related cytokines, including IL-17, TGF- β , and IL-6 in the serum of breast cancer patients, were significantly lower than a healthy individual	[92]
MA782 and 4T1	Anti-CD3 and anti-CD28 antibody	BALB/c mice	In vivo/in vitro	IL-17, one of the most critical cytokines in Th17 cells, was highly expressed in breast tumor tissue	[93]
-	Goat polyclonal anti-human IL-17 antibody	Human	In vivo	IL-17-producing cells accumulated in breast cancer tumor tissue, and these cells were a poor prognostic factor	[56]
-	Anti-PD-1, anti-CD25, anti-CD45RO, anti-CTLA-4, anti-CD103, anti-GITR, anti-CD8, anti-CD3, anti-Foxp3, anti-CD4 and anti-IL-17A antibody	Human	In vivo	Th17 and Treg cells increased significantly in the breast cancer tissue	[96]
MCF-7	Anti-Ly6C, anti-CD11b, anti-CD11c, anti-CD40, anti-Gr-1, and anti-F4 80	Human/ BALB/c mice	In vivo/in vitro	IL-17 produced in tumor tissue prevented the accumulation of MDSCs in breast cancer tumor tissue by activating STAT3 signaling	[107]
JB6 Cl41, MCF7, MEF and MDA-MB231	MAP3K8 inhibitor, PD98059, anti-ERK1, SP600125, anti-MEK1/2, anti-STAT3, anti-MAP3K8, anti-c-Jun, anti-IL-22R1, anti-JNK1/2, and anti-IL22 antibody	BALB/c mice	In vivo/in vitro	IL-22, by inducing the expression of Pim1 and MAP3K8, increase the rate of angiogenesis, proliferation, and tumorigenesis of tumor cell	[94]
-	IL-17A, anti-CD4, anti-CD25, anti-CD127, IgG2a, G1b, anti-CD3 and anti-CD28 antibody	Human	In vivo	Gradually, with breast cancer progression, Treg cells' accumulation increased, and the population of Th17 cells decreased	[95]
-	Anti-CD28, anti-CCR4, anti-CD39, anti-CD4, anti-CD45RA, anti-CD25, anti-CXCR3, anti-CD3, anti-IL-17A, anti-ROR- γ t, anti-CCR6 and anti-Foxp3 antibody	Human	In vivo	The ectonucleotidase-expressing CD25 ^{high} + Th17 increases dramatically in tumor tissue and exhibits suppressive function inhibiting CD8 and CD4 cell activation	[97]
MDA-MB-231 and MCF-7	Rabbit anti-sheep anti-rabbit IgG, rabbit anti-human pNF- κ B, rabbit anti-human pAKT, Rabbit anti-human IL-17 antibody, rabbit anti-human VEGF, anti-human MMP-9, rabbit anti-human Bcl-2, rabbit anti-human CyclinD1, rabbit anti-human EGFR, anti-human CXCR2, human CXCL1, mouse anti-IL-17A, and mouse anti-CD4 antibody	Human	In vivo/in vitro	Th17 cells regulate CXCL1 expression during cancer progression, and CXCL1 by binding to the CXCR2 receptor could promote the NF- κ B/ AKT pathway activation, thereby causing the progression of breast cancer	[98]

Table 2 (continued)

Cell line	Treatment	Mice/ Human	In vitro/in vivo	Result	References
MDA-MB-231	Anti- TNF- α , anti-IL-17A, anti-CD3, anti- IFN- γ , anti-IL-18R α , anti-CD161, anti-MAIT, anti-V α 7.2 TCR, and anti- CD56 and anti-CD45 antibody	Human	In vitro	Breast tumor cells exposed to bacteria selectively activate Th17-polarized MAIT cells from the mammary ducts, thereby increasing breast cancer progression	[99]
-	-	Human	In vivo	A positive relationship between Th17 cells expressing IL-17A and MIF and increasing the expression of both Th17 and MIF increased breast cancer risk	[100]
4T1	PRI-2191 and Calcitriol anti-mouse CD4, anti-CD35, anti-CD25, anti-CD3, anti-CD19, and anti-CD8a antibody	BALB/c mice	In vivo/in vitro	In young mice treated with PRI-2191, unlike older O VX mice, the expression of Th17, ROR- γ t, and ROR- α transcription factors, as well as genes encoding vitamin D receptor and osteopontin, were significantly increased	[108]
-	Anti-Ly6G-, anti-CCR6, anti-V γ 1, anti-VEGFR1, anti- CCR2, anti-IL23R, and anti-IL1R1	KEP; Tcr δ -/-	In vivo	The $\gamma\delta$ T cells by production of IL-17A induces neutrophils to promote distant metastases and inhibit CD8+ T Cells and	[101]
MCF7, JB6 Cl41	anti-TPL2, anti-IL-17A, anti- DAB, and anti-PCNA	-	In vitro	IL-17A induces AP-1 activity and the growth and proliferation of breast cancer cells by activating TPL2	[102]
EMT6, MA782,4THM	Anti-IL-17, anti-CD-3, anti-CD28	BALB/c	In vitro/ In vivo	IL-17 cytokine significantly increased tumor cell proliferation	[93]
4T1, MDA-MB231, EMT6, MDA-MB435, Hs578t	Anti- CD24, anti- CD29, anti- CD25, anti- CD4, anti-CD45, anti-GPR56, anti-Scara5, anti-Tgf β r1, anti-Smad4, anti-Smad2/3, anti-Smad5, anti-Smad6, anti-NF-kB	BALB/c	In vitro/ In vivo	Treg cells induced IL-17RB expression in breast cancer cells by secreting TGF- β 1	[103]
MDA-MB-435	Anti- CCR3, anti- GAPDH, anti- I κ B alpha, anti-VEGF	CD1-nude	In vivo/ In vitro	The IL-17E cytokine has a high ability to induce anti-tumor responses in vitro and in vivo	[104]
-	Anti- IL-25, anti-CD45, anti-CD45R, anti-CD3, anti-CD11b, anti-CD4, anti-CD8, anti-CD49b, anti- $\gamma\delta$ TCR, anti-MHC class II, anti-F4/80 anti-CD25, anti-Ly6G, anti-ST2 and anti-IL-17RB	MMTV-PyMT mice	In vivo/ In vitro	IL-17E increase the growth, proliferation, and metastasis of tumor cells	[105]

immunohistochemical staining, western blot, and CAM assay. It was suggested that targeting Pin1 and MAP3K8 could be an ideal approach to reduce BC progression [94].

In a study of 32 patients with BC who underwent surgery (stage I and II-IV), Wang et al. measured the frequency of Th17 and Treg cells as well as their cytokine levels using flow cytometry and ELISA assay, respectively. They found that Treg and Th17 cells were highly accumulated in the early stage of BC. In BC progression, the population of Treg and Th17 cells was gradually increased and decreased, respectively. In advanced disease, it was stated that reduced Th17 might reflect the cytokine profile in recurrent and chronic inflammation. In contrast, enhanced Th17 levels in early BC may reflect the cytokine profile in acute inflammation. As a result, to identify the involved mechanisms to regulate Th17/Treg cell balance, BC progression could be effectively prevented [95]. Additionally, in another study on the peripheral blood samples and tumor tissue of 39 patients with BC by immunohistochemistry and RT-PCR assays, it is observed that Th17 and Treg cells significantly increased in the BC tissue, promoting the growth and development of tumor cells. They also found that IL-17A is produced by CD4⁺ and CD8⁺ T lymphocytes was positively related to Th17-related molecules such as CCR6, IL-17A, and ROR- γ t. In addition, they reported that vascular endothelial growth factors CXCL8, MMP-2, and MMP-9 were highly expressed by IL-17, there by promoting the growth of tumor cells. These data showed that both tumor invasion and increased IL-17A expression were directly related to the enrichment of Treg cells in invasive breast tumors, suggesting that the regulation of the Treg/Th17 axis could help control BC progression [96].

In a cohort study, Thibaudin and colleagues studied tumor tissues and PBMCs of BC patients (Cohort 1:36 patients who underwent surgery, Cohort 2: 145 patients treated by adjuvant therapy and surgery). ELISA was used to assess cytokine secretion, the immunofluorescence assay to detect the CD73 and CD39 marker, and the immunohistochemistry analysis to investigate the IL-17 and CD8. They reported that ectonucleotidase-expressing CD25^{high} Th17 cells dramatically increased in tumor tissue and exhibited suppressive function via inhibition and activation of CD8 and CD4 cells, respectively. The results showed that these cells not only produced Th17-related cytokines but also induced the Foxp3 and ROR- γ t genes. They also found that the cytokines TGF- β and IL-6 were responsible for expressing CD39 ectonucleotidase on these cells. They suggested that the accumulation of ectonucleotidase-expressing CD25^{high} Th17 cells in tumor tissue interfered with the immune system's antitumor responses. Therefore, targeting these cells

could be an attractive way to boost the immune system [97].

Additionally, in another study on 46 patients (age: 23–65) with BC (stage I, II, III, and IV) by immunohistochemistry, ELISA, and flow cytometry assays, it has been shown that Th17 cells regulate CXCL1 expression during cancer progression. It is demonstrated that CXCL1 expressed on tumor cells and its binding to the CXCR2 receptor could promote the activation of the NF- κ B/AKT pathway, thereby causing metastasis, angiogenesis, growth, and progression of BC. It is recommended that both downregulation of Th17 and blockade of CXCL1 could help to prevent BC progression [98]. In another study, it is reported that there is a direct link among the breast microbiome functions, mammary duct epithelial cells, and MAIT cells which might play a role in BC progression. These findings showed that breast tumor cells exposed to bacteria selectively activate Th17-polarized MAIT cells from the mammary ducts and enhance BC progression. Therefore, it can be identified how these cells promote the progression of BC, and targeting them may be an ideal therapeutic strategy in future studies [99].

Avalos-Navarro and colleagues, by a study on 150 patients with BC (Luminal A, Luminal B, HER2, Triple-negative), assessed Th17 cytokines profile and the macrophage migration inhibitory factor (MIF) by ELISA assay. They reported there is a positive relationship between Th17 cells expressing IL-17A and MIF. They observed that IL-17A and MIF are higher in aggressive molecular subtypes TN, Luminal B, and HER2 compared to less aggressive Luminal A. Their results showed that an increase in the expression of both Th17 and MIF enhanced the risk of BC, indicating that simultaneous blockade of these factors could be an effective strategy to inhibit BC progression [100].

There are studies that have examined the effect of IL-17 on BC cells and the importance of this cytokine and Th17 cells in the metastasis and invasion of BC. Coffelt and colleagues reported that IL-17-producing neutrophils and $\gamma\delta$ cells increased the potency of the invasion and metastasis in BC cells.

It was shown that breast tumor cells induced polarization and proliferation of neutrophils, mediated by granulocyte colony-stimulating factor (G-CSF), which led to the suppression of CD8⁺ T cells as well as an increase in metastasis to surrounding organs. Notably, inhibiting the $\gamma\delta$ T cell/IL-17/neutrophil axis could be an effective therapeutic approach in metastatic disease [101]. Kim et al. also reported that IL-17A induced AP-1 function and TPL2 activation, which resulted in the growth and proliferation of BC cells. They showed that IL-17A increased the accumulation and growth of JB6 Cl41 cells,

while the TPL2 kinase inhibitor prevented IL-17A-induced tumorigenesis. Therefore, blocking this pathway could prevent the proliferation and invasion of BC cells [102]. In another study, Du et al. examined the role of IL-17 producing cells in BC cells growth and angiogenesis. Although the exposure of 4T1 mouse BC cells to recombinant IL-17 had little effect on cancer cell proliferation in vitro, administration of this cytokine to cancer mice significantly increased tumor size, associated with increased angiogenesis. In the tumor area, these effects were due to IL-17 secretion from lymphocytes and to a less extent from cancer cells [93]. In another study, Huang and colleagues reported that Treg cells induced IL-17RB expression in BC cells by secreting TGF- β 1 and activating the Smad2/4/3 signaling pathway in the tumor-draining LNs (TDLNs), thereby increasing proliferation, angiogenesis, and metastasis of cancer cells. They suggested that blocking IL-17RB could be an effective therapeutic target for inhibiting metastasis and cancer progression [103].

Also, it has been reported that the IL-17E cytokine has a high ability to induce anti-tumor responses in vitro and in vivo. The number of eosinophils and serum level of IL-5 was significantly increased in the peripheral blood of tumor-bearing mice treated with IL-17E, which was related to the anti-tumor activity of IL-17E. They suggested that IL-17E in combination with immunotherapy or chemotherapy could be an ideal treatment approach to reduce the progression of BC [104]. In contrast, Jiang and colleagues reported that IL-17E was significantly secreted by macrophages and CD4⁺ T cells in the MMTV-PyMT breast tumor model. It was demonstrated that inhibition of IL-17E decreased macrophages and type 2T cells in the tumor microenvironment as well as causing a reduction in growth, proliferation, and metastasis of tumor cells. They reported that blocking this cytokine could be a valuable treatment for metastatic BC [105]. In these two current studies, because of the differences in the protocols and working conditions, it is necessary to study the role of IL-17E in the treatment or progression of BC in another investigation.

The role of Th17 in the preventing breast cancer

Through immunohistological staining and flow cytometry assays, Yang and colleagues indicated that the number of Th17 cells was dramatically increased compared to normal subjects by studying 30 patients with BC. Also, it was found that Th17 cell count was positively related to the expression of IL-6, IL-17, and IL-1 β cytokines and negatively associated with an increase in the number of metastatic lymph nodes and tumor cell angiogenesis. Notably, an increase in the number of Th17 cells enhanced anti-tumor immune responses in BC tissues.

Therefore, it can be an appropriate treatment choice to enhance the population of Th17 cells in cancerous tissue of BC patients [26]. Accordingly, Faucheux and colleagues studied 106 patients with BC (Luminal A-B, TN, and HER2+) through Luminex and flow cytometry assays. They reported that Th17 is a new prognostic biomarker in TNBC patients. Due to an increase in Th17 population, it has a good prognosis to identify TNBC. Also, their results confirm a direct correlation between increasing the population of Th17 cells and improving the patients' survival. Thus, the accumulation of Th17 cells can effectively induce anti-tumor immune responses, preventing BC progression in BC tissues [106].

By studying 80 patients with BC and MCF-7 tumor-bearing mice through western blot, RT-PCR, flow cytometry, and ELISA assays, Ma and colleagues demonstrated that myeloid-derived suppressor cells (MDSCs) significantly increased in BC tissue. They reported that produced IL-17 prevented the accumulation of MDSCs in BC tumor tissue by activating STAT3 signaling. They suggested that STAT3 signaling can regulate IL-17 expression. Therefore, a combination of p-STAT3 and IL-17 expressing cells could be an effective treatment strategy for BC [107].

Pawlik et al. also examined the expression and activity of Th17 cells in young and old BALB/c mice with 4T1 tumors treated with tacalcitol (PRI-2191) and calcitriol. Their results showed that in young mice treated with PRI-2191, unlike older OVX mice, the expression of Th17 and its cytokines, ROR- γ t and ROR- α as transcription factors as well as genes encoding vitamin D receptor and osteopontin were significantly increased. In young mice, it was suggested that treatment with PRI-2191 increased osteopontin expression, which could enhance the differentiation of Th17 cells as an ideal method to treat BC [108].

Conclusion

Due to the increased mortality and global prevalence of BC especially among women, this disease is a global problem. Today with extensive advances in the medical field, no definitive cure has been reported for BC [27]. However, much research has been done to develop effective treatment strategies and identify the mechanisms and factors involved in the progression of BC. Common systematic treatments for BC include immunotherapy, molecular target therapy, endocrine therapy, radiotherapy, chemotherapy, and surgery [109]. Numerous factors and mechanisms have been identified in BC spread, among which Th17 cells can cause the expansion of BC by producing chemokines, cytokines, and inflammatory pathways [23]. Numerous studies have reported that the frequency of Th17 cells is increased

in tumor tissue and PBMC in BC patients. The activity of these cells is dramatically enhanced in patients with advanced disease. However, some studies have reported that Th17 cells inhibit BC progression by promoting the immune system [110, 111]. Observing these contradictions can have various reasons. The first point about these contradictions is the very small number of studies that declare the protective role of Th17 cells in BC patients. Some of these few studies are based on animal models, and it is not easy to generalize the results to human studies. Therefore, it is highly recommended that more comprehensive studies should be conducted in this field. Another point that may shed some light on these discrepancies is the within-patient difference in BC phenotypes, evaluated in different investigations. As we know, different phenotypes of BC patients have many differences in terms of immune responses, disease pathogenesis, and response to treatment. Therefore, in future studies, comprehensive comparisons should be made on the frequency and function of Th17 cells in different disease phenotypes. It is possible to compare these cells in different phenotypes of BC accurately. In many investigations, another point has probably been overlooked is the study of Th17 cells expressing Treg factors such as FoxP3. Studies show that Th17 cells are involved in BC progression by inducing cells that express the markers of Th17 and Treg cells (CD25^{high} Th17 cells). Therefore, it is essential to evaluate these cell population in future studies. Another group of studies that can be beneficial is to use BC transgenic mice lacking Th17 cells. Finally, in mouse models of BC, silencing gene expression of key transcription factors in Th17 cell differentiation can answer many questions about the role of Th17 cells in BC. In addition to cancer, Th17 cells can promote autoimmune and inflammatory diseases such as psoriasis and RA [112, 113]. Due to the critical role of Th17 population in progression of various disorders, understanding mechanisms involved in inducing and activating Th17 cells as well as pathways inhibiting Th17 inflammatory cytokines could be the subject of the exciting field for future studies.

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Authors' contributions

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Competing interests

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References

- Ishaque N, Asad M. Clinicopathological characteristics of breast carcinoma in premenopausal women. *J Islamabad Med Dental College*. 2019;8(4):193–7.
- Castelló A, Martín M, Ruiz A, Casas AM, Baena-Cañada JM, Lope V, Antolín S, Sánchez P, Ramos M, Antón A. Lower A risk among women following the World Cancer Research Fund and American Institute for Cancer Research lifestyle recommendations: EpiGEICAM case-control study. *PLoS ONE*. 2015;10(5):e0126096.
- Villarreal-Garza C, Aguila C, Magallanes-Hoyos MC, Mohar A, Bargalló E, Meneses A, Cazap E, Gomez H, López-Carrillo L, Chávarri-Guerra Y. Breast cancer in young women in Latin America: an unmet, growing burden. *Oncologist*. 2013;18(12):1298.
- Alvarez RH, Valero V, Hortobagyi GN. Emerging targeted therapies for breast cancer. *J Clin Oncol*. 2010;28(20):3366–79.
- Coleman RE, Gregory W, Marshall H, Wilson C, Holen I. The metastatic microenvironment of breast cancer: clinical implications. *The Breast*. 2013;22:S50–6.
- Karpisheh V, Joshi N, Zekiy AO, Beyzai B, Hojjat-Farsangi M, Namdar A, Edalati M, Jadidi-Niaragh F. EP4 receptor as a novel promising therapeutic target in colon cancer: running title: EP4 receptor in colon cancer. *Pathol-Res Pract*. 2020;216:153247.
- Karpisheh V, Nikkhoo A, Hojjat-Farsangi M, Namdar A, Azizi G, Ghalamfarsa G, Sabz G, Yousefi M, Yousefi B, Jadidi-Niaragh F. Prostaglandin E2 as a potent therapeutic target for treatment of colon cancer. *Prostaglandins Other Lipid Mediators*. 2019;144:106338.
- Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. *J Immunol Res*. 2014;2014:1.
- Mantovani A, Marchesi F, Porta C, Sica A, Allavena P. Inflammation and cancer: breast cancer as a prototype. *The Breast*. 2007;16:27–33.
- Su X, Ye J, Hsueh EC, Zhang Y, Hoft DF, Peng G. Tumor microenvironments direct the recruitment and expansion of human Th17 cells. *J Immunol*. 2010;184(3):1630–41.
- Nam J-S, Terabe M, Kang M-J, Chae H, Voong N, Yang Y-a, Laurence A, Michalowska A, Mamura M, Lonning S. Transforming growth factor β subverts the immune system into directly promoting tumor growth through interleukin-17. *Cancer Res*. 2008;68(10):3915–23.

12. Neel J-C, Humbert L, Lebrun J-J. The dual role of TGF β in human cancer: from tumor suppression to cancer metastasis. *Int Scholarly Res Notices*. 2012;2012:1.
13. Shime H, Yabu M, Akazawa T, Kodama K, Matsumoto M, Seya T, Inoue N. Tumor-secreted lactic acid promotes IL-23/IL-17 proinflammatory pathway. *J Immunol*. 2008;180(11):7175–83.
14. Volpe E, Servant N, Zollinger R, Bogiatzi SI, Hupé P, Barillot E, Soumelis V. A critical function for transforming growth factor- β , interleukin 23 and proinflammatory cytokines in driving and modulating human T H-17 responses. *Nat Immunol*. 2008;9(6):650.
15. Manel N, Unutmaz D, Littman DR. The differentiation of human T H-17 cells requires transforming growth factor- β and induction of the nuclear receptor ROR γ t. *Nat Immunol*. 2008;9(6):641.
16. Zhou Z, Sun W, Liang Y, Gao Y, Kong W, Guan Y, Feng J, Wang X. Fenofibrate inhibited the differentiation of T helper 17 cells in vitro. *PPAR Res*. 2012;2012:1.
17. Karpisheh V, Mousavi SM, Sheykholeslami PN, Fathi M, Mohammadpour Saray M, Aghebbati-Maleki L, Jafari R, Zolbanin NM, Jadidi-Niaragh F. The role of regulatory T cells in the pathogenesis and treatment of prostate cancer. *Life Sci*. 2021;119132.
18. Wilson NJ, Boniface K, Chan JR, McKenzie BS, Blumenschein WM, Mattson JD, Basham B, Smith K, Chen T, Morel F. Development, cytokine profile and function of human interleukin 17–producing helper T cells. *Nat Immunol*. 2007;8(9):950–7.
19. Tesmer LA, Lundy SK, Sarkar S, Fox DA. Th17 cells in human disease. *Immunol Rev*. 2008;223(1):87–113.
20. Weaver CT, Hatton RD. Interplay between the TH 17 and T Reg cell lineages: a (co-) evolutionary perspective. *Nat Rev Immunol*. 2009;9(12):883–9.
21. Chen Z, Laurence A, O’Shea JJ. Signal transduction pathways and transcriptional regulation in the control of Th17 differentiation. In: Elsevier, 2007; 400–408.
22. Zou W, Restifo NP. TH 17 cells in tumour immunity and immunotherapy. *Nat Rev Immunol*. 2010;10(4):248–56.
23. Guéry L, Hugues S. Th17 cell plasticity and functions in cancer immunity. *BioMed Res Int*. 2015;2015:1.
24. Wang S, Li Z, Hu G. Prognostic role of intratumoral IL-17A expression by immunohistochemistry in solid tumors: a meta-analysis. *Oncotarget*. 2017;8(39):66382.
25. Muranski P, Boni A, Antony PA, Cassard L, Irvine KR, Kaiser A, Paulos CM, Palmer DC, Touloukian CE, Ptak K. Tumor-specific Th17-polarized cells eradicate large established melanoma. *Blood, J Am Soc Hematol*. 2008;112(2):362–73.
26. Yang L, Qi Y, Hu J, Tang L, Zhao S, Shan B. Expression of Th17 cells in breast cancer tissue and its association with clinical parameters. *Cell Biochem Biophys*. 2012;62(1):153–9.
27. DeSantis C, Ma J, Bryan L, Jemal A. Breast cancer statistics, 2013. *CA Cancer J Clin*. 2014;64(1):52–62.
28. Anastasiadi Z, Lianos GD, Ignatiadou E, Harissis HV, Mitsis M. Breast cancer in young women: an overview. *Updat Surg*. 2017;69(3):313–7.
29. Reyna C, Lee MC. Breast cancer in young women: special considerations in multidisciplinary care. *J Multidiscip Healthc*. 2014;7:419.
30. Cavalieri EL, Rogan EG. The etiology and prevention of breast cancer. *Drug Discovery Today Disease Mechanisms*. 2012;9(1–2):e55–69.
31. Coughlin SS, Smith SA. The impact of the natural, social, built, and policy environments on breast cancer. *J Environ Health Sci*. 2015;1(3):1.
32. Howell A, Anderson AS, Clarke RB, Duffy SW, Evans DG, Garcia-Closas M, Gescher AJ, Key TJ, Saxton JM, Harvie MN. Risk determination and prevention of breast cancer. *Breast Cancer Res*. 2014;16(5):1–19.
33. Podkowa N, Komasińska P, Roszak M, Gryczka R, Grodecka-Gazdecka S, Łacka K. Health behaviours among women diagnosed with breast tumours. *Polski Merkuriusz Lekarski: Organ Polskiego Towarzystwa Lekarskiego*. 2014;37(219):153–8.
34. Kruk J. Intensity of recreational physical activity in different life periods in relation to breast cancer among women in the region of Western Pomerania. *Contemp Oncol*. 2012;16(6):576.
35. Barlow WE, Lehman CD, Zheng Y, Ballard-Barbash R, Yankaskas BC, Cutter GR, Carney PA, Geller BM, Rosenberg R, Kerlikowske K. Performance of diagnostic mammography for women with signs or symptoms of breast cancer. *J Natl Cancer Inst*. 2002;94(15):1151–9.
36. Wang L. Early diagnosis of breast cancer. *Sensors*. 2017;17(7):1572.
37. Alnoqaidan EA, Rubui RM, Alshehri AS, Aljarba ZFA, Alhelali NNG, Aldablan AS, Albalawi AZ, Alamri AHA, Shaa-ban AS, Albaghdadi AT. An overview on diagnosis and management approach of breast cancer.
38. Tinoco G, Warsch S, Glück S, Avancha K, Montero AJ. Treating breast cancer in the 21st century: emerging biological therapies. *J Breast Cancer*. 2013;4(2):117–32.
39. Matsen CB, Neumayer LA. Breast cancer: a review for the general surgeon. *JAMA Surg*. 2013;148(10):971–80.
40. Dai X, Li T, Bai Z, Yang Y, Liu X, Zhan J, Shi B. Breast cancer intrinsic subtype classification, clinical use and future trends. *Am J Cancer Res*. 2015;5(10):2929.
41. Emens LA. Breast cancer immunotherapy: facts and hopes. *Clin Cancer Res*. 2018;24(3):511–20.
42. Infante-Duarte C, Horton HF, Byrne MC, Kamradt T. Microbial lipopeptides induce the production of IL-17 in Th cells. *J Immunol*. 2000;165(11):6107–15.
43. Bettelli E, Korn T, Kuchroo VK. Th17: the third member of the effector T cell trilogy. *Curr Opin Immunol*. 2007;19(6):652–7.
44. Laprevotte E, Bastid J, Cochaud S, Giustiniani J, Philippe M, Frewer KA, Sanders AJ, Jiang WG, Bensusan A, Alberici G. Interleukin-17B promotes chemoresistance of breast tumors through ERK1/2 anti-apoptotic pathway. In: AACR; 2015.
45. Khader SA, Gaffen SL, Kolls JK. Th17 cells at the crossroads of innate and adaptive immunity against infectious diseases at the mucosa. *Mucosal Immunol*. 2009;2(5):403–11.
46. Fossiez F, Djossou O, Chomarat P, Flores-Romo L, Ait-Yahia S, Maat C, Pin J-J, Garrone P, Garcia E, Saeland S. T cell interleukin-17 induces stromal cells to produce proinflammatory and hematopoietic cytokines. *J Exp Med*. 1996;183(6):2593–603.
47. Pelletier M, Maggi L, Micheletti A, Lazzeri E, Tamassia N, Costantini C, Cosmi L, Lunardi C, Annunziato F, Romagnani S. Evidence for a cross-talk between human neutrophils and Th17 cells. *Blood*. 2010;115(2):335–43.
48. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, Oliver P, Huang W, Zhang P, Zhang J. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. *J Exp Med*. 2001;194(4):519–28.
49. Liang SC, Tan X-Y, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, Fouser LA. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med*. 2006;203(10):2271–9.
50. Martin-Orozco N, Muranski P, Chung Y, Yang XO, Yamazaki T, Lu S, Hwu P, Restifo NP, Overwijk WW, Dong C. T helper 17 cells promote cytotoxic T cell activation in tumor immunity. *Immunity*. 2009;31(5):787–98.
51. Mitsdoerffer M, Lee Y, Jäger A, Kim H-J, Korn T, Kolls JK, Cantor H, Bettelli E, Kuchroo VK. Proinflammatory T helper type 17 cells are effective B-cell helpers. *Proc Natl Acad Sci*. 2010;107(32):14292–7.
52. Munegowda MA, Deng Y, Mulligan SJ, Xiang J. Th17 and Th17-stimulated CD8+ T cells play a distinct role in Th17-induced preventive and therapeutic antitumor immunity. *Cancer Immunol Immunother*. 2011;60(10):1473.
53. Ronchi F, Basso C, Preite S, Reboldi A, Baumjohann D, Perlini L, Lanzavecchia A, Sallusto F. Experimental priming of encephalitogenic Th1/Th17 cells requires pertussis toxin-driven IL-1 β production by myeloid cells. *Nat Commun*. 2016;7(1):1–11.
54. Pang B, Zhen Y, Hu C, Ma Z, Lin S, Yi H. Myeloid-derived suppressor cells shift Th17/Treg ratio and promote systemic lupus erythematosus progression through arginase-1/miR-322-5p/TGF- β pathway. *Clin Sci*. 2020;134(16):2209–22.
55. Shahid A, Bharadwaj M. The connection between the Th17 cell related cytokines and cancer stem cells in cancer: Novel therapeutic targets. *Immunol Lett*. 2019;213:9–20.
56. Chen WC, Lai YH, Chen HY, Guo HR, Su IJ, Chen HH. Interleukin-17-producing cell infiltration in the breast cancer tumour microenvironment is a poor prognostic factor. *Histopathology*. 2013;63(2):225–33.
57. Fujii Y, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, Bamba T, Fujiyama Y. Increased expression of interleukin 17 in inflammatory bowel disease. *Gut*. 2003;52(1):65–70.

58. Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, Sudo K, Iwakura Y. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *J Immunol*. 2006;177(1):566–73.
59. Nakae S, Nambu A, Sudo K, Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J Immunol*. 2003;171(11):6173–7.
60. Tzartos JS, Friese MA, Craner MJ, Palace J, Newcombe J, Esiri MM, Fugger L. Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol*. 2008;172(1):146–55.
61. Alizadeh D, Katsanis E, Larmonier N. The multifaceted role of Th17 lymphocytes and their associated cytokines in cancer. *Clin Dev Immunol*. 2013;2013:1.
62. Kurebayashi Y, Nagai S, Ikejiri A, Koyasu S. Recent advances in understanding the molecular mechanisms of the development and function of Th17 cells. *Genes Cells*. 2013;18(4):247–65.
63. Yang X-P, Ghoreschi K, Steward-Tharp SM, Rodriguez-Canales J, Zhu J, Grainger JR, Hirahara K, Sun H-W, Wei L, Vahedi G. Opposing regulation of the locus encoding IL-17 through direct, reciprocal actions of STAT3 and STAT5. *Nat Immunol*. 2011;12(3):247–54.
64. Chen Z, Tato CM, Muul L, Laurence A, O'Shea JJ. Distinct regulation of interleukin-17 in human T helper lymphocytes. *Arthritis Rheum*. 2007;56(9):2936–46.
65. Gorelik L, Constant S, Flavell RA. Mechanism of transforming growth factor β -induced inhibition of T helper type 1 differentiation. *J Exp Med*. 2002;195(11):1499–505.
66. Gorelik L, Fields PE, Flavell RA. Cutting edge: TGF- β inhibits Th type 2 development through inhibition of GATA-3 expression. *J Immunol*. 2000;165(9):4773–7.
67. Hirota K, Martin B, Veldhoen M. Development, regulation and functional capacities of Th17 cells. In: Springer, 2010;3–16.
68. Schumann J, Muller U, Blessing M. TGF- β signaling in T cells is not essential for Th17 cell development in the mouse. *J Biol Regul Homeost Agents*. 2012;26(3):357–66.
69. Sonderegger I, Kisielow J, Meier R, King C, Kopf M. IL-21 and IL-21R are not required for development of Th17 cells and autoimmunity in vivo. *Eur J Immunol*. 2008;38(7):1833–8.
70. Korn T, Bettelli E, Gao W, Awasthi A, Jäger A, Strom TB, Oukka M, Kuchroo VK. IL-21 initiates an alternative pathway to induce proinflammatory TH17 cells. *Nature*. 2007;448(7152):484–7.
71. Leonard WJ, Spolski R. Interleukin-21: a modulator of lymphoid proliferation, apoptosis and differentiation. *Nat Rev Immunol*. 2005;5(9):688–98.
72. Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, Schluns K, Tian Q, Watowich SS, Jetten AM. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. *Nature*. 2007;448(7152):480–3.
73. Zhou L, Ivanov II, Spolski R, Min R, Shenderov K, Egawa T, Levy DE, Leonard WJ, Littman DR. IL-6 programs TH-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. *Nat Immunol*. 2007;8(9):967–74.
74. Karpisheh V, Afjadi JF, Afjadi MN, Haeri MS, Sough TSA, Asl SH, Edalati M, Atyabi F, Masjedi A, Hajizadeh F. Inhibition of HIF-1 α /EP4 axis by hyaluronate-trimethyl chitosan-SPION nanoparticles markedly suppresses the growth and development of cancer cells. *Int J Biol Macromol*. 2021;167:1006–19.
75. Rostamzadeh D, Razavi SR, Esmaeili S, Dolati S, Ahmahi M, Sadreddini S, Jadidi-Niaragh F, Yousefi M. Application of nanoparticle technology in the treatment of systemic lupus erythematosus. *Biomed Pharmacother*. 2016;83:1154–63.
76. Durant L, Watford WT, Ramos HL, Laurence A, Vahedi G, Wei L, Takahashi H, Sun HW, Kanno Y, Powrie F, et al. Diverse targets of the transcription factor STAT3 contribute to T cell pathogenicity and homeostasis. *Immunity*. 2010;32(5):605–15.
77. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, Dong C. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *J Biol Chem*. 2007;282(13):9358–63.
78. Leonardi C, Matheson R, Zachariae C, Cameron G, Li L, Edson-Heredia E, Braun D, Banerjee S. Anti-interleukin-17 monoclonal antibody ixekizumab in chronic plaque psoriasis. *N Engl J Med*. 2012;366(13):1190–9.
79. Patel DD, Lee DM, Kolbinger F, Antoni C. Effect of IL-17A blockade with secukinumab in autoimmune diseases. *Ann Rheumatic Dis*. 2013;72(suppl 2):iii116–23.
80. Ridker PM, Thuren T, Zalewski A, Libby P. Interleukin-1 β inhibition and the prevention of recurrent cardiovascular events: rationale and design of the Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS). *Am Heart J*. 2011;162(4):597–605.
81. Ritchlin C, Rahman P, Kavanaugh A, McInnes IB, Puig L, Li S, Wang Y, Shen Y-K, Doyle MK, Mendelsohn AM. Efficacy and safety of the anti-IL-12/23 p40 monoclonal antibody, ustekinumab, in patients with active psoriatic arthritis despite conventional non-biological and biological anti-tumour necrosis factor therapy: 6-month and 1-year results of the phase 3, multicentre, double-blind, placebo-controlled, randomised PSUMMIT 2 trial. *Ann Rheum Dis*. 2014;73(6):990–9.
82. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, Weaver CT. Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol*. 2005;6(11):1123–32.
83. Gocke AR, Cravens PD, Ben L-H, Hussain RZ, Northrop SC, Racke MK, Lovett-Racke AE. T-bet regulates the fate of Th1 and Th17 lymphocytes in autoimmunity. *J Immunol*. 2007;178(3):1341–8.
84. Lighvani AA, Frucht DM, Jankovic D, Yamane H, Aliberti J, Hissong BD, Nguyen BV, Gadina M, Sher A, Paul WE. T-bet is rapidly induced by interferon- γ in lymphoid and myeloid cells. *Proc Natl Acad Sci*. 2001;98(26):15137–42.
85. Batten M, Li J, Yi S, Klijavin NM, Danilenko DM, Lucas S, Lee J, de Sauvage FJ, Ghilardi N. Interleukin 27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. *Nat Immunol*. 2006;7(9):929–36.
86. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang Y-H, Wang Y, Hood L, Zhu Z, Tian Q. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol*. 2005;6(11):1133–41.
87. Kleinschek MA, Owyang AM, Joyce-Shaikh B, Langrish CL, Chen Y, Gorman DM, Blumenschein WM, McClanahan T, Brombacher F, Hurst SD. IL-25 regulates Th17 function in autoimmune inflammation. *J Exp Med*. 2007;204(1):161–70.
88. Kryczek I, Wei S, Vatan L, Escara-Wilke J, Szeliga W, Keller ET, Zou W. Cutting edge: opposite effects of IL-1 and IL-2 on the regulation of IL-17+ T cell pool IL-1 subverts IL-2-mediated suppression. *J Immunol*. 2007;179(3):1423–6.
89. Kryczek I, Wei S, Zou L, Altuwajri S, Szeliga W, Kolls J, Chang A, Zou W. Cutting edge: Th17 and regulatory T cell dynamics and the regulation by IL-2 in the tumor microenvironment. *J Immunol*. 2007;178(11):6730–3.
90. Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, Blank RB, Meylan F, Siegel R, Hennighausen L. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. *Immunity*. 2007;26(3):371–81.
91. Moisan J, Grenningloh R, Bettelli E, Oukka M, Ho IC. Ets-1 is a negative regulator of Th17 differentiation. *J Exp Med*. 2007;204(12):2825–35.
92. Baharlou R, Atashzar MR, Vasmehjani AA, Rahimi E, Khoshmirsafa M, Seif F, Mahdiyari M. Reduced levels of T-helper 17-associated cytokines in the serum of patients with breast cancer: indicators for following the course of disease. *Central-Eur J Immunol*. 2016;41(1):78.
93. Du J-W, Xu K-Y, Fang L-Y, Qi X-L. Interleukin-17, produced by lymphocytes, promotes tumor growth and angiogenesis in a mouse model of breast cancer. *Mol Med Rep*. 2012;6(5):1099–102.
94. Kim K, Kim G, Kim J-Y, Yun HJ, Lim S-C, Choi HS. Interleukin-22 promotes epithelial cell transformation and breast tumorigenesis via MAP3K8 activation. *Carcinogenesis*. 2014;35(6):1352–61.
95. Wang J, Cai D, Ma B, Wu G, Wu J. Skewing the balance of regulatory T-cells and T-helper 17 cells in breast cancer patients. *J Int Med Res*. 2011;39(3):691–701.
96. Benevides L, Cardoso CRB, Tiezzi DG, Marana HRC, Andrade JM, Silva JS. Enrichment of regulatory T cells in invasive breast tumor correlates with the upregulation of IL-17A expression and invasiveness of the tumor. *Eur J Immunol*. 2013;43(6):1518–28.
97. Thibaudin M, Chaix M, Boidot R, Végran F, Derangère V, Limagne E, Berger H, Ladoire S, Apetoh L, Ghiringhelli F. Human ectonucleotidase-expressing CD25high Th17 cells accumulate in breast cancer tumors and exert immunosuppressive functions. *Oncoimmunology*. 2016;5(1):e1055444.

98. Ma K, Yang L, Shen R, Kong B, Chen W, Liang J, Tang G, Zhang B. Th17 cells regulate the production of CXCL1 in breast cancer. *Int Immunopharmacol*. 2018;56:320–9.
99. Zumwalde NA, Haag JD, Gould MN, Gumperz JE. Mucosal associated invariant T cells from human breast ducts mediate a Th17-skewed response to bacterially exposed breast carcinoma cells. *Breast Cancer Res*. 2018;20(1):1–14.
100. Avalos-Navarro G, Muñoz-Valle JF, Daneri-Navarro A, Quintero-Ramos A, Franco-Topete RA, de Jesus M-M, Ocegüera-Villanueva A, Bautista-Herrera LA, Topete-Camacho A, Del Toro-Arreola A. Circulating soluble levels of MIF in women with breast cancer in the molecular subtypes: relationship with Th17 cytokine profile. *Clin Exp Med*. 2019;19(3):385–91.
101. Coffelt SB, Kersten K, Doornebal CW, Weiden J, Vrijland K, Hau C-S, Versteegen NJM, Ciampricotti M, Hawinkels LJAC, Jonkers J. IL-17-producing $\gamma\delta$ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature*. 2015;522(7556):345–8.
102. Kim G, Khanal P, Lim S-C, Yun HJ, Ahn S-G, Ki SH, Choi HS. Interleukin-17 induces AP-1 activity and cellular transformation via upregulation of tumor progression locus 2 activity. *Carcinogenesis*. 2013;34(2):341–50.
103. Huang SC, Wei PC, Hwang-Verslues WW, Kuo WH, Jeng YM, Hu CM, Shew JY, Huang CS, Chang KJ, Lee EYH. TGF- β 1 secreted by Tregs in lymph nodes promotes breast cancer malignancy via up-regulation of IL-17RB. *EMBO Mol Med*. 2017;9(12):1660–80.
104. Benatar T, Cao MY, Lee Y, Lightfoot J, Feng N, Gu X, Lee V, Jin H, Wang M, Wright JA. IL-17E, a proinflammatory cytokine, has antitumor efficacy against several tumor types in vivo. *Cancer Immunol Immunother*. 2010;59(6):805–17.
105. Jiang Z, Chen J, Du X, Cheng H, Wang X, Dong C. IL-25 blockade inhibits metastasis in breast cancer. *Protein Cell*. 2017;8(3):191–201.
106. Fauchoux L, Grandclaudon M, Perrot-Dockès M, Sirven P, Berger F, Hamy AS, Fourchette V, Vincent-Salomon A, Mechta-Grigoriou F, Reyat F. A multivariate Th17 metagene for prognostic stratification in T cell non-inflamed triple negative breast cancer. *Oncoimmunology*. 2019;8:e1624130.
107. Ma M, Huang W, Kong D. IL-17 inhibits the accumulation of myeloid-derived suppressor cells in breast cancer via activating STAT3. *Int Immunopharmacol*. 2018;59:148–56.
108. Pawlik A, Anisiewicz A, Filip-Psurska B, Klopotowska D, Maciejewska M, Mazur A, Wietrzyk J. Divergent effect of tacalcitol (PRI-2191) on Th17 cells in 4T1 tumor bearing young and old ovariectomized mice. *Aging Dis*. 2020;11(2):241.
109. Yu L-Y, Tang J, Zhang C-M, Zeng W-J, Yan H, Li M-P, Chen X-P. New immunotherapy strategies in breast cancer. *Int J Environ Res Public Health*. 2017;14(1):68.
110. Asadzadeh Z, Mohammadi H, Safarzadeh E, Hemmatzadeh M, Mahdian-Shakib A, Jadidi-Niaragh F, Azizi G, Baradaran B. The paradox of Th17 cell functions in tumor immunity. *Cell Immunol*. 2017;322:15–25.
111. Joshi N, Hajizadeh F, Dezfouli EA, Zekiy AO, Afjadi MN, Mousavi SM, Hojjat-Farsangi M, Karpisheh V, Mahmoodpoor A, Hassannia H. Silencing STAT3 enhances sensitivity of cancer cells to doxorubicin and inhibits tumor progression. *Life Sci*. 2021;275:119369.
112. Amedei A, Munari F, Della Bella C, Niccolai E, Benaglio M, Bencini L, Cianchi F, Farsi M, Emmi G, Zanotti G. Helicobacter pylori secreted peptidyl prolyl cis, trans-isomerase drives Th17 inflammation in gastric adenocarcinoma. *Intern Emerg Med*. 2014;9(3):303–9.
113. Segura E, Touzot M, Bohineust A, Cappuccio A, Chiocchia G, Hosmalin A, Dalod M, Soumelis V, Amigorena S. Human inflammatory dendritic cells induce Th17 cell differentiation. *Immunity*. 2013;38(2):336–48.

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